



# 3D Imaging using Coherent Synchrotron Radiation



P. Cloetens

W. Ludwig<sup>1</sup>, O. Hignette, R. Mokso,  
E. Boller, X. Thibault, E. Pagot

*ESRF, Grenoble*

<sup>1</sup> *GEMPPM, INSA, Lyon*

## X-ray Imaging Group

ID17 Medical Beamline

ID19 Micro-Tomography

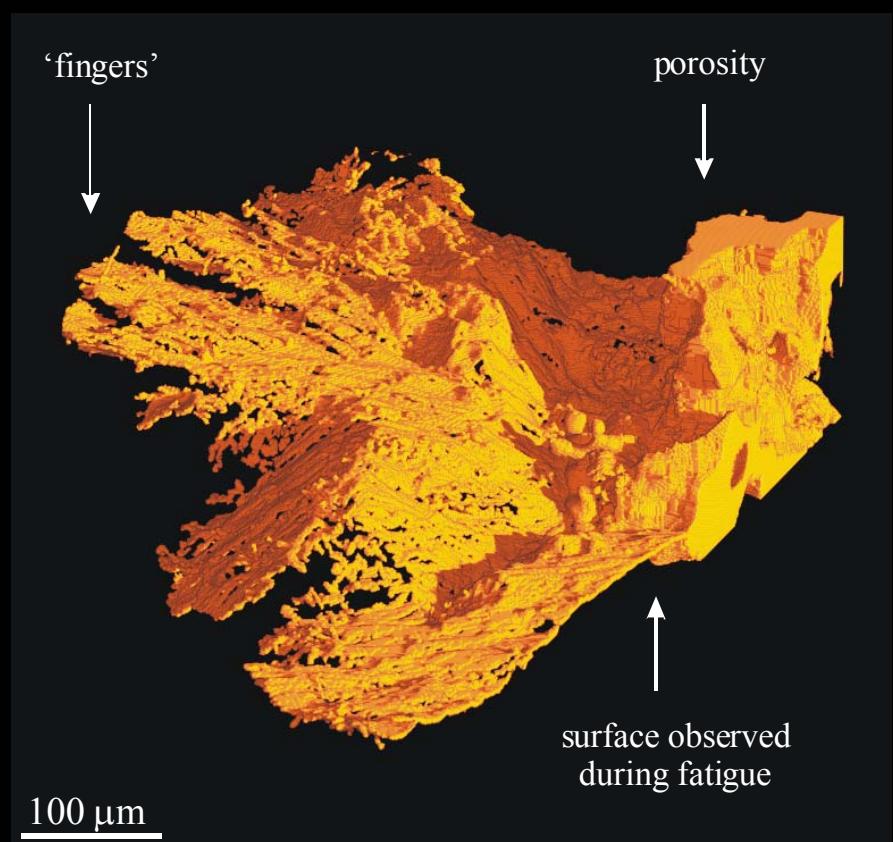
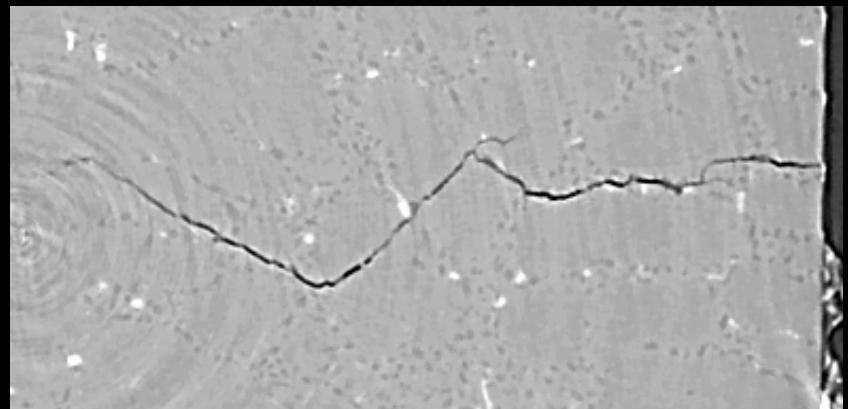
ID21 Microscopy / Micro-analysis

ID22/ID18F Micro-Fluorescence

150 of 900 ESRF proposals

# 2D vs 3D Imaging

- Microscopist:  
knows the answer to his  
problem  
looks for the image that  
confirms his idea
- Tomographist  
tries to get the whole picture



# Microtomography using synchrotron radiation

High flux, monochromatic, parallel, coherent beam:

High spatial resolution (~ micron)

Quantitative and ‘exact’ tomography

Fast Tomography (*in-situ*)

Phase Contrast Tomography

Trends:

Combined techniques: Fluo-Tomography

Multidisciplinary Technique

Nano-tomography

# 3-D Imaging: Better X-ray Sources



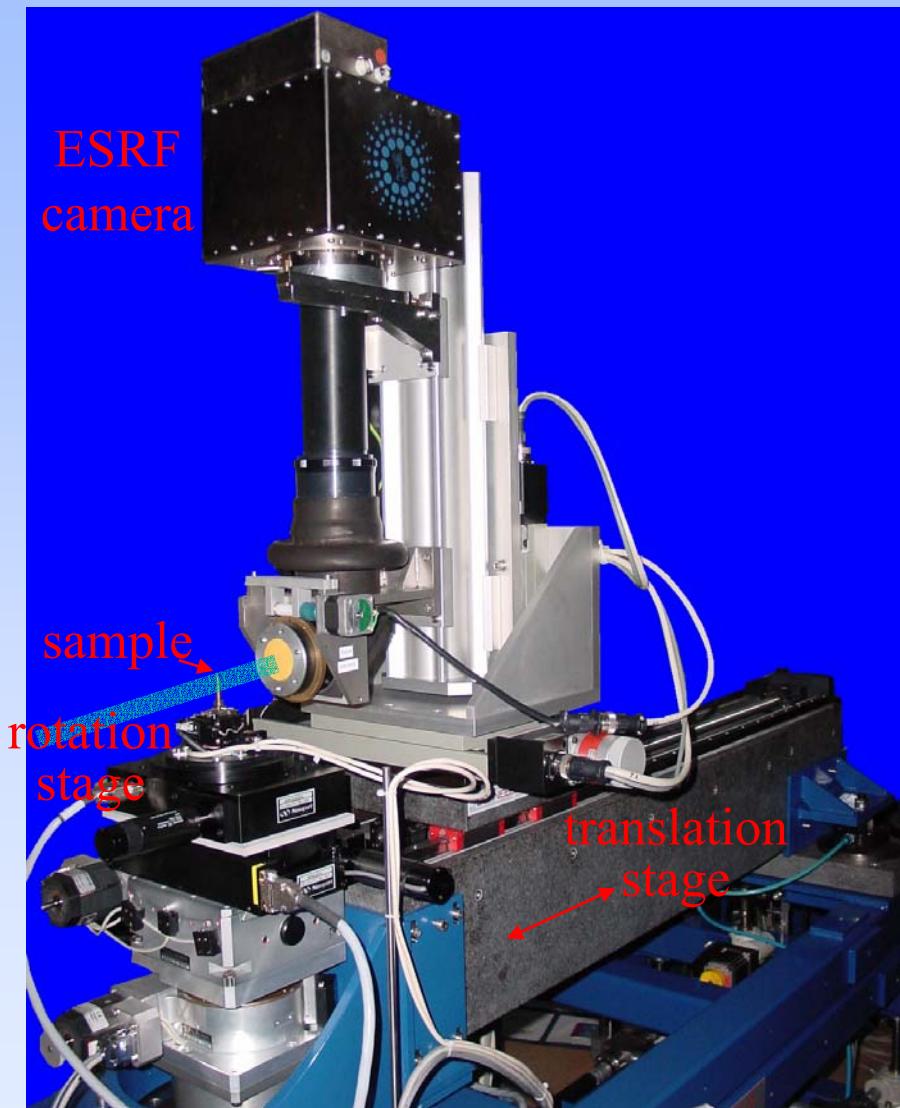
- High photon flux
- Monochromatic and parallel beam
- Partially Coherent beam

ESRF, ID19 Beamline:

source     $L = 145 \text{ m}$      $s_v \approx 25 \mu\text{m}$      $s_h \approx 125 \mu\text{m}$  }     $\alpha \approx 0.2 \mu\text{rad}$      $\rightarrow$      $l_{\text{coh}} = \lambda / 2\alpha \approx 250 \mu\text{m}$

and horizontally  $< 1 \mu\text{rad}$

# 3-D Imaging: Digital Detectors



Implemented on ID19 / ESRF

Dedicated microtomograph (P. Bernard)

Monochromator:

double Si crystal ( $\Delta\lambda/\lambda=10^{-4}$ )  
or multilayer ( $\Delta\lambda/\lambda=10^{-2}$ )

Sample stage

rotation stage (tomography)

Detector

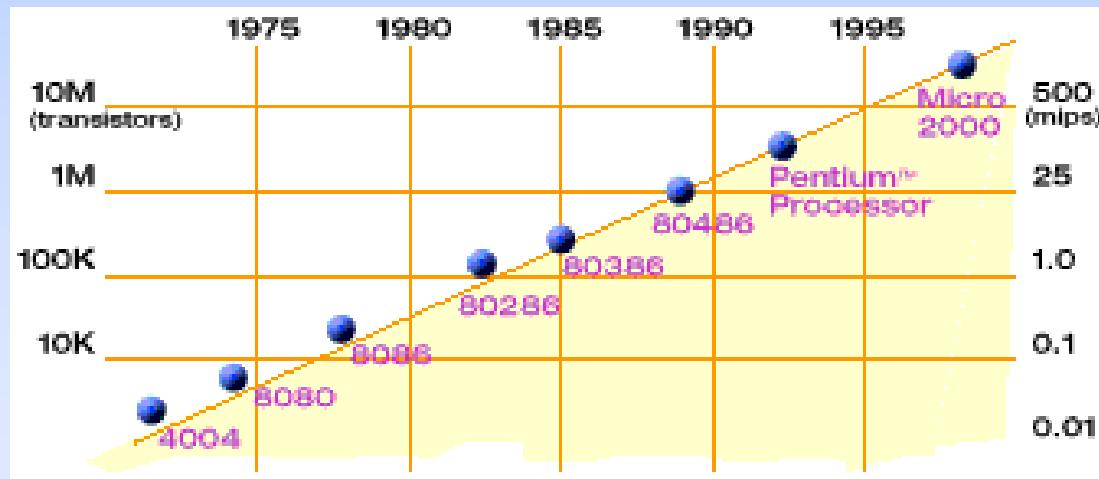
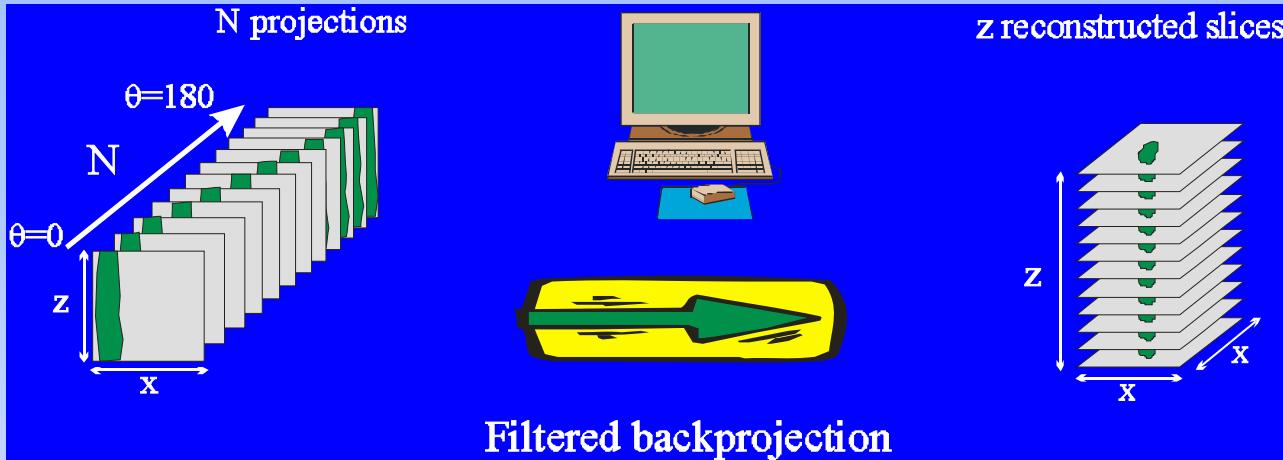
CCD-based (FReLoN)  
on translation (propagation)  
soon *FReLoN2* (frame transfer)

