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(54) **CLONING AND EXPRESSION VECTORS
AND SYSTEMS**

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ABSTRACT

Vectors for cloning, maintaining and expressing a wide range of coding sequences in inducible T7 expression systems in *Escherichia coli* expression hosts are disclosed herein. Target genes that can be stably maintained and expressed include those that specify proteins that are highly toxic to the host cell. Different configurations of vectors and expression hosts provide different rates of transcription and translation of target genes and therefore different rates of accumulation of target proteins. Methods for cloning by asymmetric ligation and co-expression of more than one target protein in a single vector are also disclosed, as are variants of BL21(DE3) having lower basal transcription by T7 RNA polymerase.

8 Claims, 6 Drawing Sheets

Specification includes a Sequence Listing.

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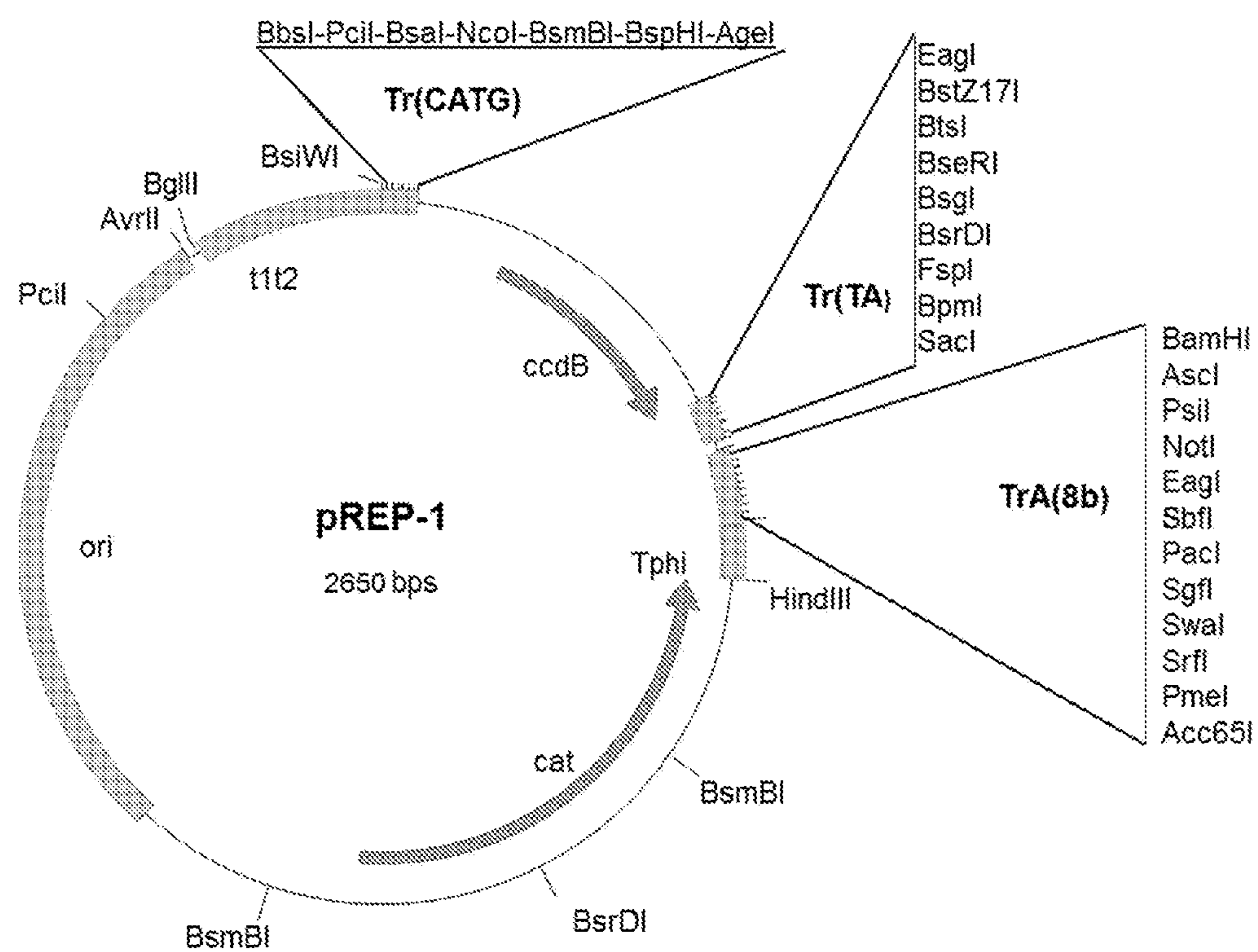