Scan time

$1024^2 * 900$  proj.: 5 minutes

$2048^2 * 1500$  proj.: 15 minutes

# 3-D Imaging: Faster Computers



Computing power grows exponentially

$1024^3$  volume reconstructed in 15 minutes

# In-situ Tomography

Follow 3D structure and composition as function of  
time (fast tomography)  
temperature (furnace, cryostat)  
strain (tensile stage)

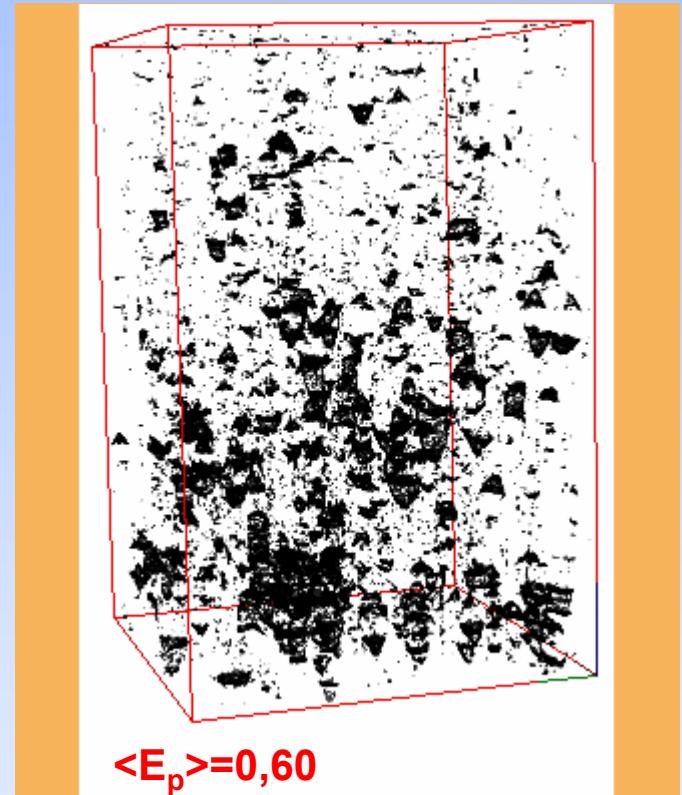
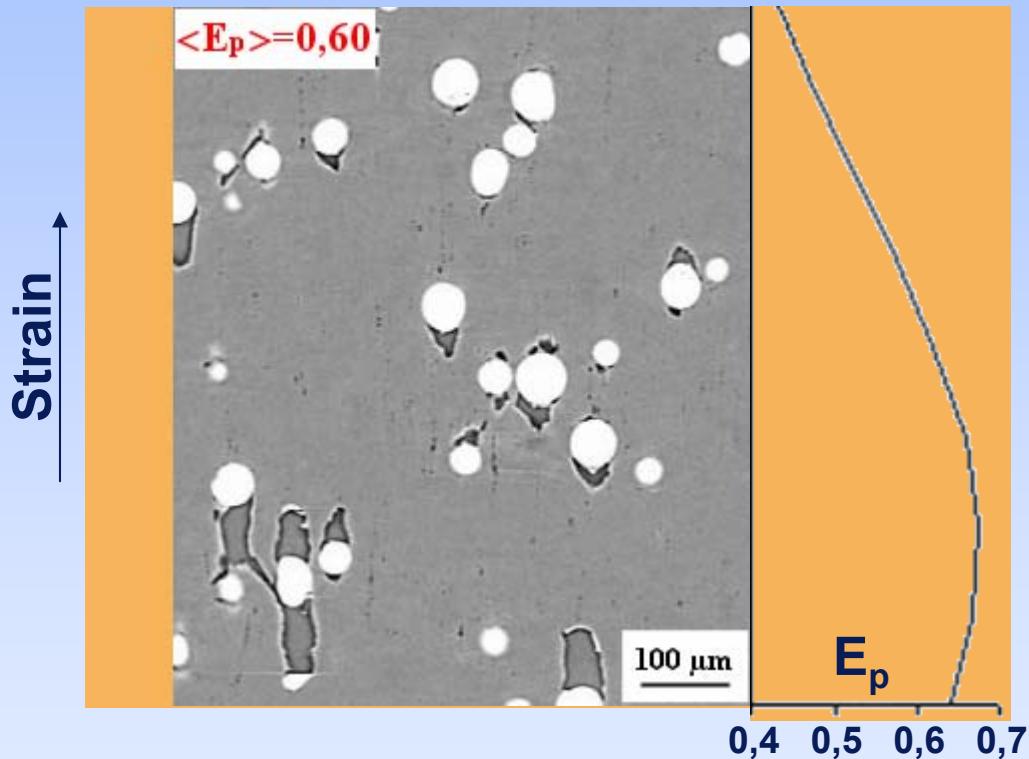
...

Sequential Tomography  
interrupt evolution between successive scans

Real-time Tomography  
scan time  $\ll$  evolution time  
High Energy Beamline

# Damage in composites

Example: soft matrix  
Al + 4%ZrO<sub>2</sub>SiO<sub>2</sub>



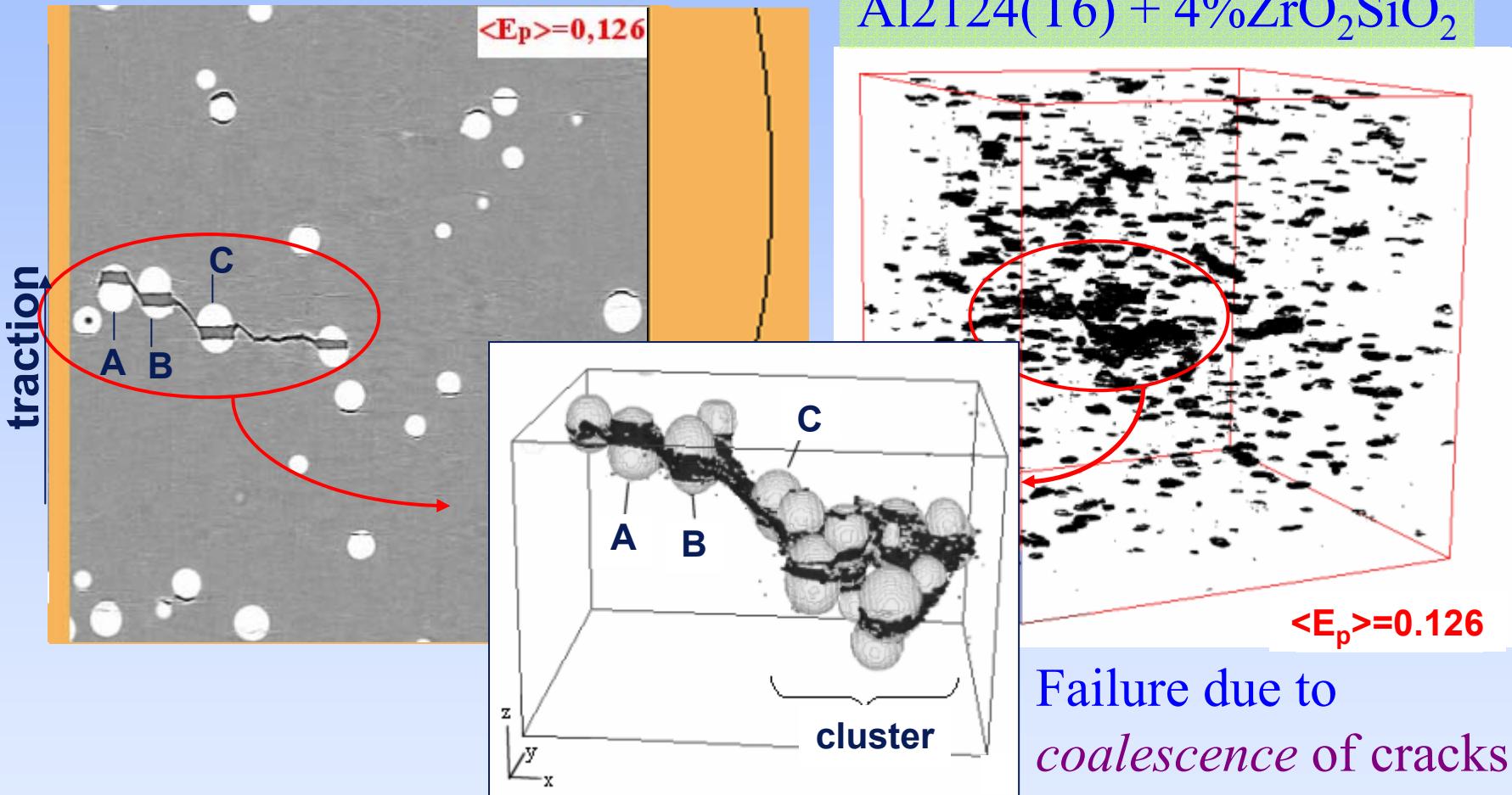
Preferred damage initiation: Matrix-particle *decohesion*

Conditions : in situ, E=30 keV abs/phase, 2  $\mu\text{m}$

L. Babout, E Maire (INSA-Lyon)

# Damage in composites

Example: hard matrix  
Al2124(T6) + 4%ZrO<sub>2</sub>SiO<sub>2</sub>



Failure due to  
*coalescence* of cracks  
and *local* configuration



Preferred damage initiation: particle rupture in mode I

Conditions : in situ, E=30 keV abs/phase, 2 μm

L. Babout, E Maire

# Damage in composites

Quantification of *damage* in the *bulk* of composite materials

Damage initiation  
mechanism  
criteria (normal stress / stored elastic energy)  
link to finite-element simulations

Damage growth: discrepancy with classical models

Failure: coalescence of cracks *under investigation*

Authors:

L. Babout, E. Maire, J.Y. Buffière  
*GEMPPM, INSA, Lyon*

# Why Phase Contrast?

- Dream 1: **Sensitivity**

Absorption contrast too low

high spatial resolution

$$n = 1 - \delta + i\beta$$

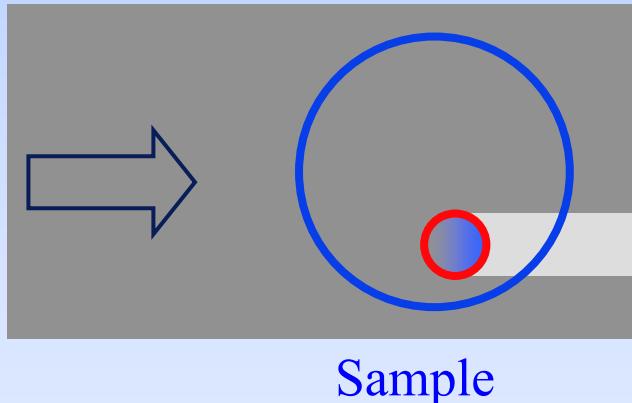
light materials

$$\delta \gg \beta$$

similar attenuation: C-C, Al-Si, Al-Al<sub>2</sub>O<sub>3</sub>

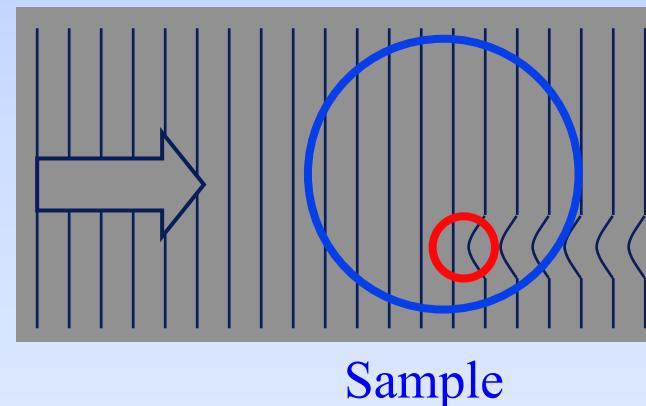
- Dream 2: **Zero Dose**

Absorption

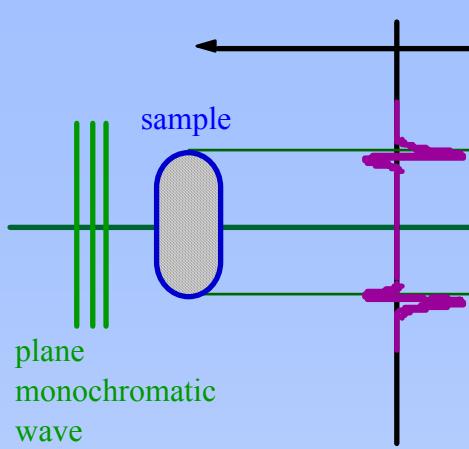


0.1% shrinkage

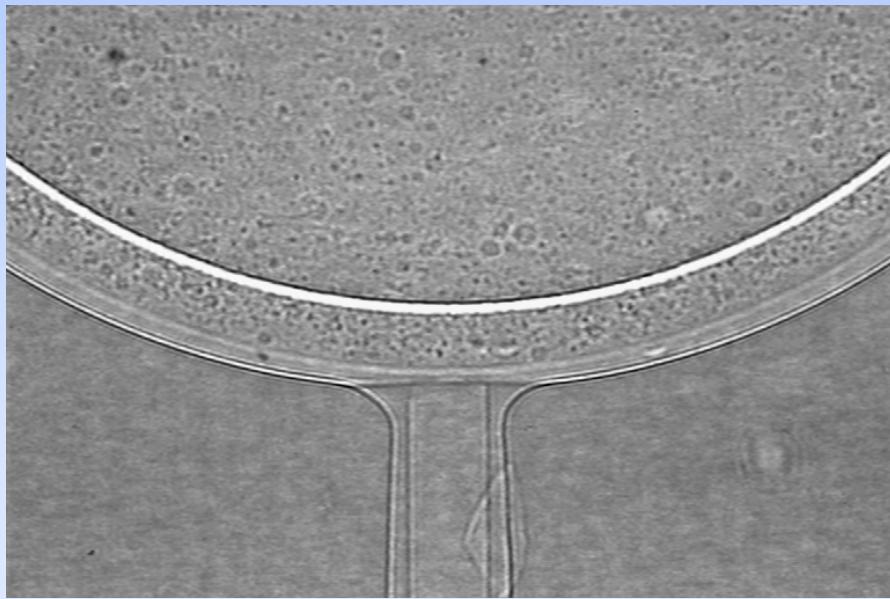
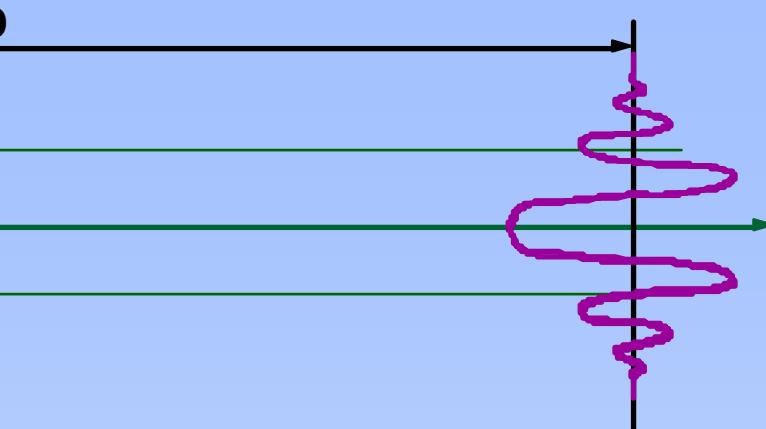
Phase



⇒ 2 voxels motion ( $N=2048$ )



edge detection

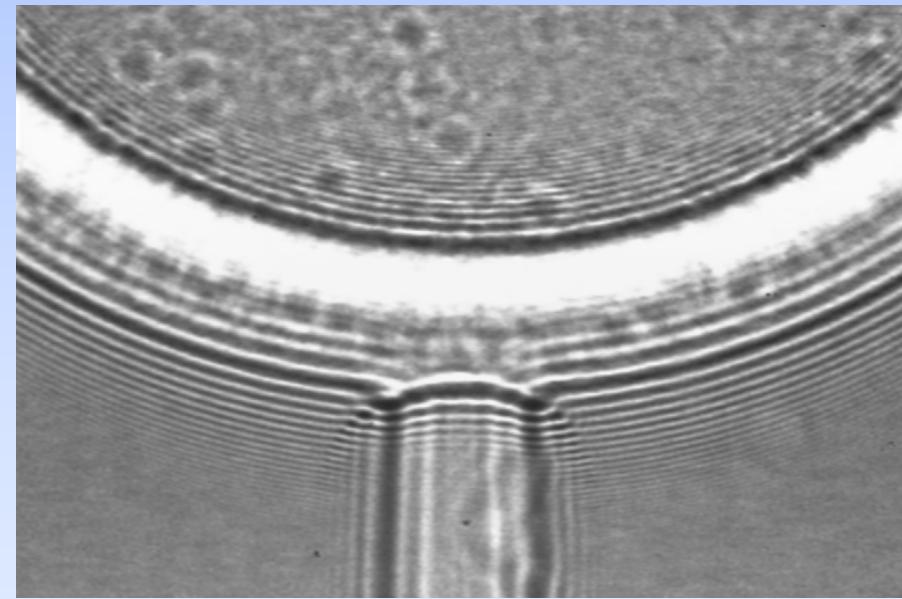


$$D = 15 \text{ cm}$$

each edge imaged independently  
no access to phase, only to border

$$\sqrt{\lambda D} \ll a$$

$$\begin{array}{c} \lambda = 0.7 \text{ \AA} \\ 50 \mu\text{m} \end{array}$$



$$D = 310 \text{ cm}$$

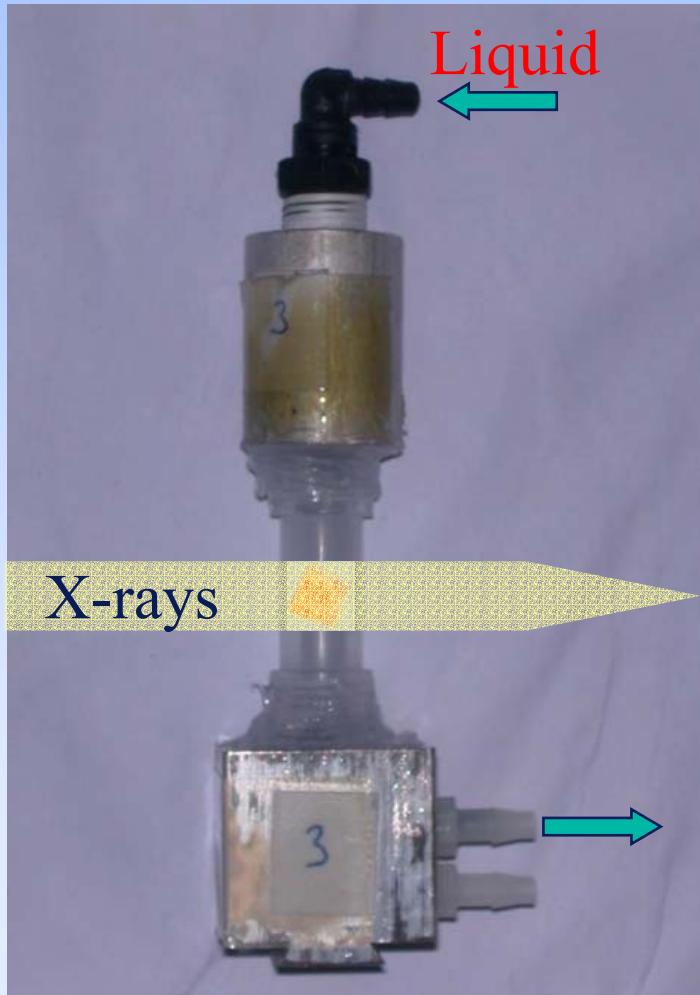
deformed *image* of whole object  
Phase retrieval needed

$$\sqrt{\lambda D} \approx a$$

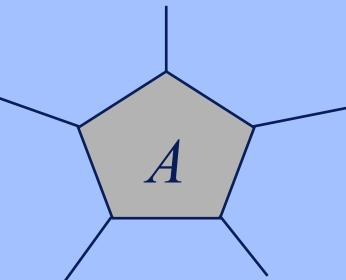
# Phase Contrast: Liquid Foams

Scientific Case:

Evolution (coarsening) of *liquid* foams in 3D



vs. in 2D



Bubble with  
surface  $A$  and  $n$  sides

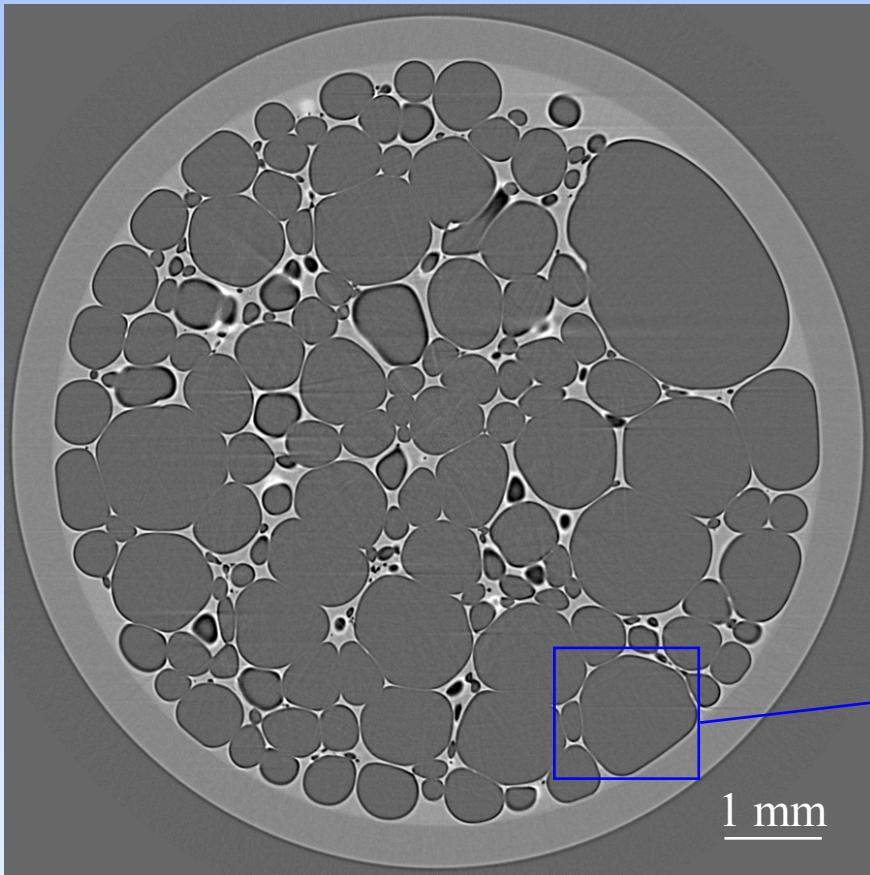
$$\frac{dA}{dt} \propto (n - 6)$$

Dry foam reaches *scaling state*  
average size grows with power law  
normalized distributions are constant

# Phase Contrast: Liquid Foams

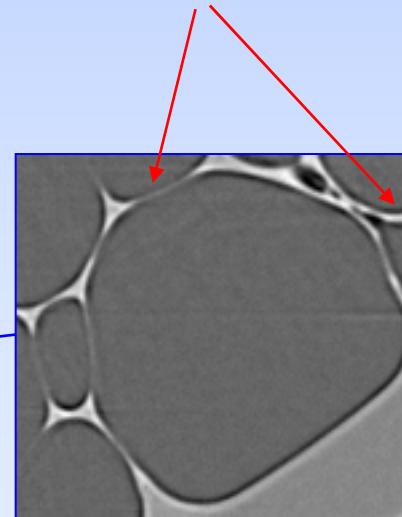
Scientific Case:

Evolution (coarsening) of liquid foams in 3D



Phase enhancement to visualise  
liquid films separating bubbles:  
Film thickness << voxel size

thin films



$E = 15 \text{ keV}$ , Sample-detector distance: 0.15 m

# Phase Contrast: Liquid Foams

Coarsening: pressure driven growth or disappearance of bubbles

QuickTime™ and a Sorenson Video 3 decompressor are needed to see this picture.