Figure 1

Tr (CATG) SEQ ID NO. 3

BsiWI BbsI PciI BsaI NcoI BsmBI BspHI AgeI

5'-cgtacg**gaagacta|catg tggctctcc|catg gcgtctct|catg aaccggt**-3'

3'-gcatgc**cttctgat gtac|accagagg gtac|cgcagaga gtac|t**tgcca-5'

Tr (TA) SEQ ID NO. 4

EagI BtsI BseRI BsgI BsrDI EciI BpmI SacI

5'-cggccgta ta|**cactgccactcctcctgcac** ta|**cattgcgcacccgcctccagt**gagctc-3'

3'-gccggcat|at **gtgacggtgaggaggacgtg|at gtaacgcgtaggcggaggtc**actcgag-5'

Tr (TG) SEQ ID NO. 5

EagI BtsI BseRI BsgI BsrDI EciI BpmI SacI

5'-cggccgta tg|**cactgccactcctcctgcac** tg|**cattgcgcacccgcctccagt**gagctc-3'

3'-gccggcat|ac **gtgacggtgaggaggacgtg|ac gtaacgcgtaggcggaggtc**actcgag-5'

Figure 2

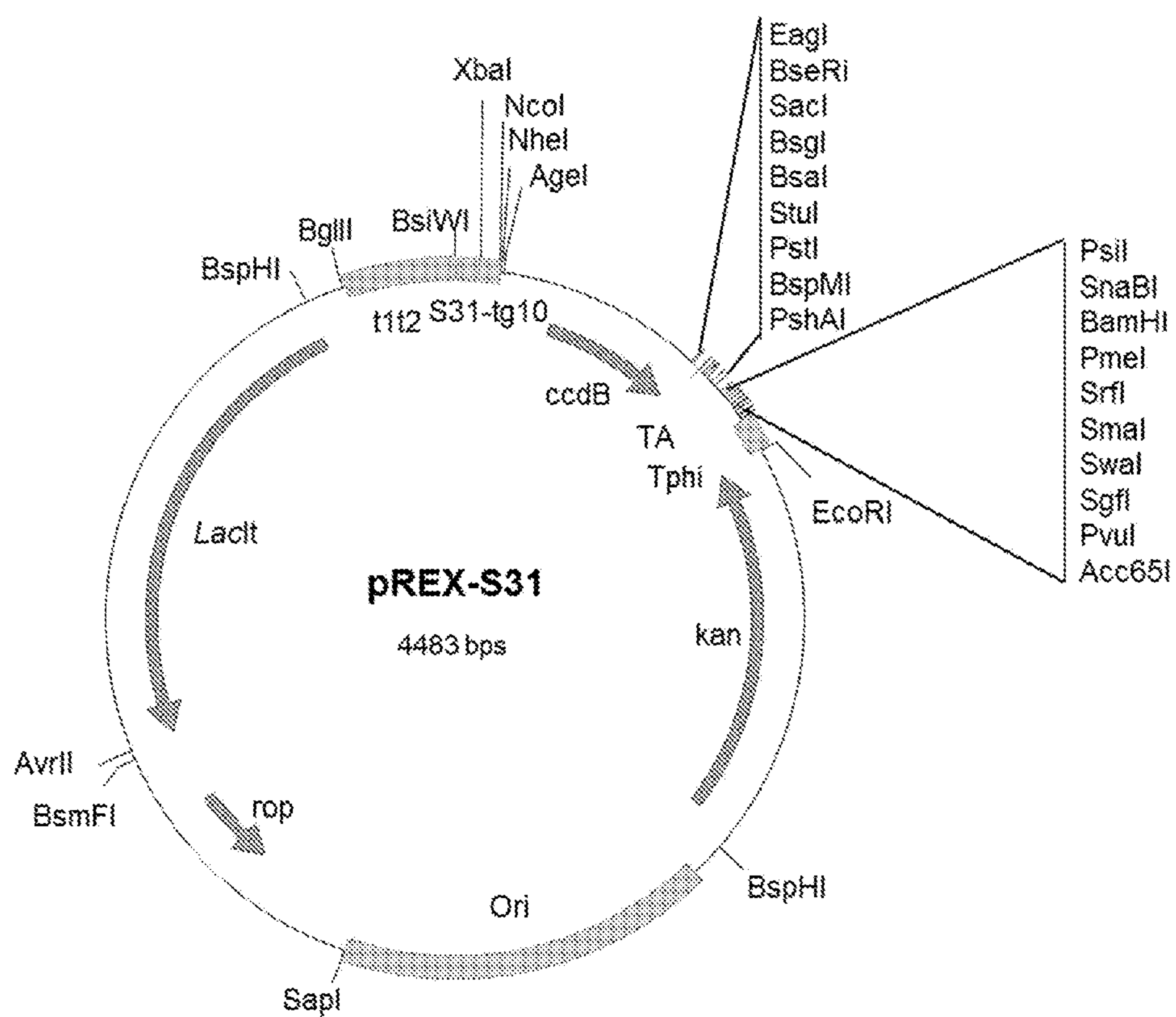