2 minutes/scan (2GB data)

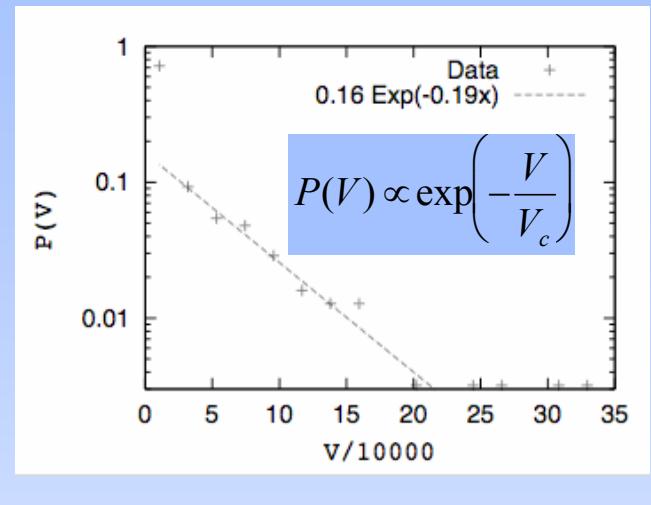
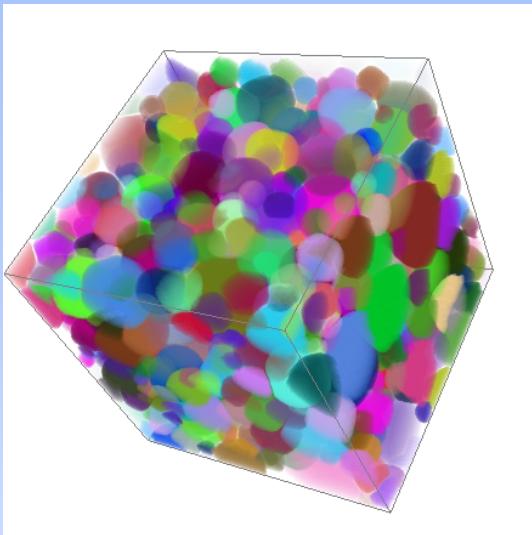
⇒ 3D Growth Law  
volume individual bubbles in time  
(cfr. grain growth, sintering)

# Liquid Foams

Model Foams: Dry Foam (liquid fraction  $\rightarrow 0$ )

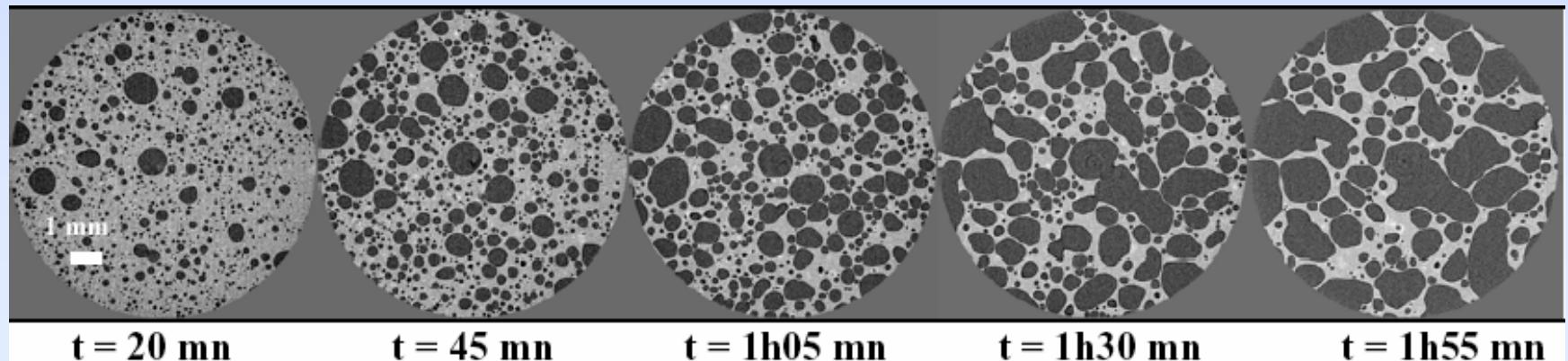
Individual growth rate

Scaling state ?



F. Graner (UJF), J. Lambert (Univ. Rennes)

Real foams: proteins, bread dough, baking

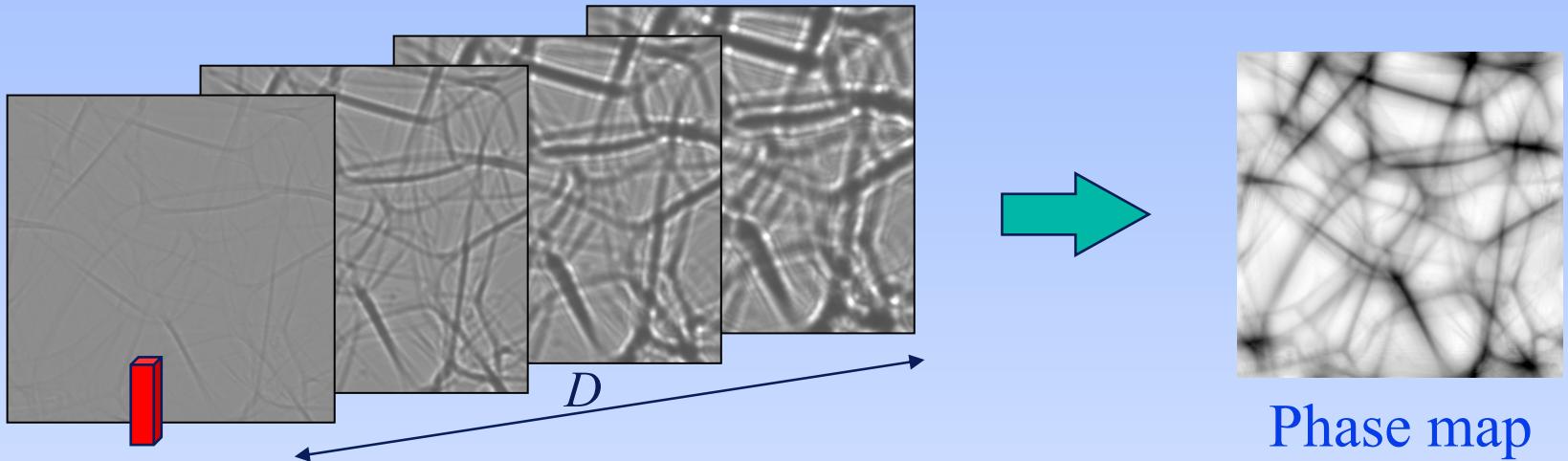


28 s / scan

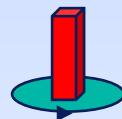
P. Babin (INPG), H. Chiron (INRA, Nantes)

# Holo-tomography

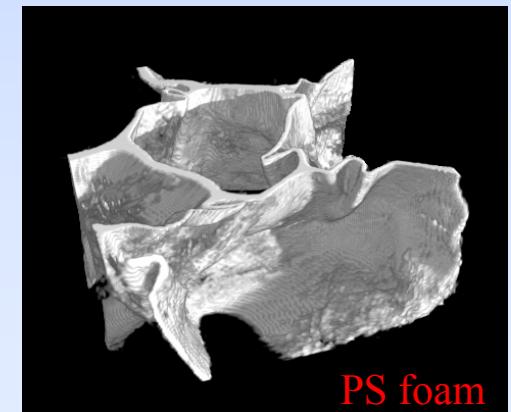
1) phase retrieval with images at different distances



2) tomography: repeated for  $\approx 1000$  angular positions



3D distribution of  $\delta$  or the electron-density  
improved resolution  
straightforward interpretation  
processing



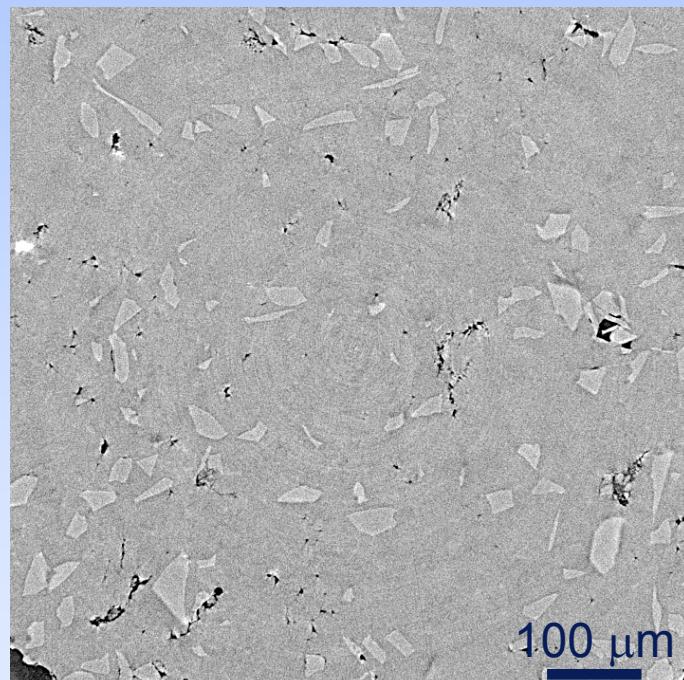
# Holotomography of composite materials

Al-SiC composite material

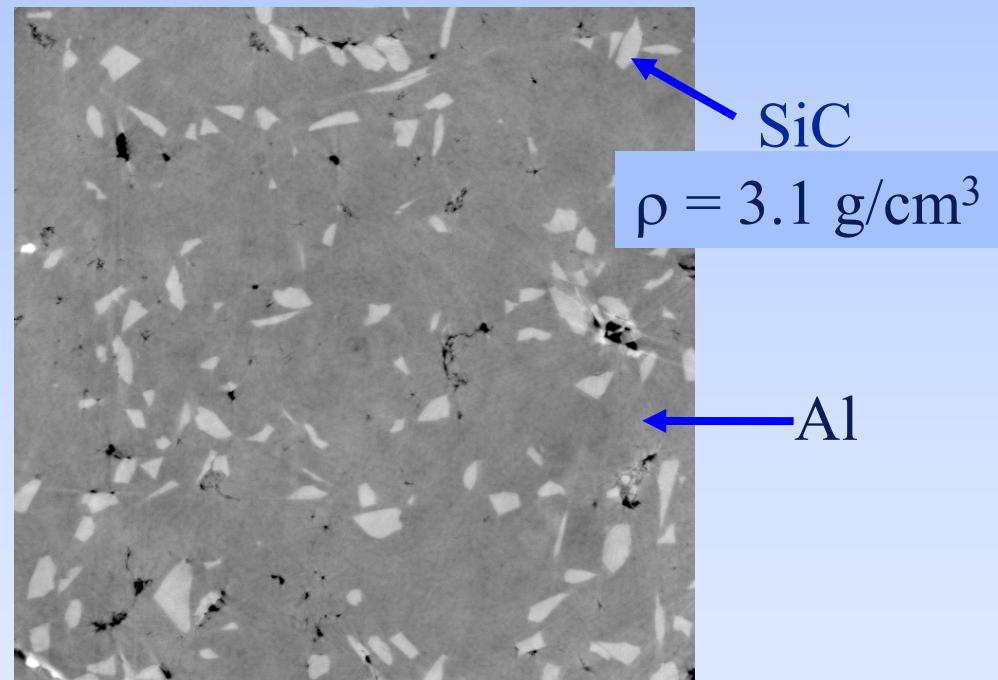
$$\Delta\delta = 3 \cdot 10^{-7} \text{ et } \Delta\beta = 3.3 \cdot 10^{-10} \text{ gain } \approx 900!$$

E = 20.5 keV

Phase retrieval with 3 distances



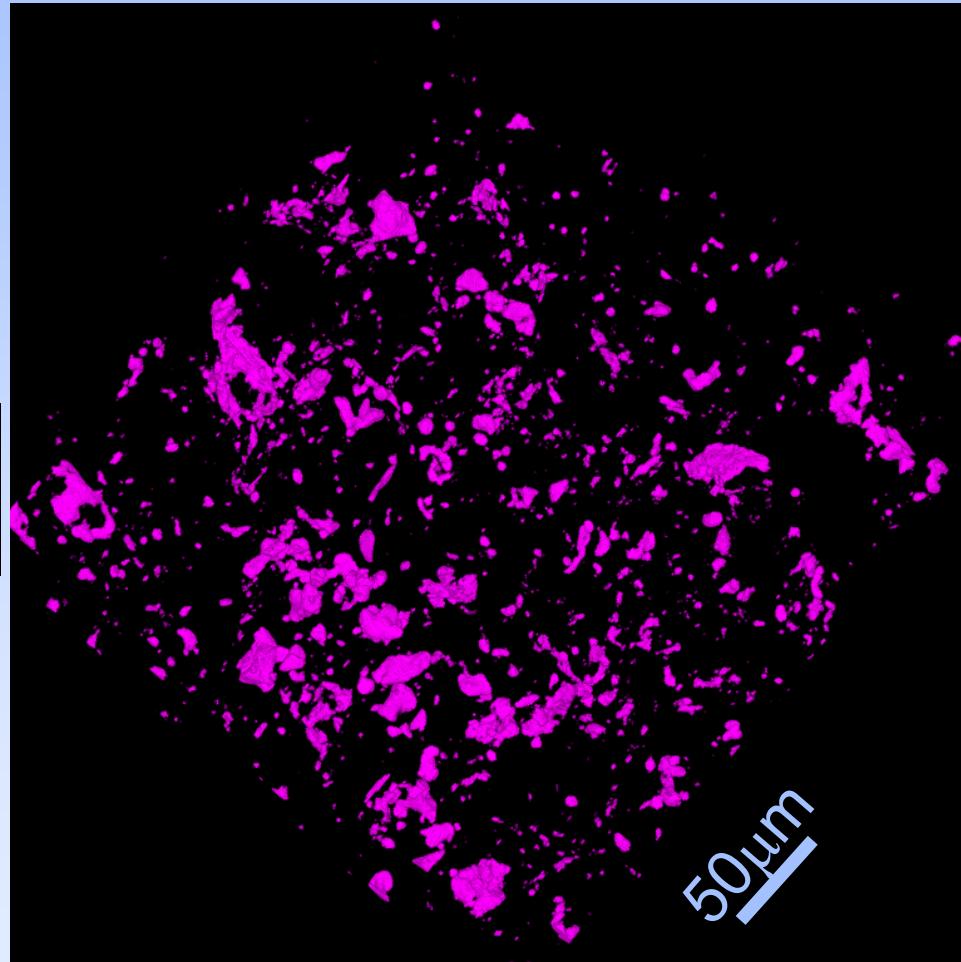
$\approx$  Absorption  
(D=7mm)



Holotomography  
density-map

# Composite materials

Al-SiC composite material

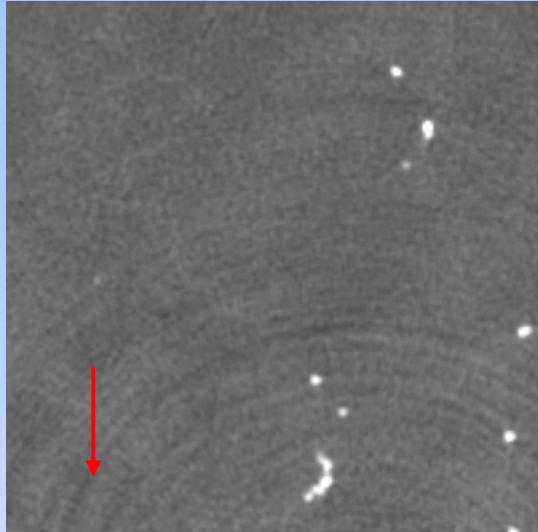


$E = 20.5 \text{ keV}$   
3 distances

Semi-solid  
Al / Al+Si

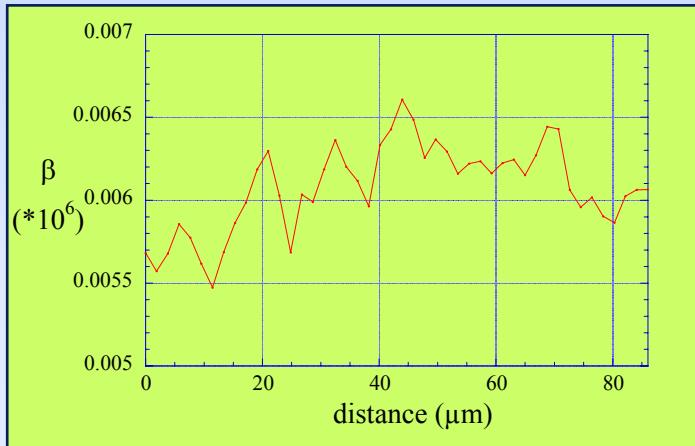
# Density Resolution

Absorption

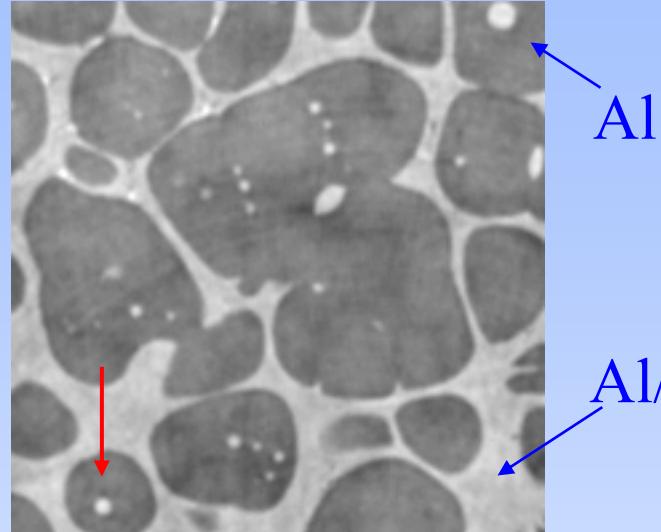


$\beta$ -map

E = 18 keV

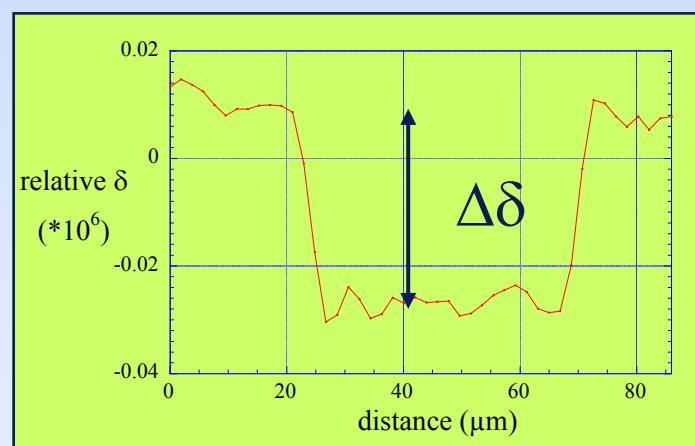


Phase contrast



50  $\mu\text{m}$

$\delta$ -map



$$\Delta\delta \approx 3.5 \cdot 10^{-8} \Rightarrow$$

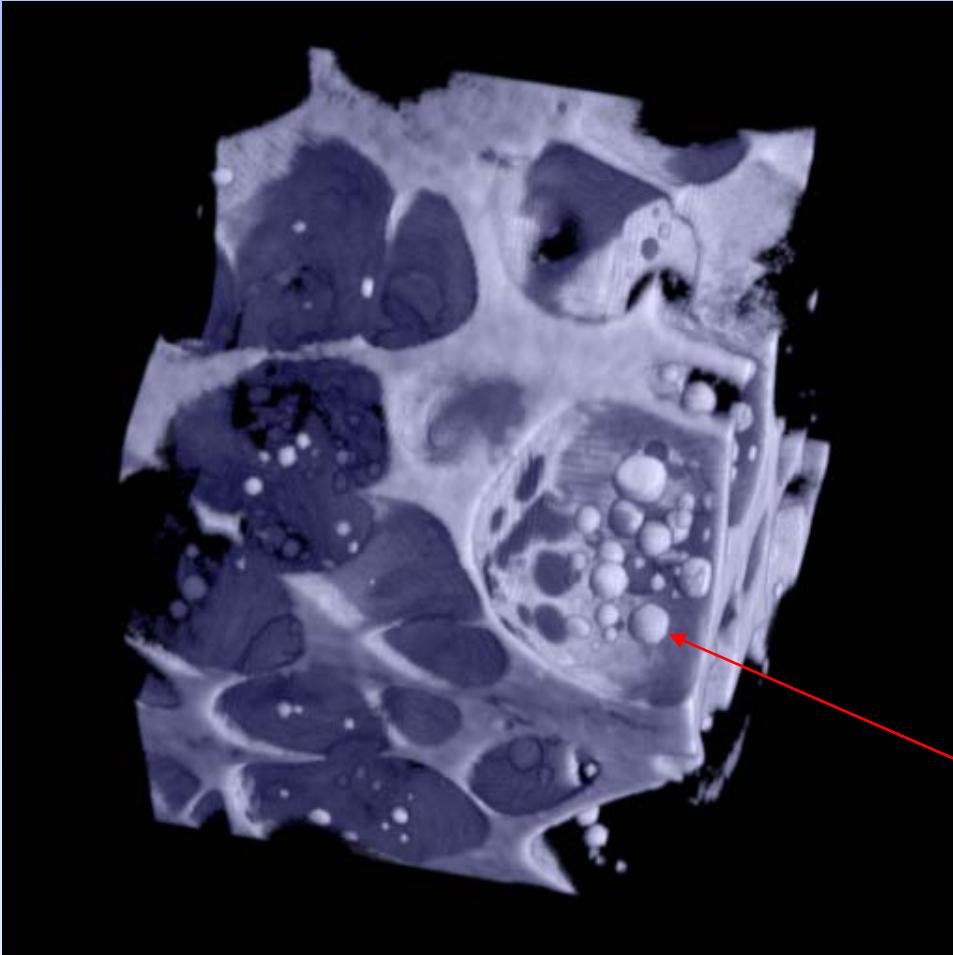
$$\Delta\rho \approx 0.05 \text{ g/cm}^3$$

15 % of Si

L. Salvo (GPM2, Grenoble)

# Holotomography of *semi-solids*

Volume Rendering (solid transparent)



Connectivity:

Liquid phase:

total

+ trapped liquid

Solid phase:

very strong

trapped liquid



input for permeability calculations

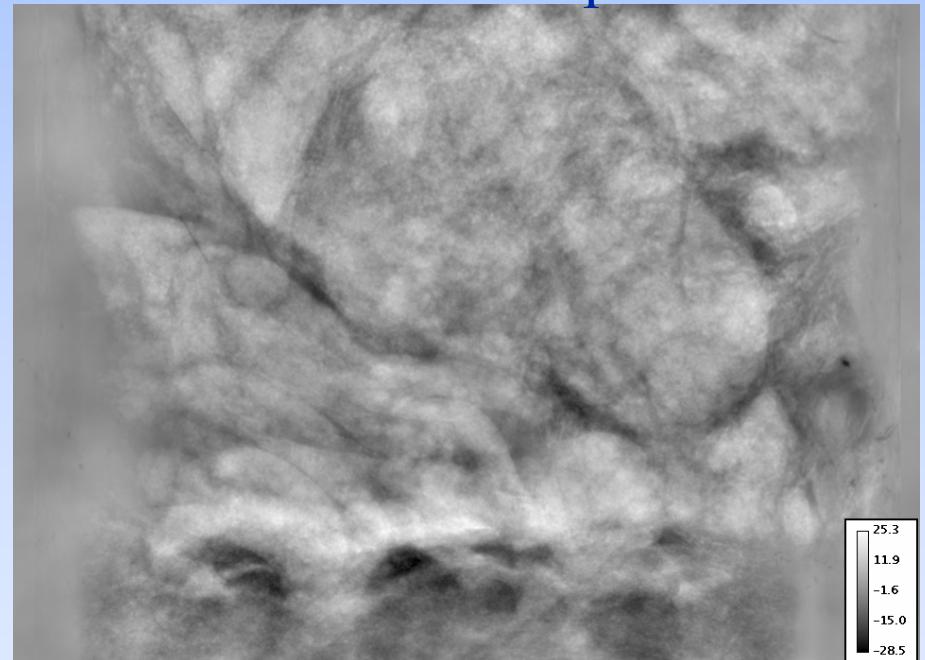
# Larger field of view

Breast Biopsy

$\varnothing \approx 15 \text{ mm}$  ; pixel size =  $7.5 \mu\text{m}$

$\Rightarrow$  Huge distances: 0.03, 1, 4.3 and 8.8 m !

Phase Map



QuickTime™ and a decompressor are needed to see this picture.

13 mm

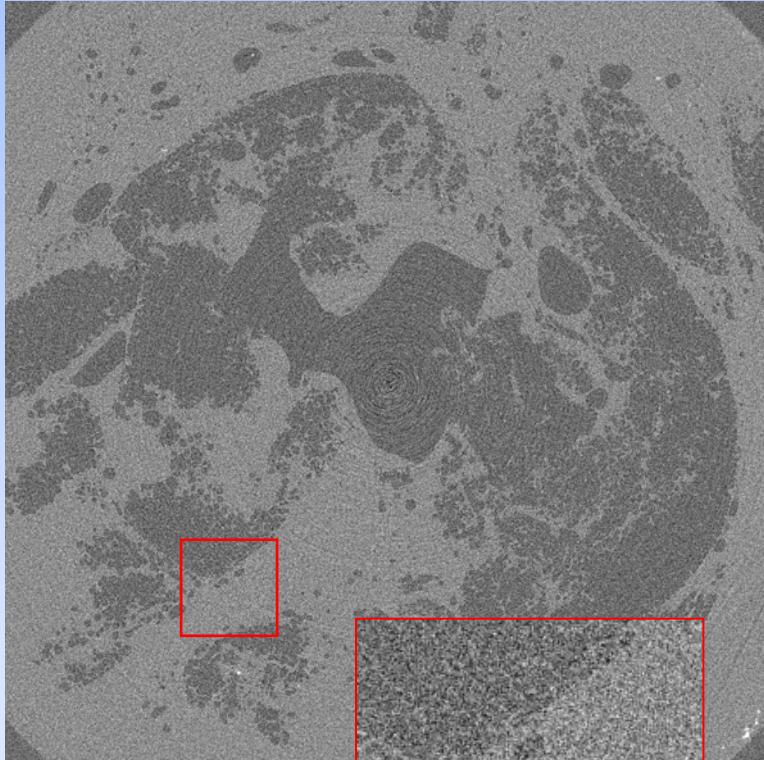
Energy = 25 keV

rad

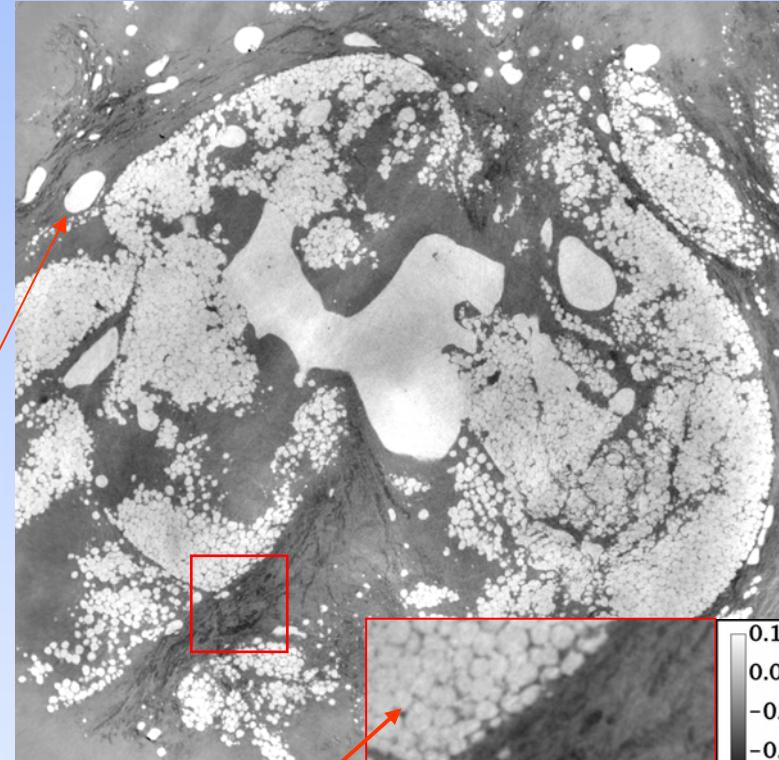
# Larger field of view

Breast Biopsy  
Absorption

$\varnothing \approx 15 \text{ mm}$  ; pixel size =  $7.5 \mu\text{m}$   
Holotomography

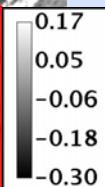


Vessel



Fatty cells

Collagen



$\rho$

$\longleftrightarrow 1.5 \text{ mm}$

# Sub-micron Tomography

- Improve the spatial resolution

$$\lambda < 1 \text{ \AA}$$



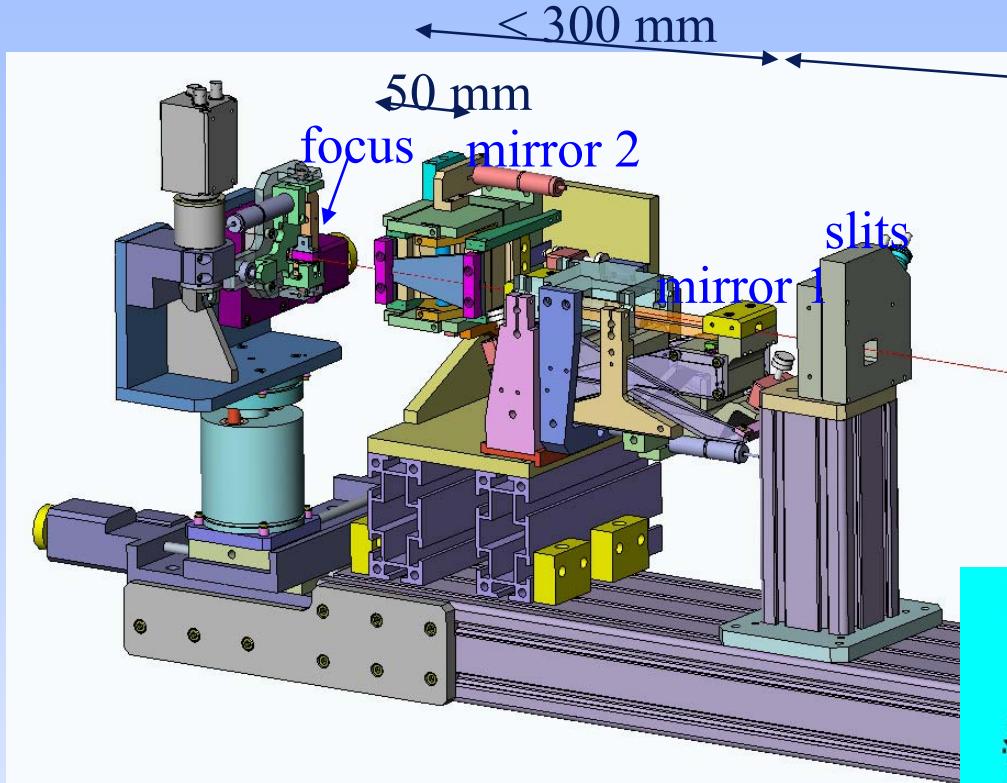
resolution  $\approx 0.5 \text{ \mu m}$

with scintillator based detectors  
parallel beam

Gabor's Microscopic principle (Nature, 1948)

*The object is illuminated by an **electron** beam brought to a **fine focus**... The object is a **small distance behind** (or in front) of the point focus, followed by a **photographic plate** at a large multiple of this distance...*

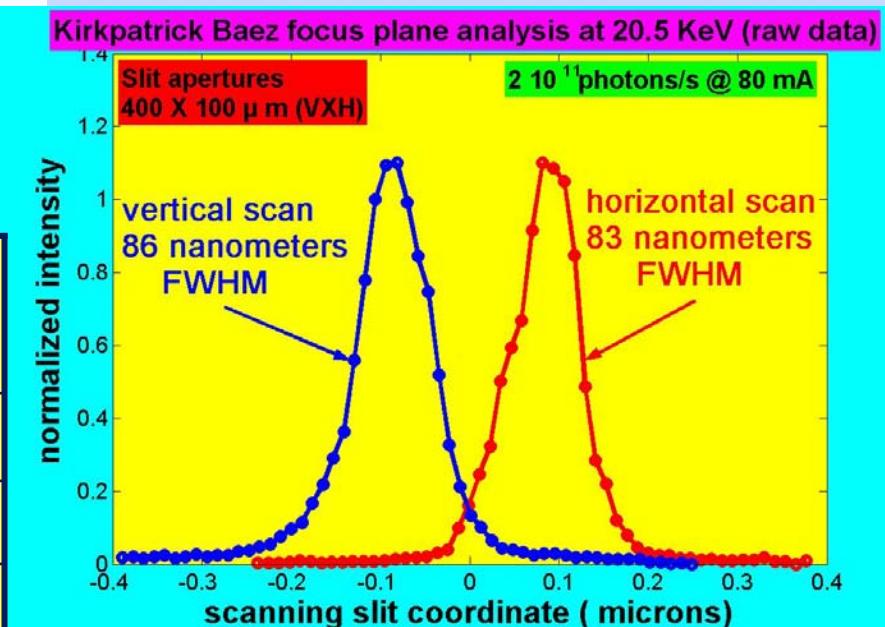
# KB focusing at 20.5 keV



source

Source size  
Mirror quality  
Diffraction

| Aperture V*H ( $\mu\text{m}$ ) | Spot fwhm V*H (nm) | Flux ( $\Delta E/E=10^{-2}$ ) ph/s @ 80 mA |
|--------------------------------|--------------------|--|
| 200 x 50                       | 118 x 109          | $5 \cdot 10^{10}$                          |
| 400 x 100                      | 86 x 83            | $2 \cdot 10^{11}$                          |
| 600 x 160                      | 116 x 90           | $4.5 \cdot 10^{11}$                        |