Figure 3

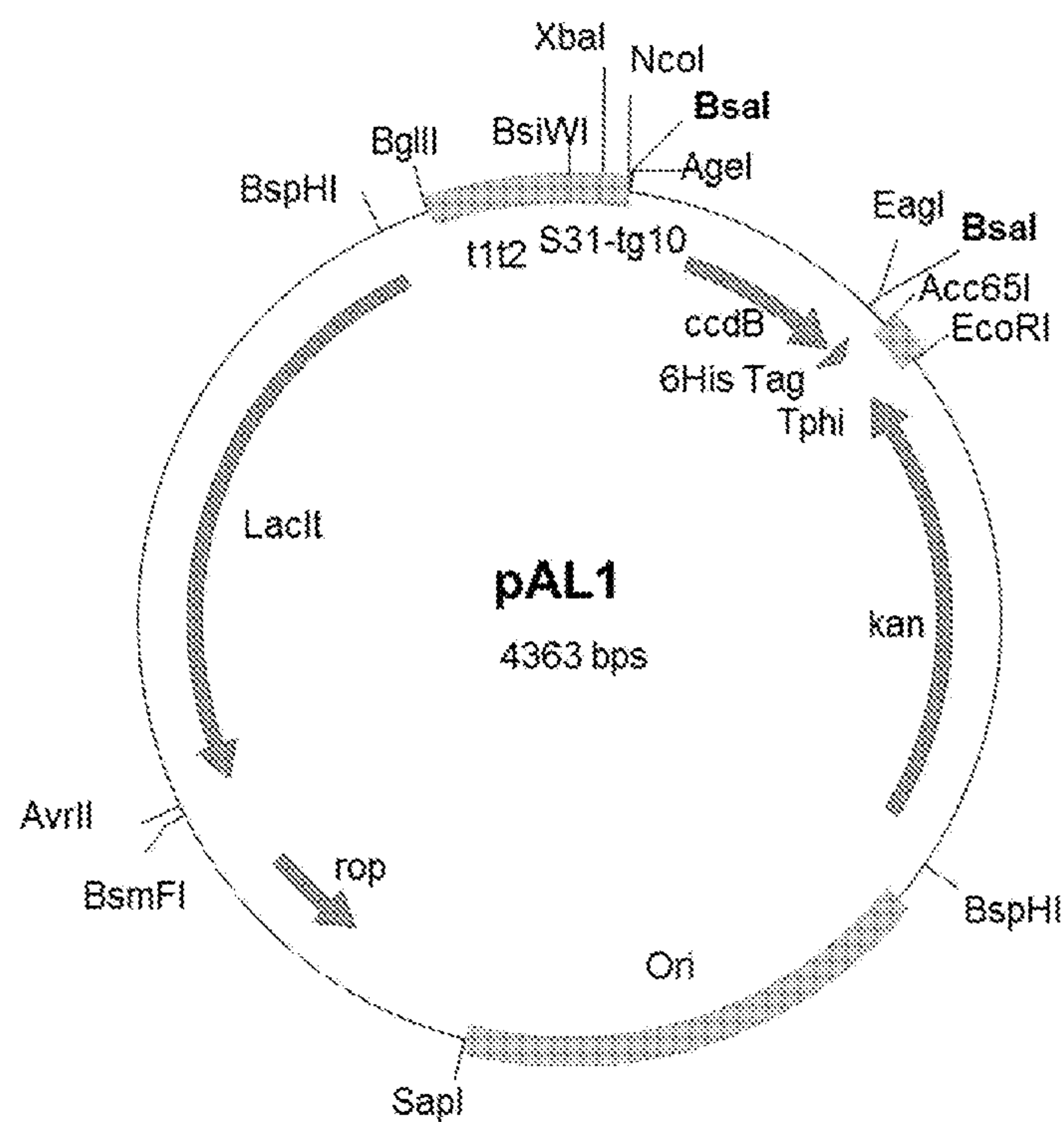


Figure 4

pAL1, 2, 3, 4, 5, 11, 12

PCR ends for generating 4-nt 5' overhangs with T4 DNA polymerase + dCTP

upstream	5'	ccat g-----c	agtg	3'	downstream
	3'	ggta c-----g	tcac	5'	

bp matches in all 10 pairwise alignments
(0 is no bp; 1 is usual DNA bp; - is GT match)

	5'	ccat	atgg	agtg	cact 3'	
upstream aa linkers RF	3' tacc	0000				RF downstream aa linkers
[not MQKEW] -his-VADEG nnc	3' ggta	1111	0--0			nnc [not MQKEW] -his-VADEG
SPTA-met ncc	3' gtga	1001	0000	0--0		nca SPTA-val
pro-CW-[not IMTNK] cca	3' tcac	0000	1001	1111	0000	cag gln-CW-[not IMTNK]

Figure 5A

pAL13, 14

PCR ends for generating 4-nt 5' overhangs with T4 DNA polymerase + dCTP

upstream	5'	actc g-----c	agtg	3'	downstream
	3'	tgag c-----g	tcac	5'	

bp matches in all 10 pairwise alignments
(0 is no bp; 1 is usual DNA bp; - is GT match)

	5'	actc	gagt	agtg	cact 3'	
upstream aa linkers RF	3' ctca	0000				RF downstream aa linkers
[not FMYHND CW] -leu-VADEG nna	3' tgag	1111	-00-			nnc [not MQKEW] -his-VADEG
YHND-ser nac	3' gtga	0--0	1010	0--0		nca SPTA-val
thr-arg act	3' tcac	0100	000-	1111	0000	cag gln-CW-[not IMTNK]

Figure 5B

pAL21, 22, 23, 24

PCR ends for generating 4-nt 5' overhangs with T4 DNA polymerase + dTTP

upstream	5'	ctgc a-----t	agcg	3'	downstream
	3'	gacg t-----a	tcgc	5'	

bp matches in all 10 pairwise alignments
(0 is no bp; 1 is usual DNA bp; - is GT match)