# Projection Microscopy

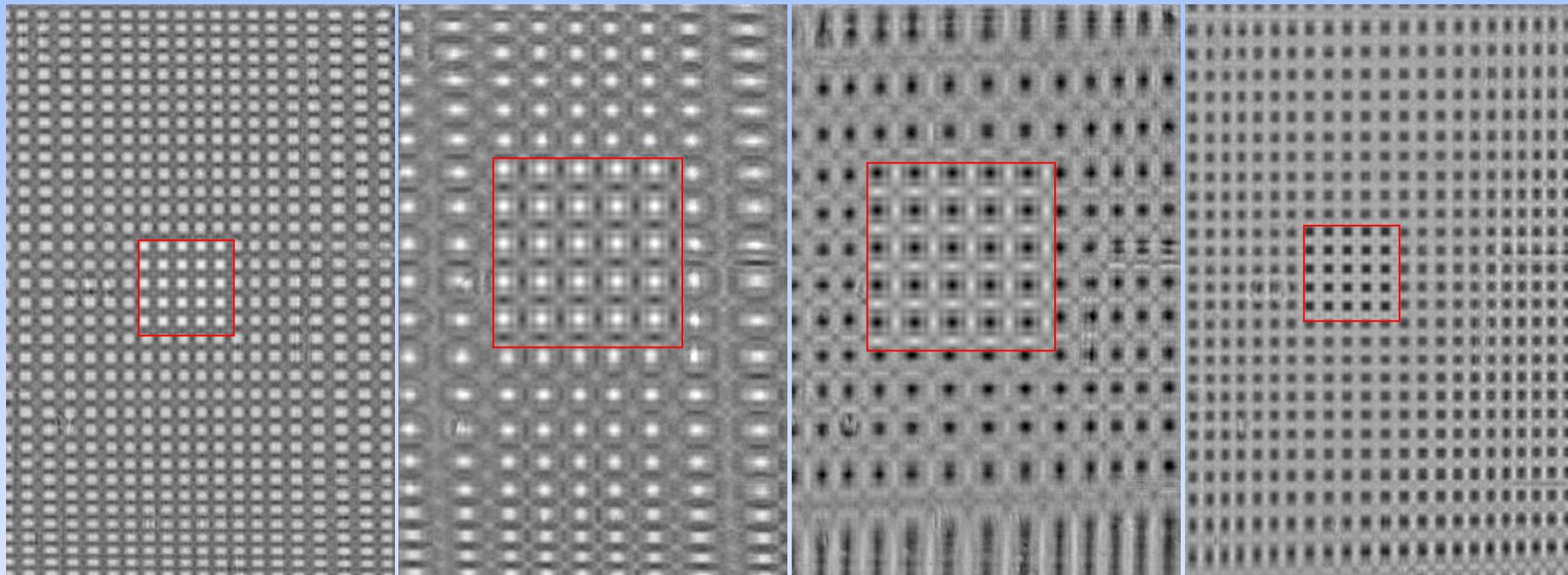
2  $\mu\text{m}$  pitch grating (Si)

C. David, PSI

10  $\mu\text{m}$

Mirrors fully illuminated

0.2 (H) x 1.1 (V) mm,  $E=20.5$  keV



$D = -20 \text{ mm}$

$M = 115$

$D = -10 \text{ mm}$

$M = 230$

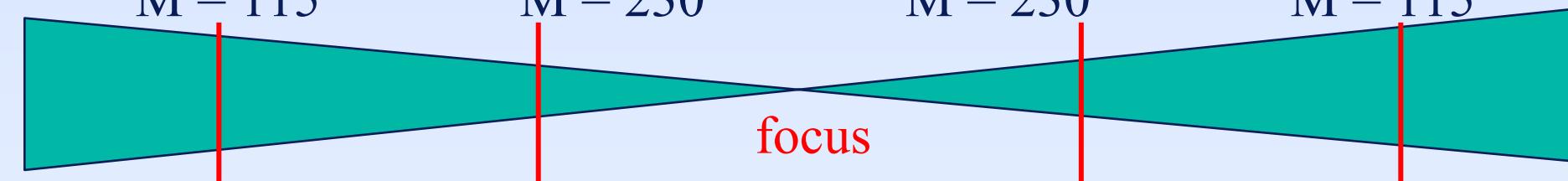
$D = 10 \text{ mm}$

$M = 230$

$D = 20 \text{ mm}$

$M = 115$

focus

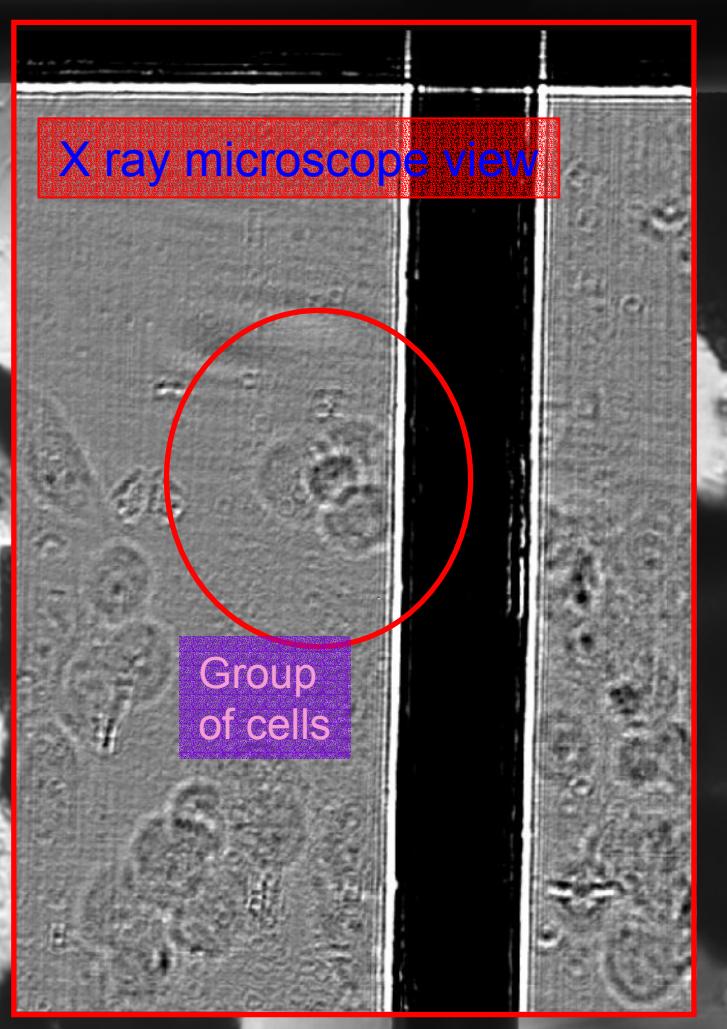
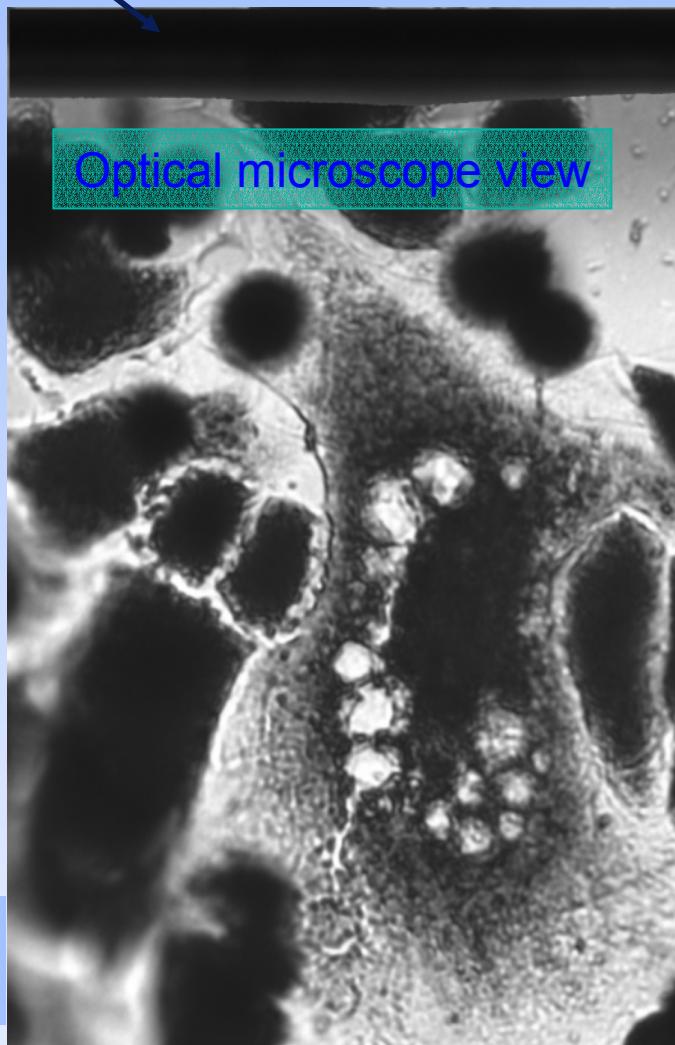


Exposure time = 0.4 s ! (16-bunch, saturation CCD)

P. Cloetens, O. Hignette

# Projection Microscopy

Cu fiber

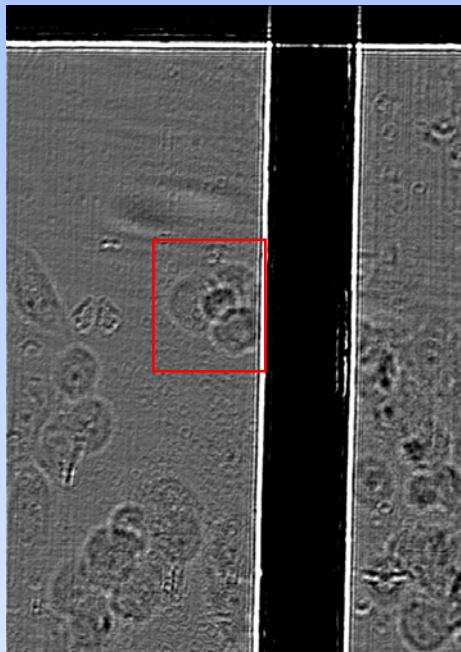


Cellular imaging      Cancerous cells  
S. Bohic

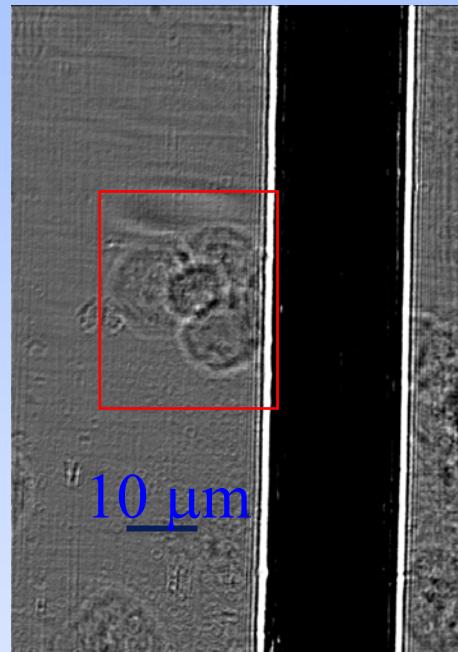
# Projection Microscopy

Cancerous cells  
S. Bohic

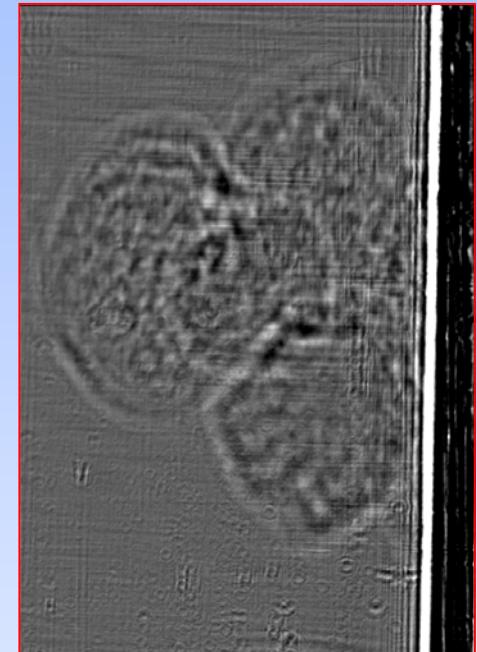
Towards focus



$D = 50 \text{ mm}$   
 $M = 45$



$D = 30 \text{ mm}$   
 $M = 75$



$E = 20.5 \text{ keV}$   
Exposure time = 0.3 s ! (16-bunch)

No X-ray optics behind the sample  $\Rightarrow$  dose efficient

P. Cloetens, W. Ludwig

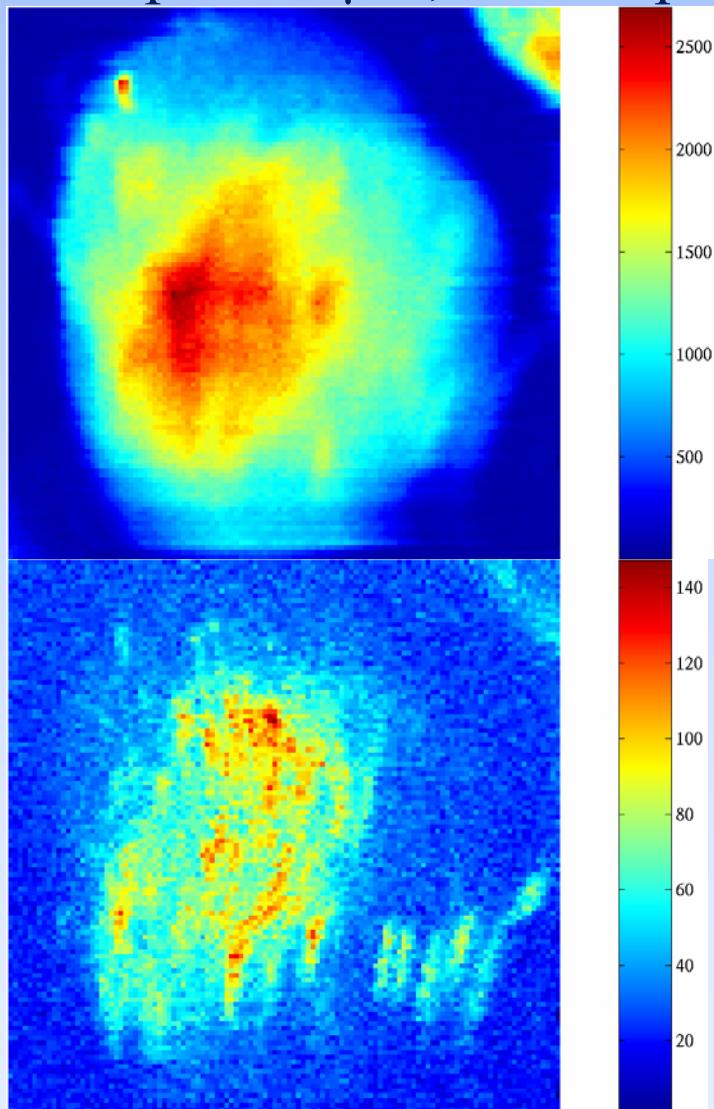
# Applications: Fluorescence

Cancerous cells treated with anti-cancer drug (Cisplatin)

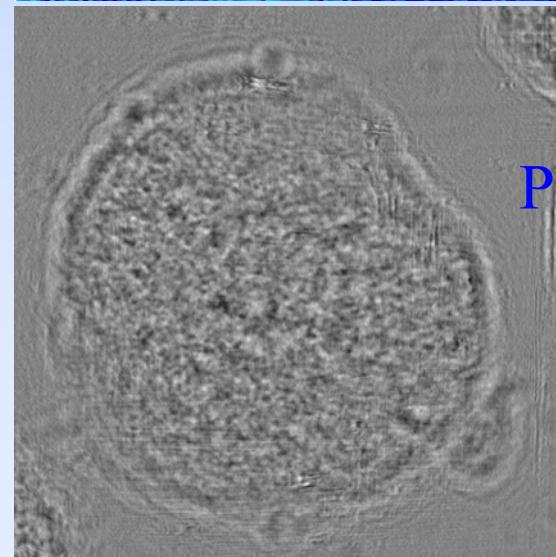
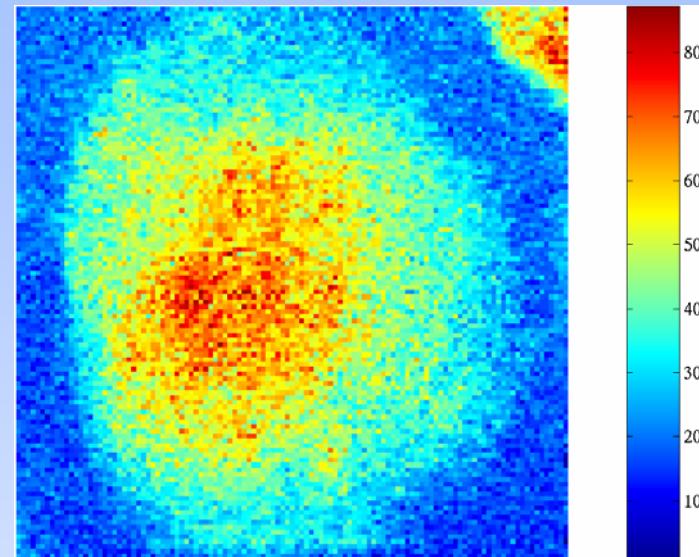
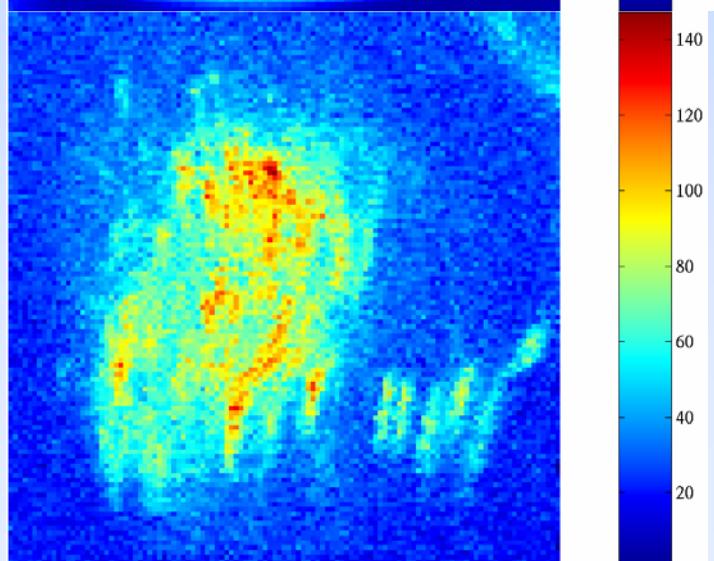
Step = 0.3  $\mu\text{m}$ , 0.9 sec/pt



K



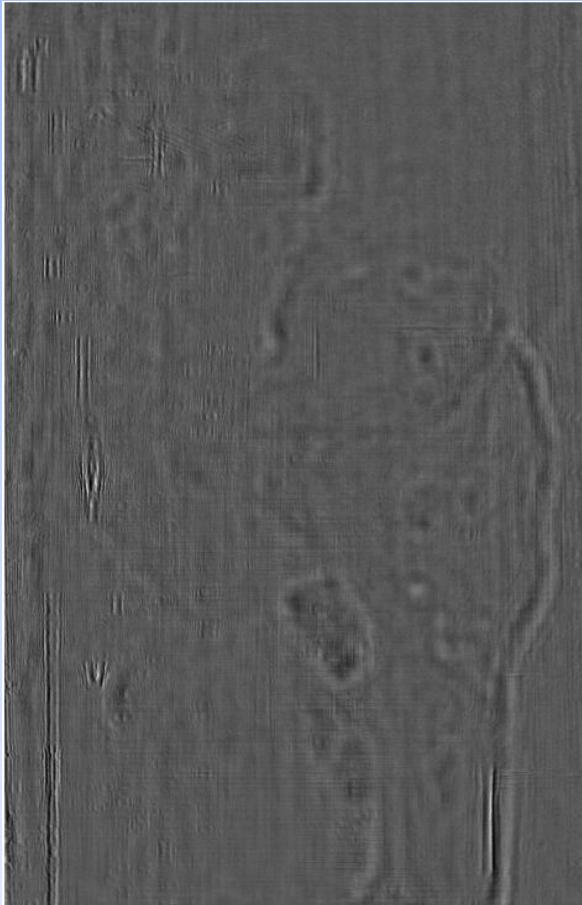
Fe



Phase Contrast

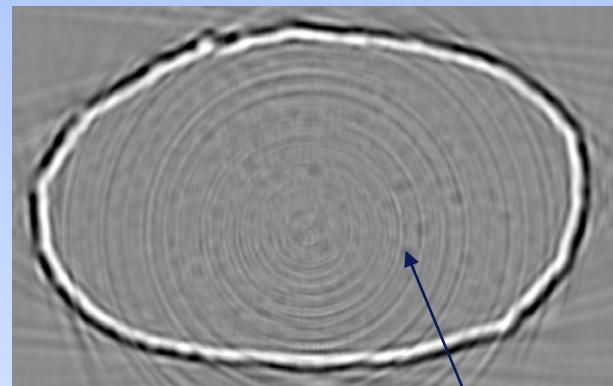
P. Cloetens,  
W. Ludwig,  
S. Bohic

# Projection Microscopy: 3D



Human hair: 80  $\mu\text{m}$  diam  
Distances:  
270, 120, 50 mm from source

Improvement X-ray optics for coherent imaging



Tomographic slice  
Localised structures  
porosity

$D \sim 60 \text{ mm}$        $M=70$

$E = 20.5 \text{ keV}$

Exposure time = 1 s (16-bunch)

# Nano-Tomography Project

Motivation:

Materials Science: relevant scale 0.1-10  $\mu\text{m}$

Nano-technology/fabrication: 50 nm scale and below

Soft condensed matter

Goal: *routine* 3D microscopy with  $< 50 \text{ nm}$  spatial resolution  
with *in-situ* and *micro-analysis* possibilities (7 - 15 keV)

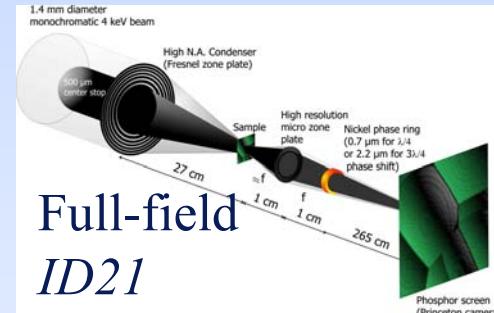
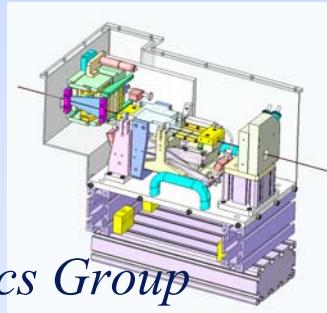
Strategy:

*Multiple-mode 3D microscope*

*Optics*

KB

*Optics Group*



*Coherent / Incoherent Illumination*  
*Full-field / Mapping*



# Conclusions

- Real space imaging often required
- Tomography more powerful than ever: absorption, phase contrast, holotomography, fluo-tomography
- *Quantitative* mapping in 2D and 3D
- *In-situ* experiments
- Multidisciplinary approach

Improve the spatial resolution  
Projection microscopy by KB focusing:  
Sub  $(100 \text{ nm})^2$  focusing made ‘easy’  
High flux ( $10^{12} \text{ ph/s}$ )

**Imaging:** Micro-structure via nano-tomography  
**Nanoprobe:** Element specificity through Fluorescence

30  $\mu\text{m}$

Seed of Arabidopsis

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