	5'	ctgc	gcag	agcg	cgct 3'	
upstream aa linkers RF	3' cgtc	0--0				RF downstream aa linkers
[not MQKEW] -cys-IMTNKSR nnc	3' gacg	1111	0000			nnt [not MQKEW] -ser-VADEG
YHND-ser nct	3' gcga	0001	0000	0110		nta LIV-gly
Leu-HQ ctg	3' tcgc	0000	100-	1111	0000	tag stop codon

Figure 5C

Cloning and co-expression of two coding sequences (CDS1 and CDS2, overhang sequences capitalized)

upstream overhang		SD		linker overhang		downstream overhang	
5' CCATg-CDS1-taaggagacTATTAATg-CDS2-cAGTG 3'							
- SEQ ID NO. 79 -							
Three perfectly base-paired and 26 imperfectly paired overhangs:							
overhangs		us	usC	1>2	1>2C	ds	dsC
		5' ccat	atgg	tattaat	attaata	agtg	cact 3'
us	3' tacc 5'	0000					
usC	3' ggta 5'	(1111)	0-0				
1>2	3' taattat 5'	0000	100-	0010100			
		0000	110-				
1>2C	3' ataatta 5'	0011	00-0	(1111111)	0010100		
		0001	0000				
ds	3' gtga 5'	1001	0000	-1-1	00-0	0-0	
				-101	01-0		
dsC	3' tcac 5'	0000	1001	0010	1010	(1111)	0000
				0000	1010		

Figure 6A

Cloning and co-expression of three coding sequences (CDS1, CDS2 and CDS3, overhang sequences capitalized)

upstream overhang		SD		linker overhang		SD		linker overhang		downstream overhang	
5' CCATg-CDS1-taaggagacTATTAATg-CDS2-taaggagacTTAATATg-CDS3-cAGTG 3'											
- SEQ ID NO. 79 -											
- SEQ ID NO. 80 -											
Four ungapped, perfectly base-paired overhangs and 48 imperfectly paired overhangs:											
		us	usC	1>2	1>2C	2>3	2>3C	ds	dsC		
overhangs		5' ccat	atgg	tattaat	attaata	ttaatat	atattaa	agtg	cact	3'	
us	3' tacc 5'	0000									
usC	3' ggta 5'	1111	0-0								
1>2	3' taattat 5'	0000	100-	0010100							
		0000	110-								
1>2C	3' ataatta 5'	0011	00-0	1111111	0010100						
		0001	0000								
2>3	3' tataatt 5'	0010	01--	0001010	1100001	0110110					
		0011	11-0								
2>3C	3' aattata 5'	0011	11-0	1000011	0101000	1111111	0110110				
		0010	01--								
ds	3' gtga 5'	1001	0000	-1-1	00-0	-000	0001	0-0			
				-101	01-0	0001	-000				
dsC	3' tcac 5'	0000	1001	0010	1010	0000	1000	1111	0000		
				0000	1010	1000	0000				

Figure 6B

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**CLONING AND EXPRESSION VECTORS
AND SYSTEMS****CROSS-REFERENCE TO RELATED
APPLICATIONS**

This application is a US National Phase Application under 35 USC § 371 of International Patent Application No. PCT/US2017/017061, filed on Feb. 8, 2017, which claims the benefit of prior U.S. Provisional Application No. 62/293,089, filed Feb. 9, 2016, all of which are hereby incorporated by reference in their entirety.

STATEMENT OF GOVERNMENT RIGHTS

The present application was made with government support under contract number DE-SC0012704 awarded by the U.S. Department of Energy. The United States government has certain rights in the invention(s).

BACKGROUND

Transcription of cloned genes by T7 RNA polymerase and translation of the mRNA directed by the highly efficient translation initiation sequence upstream of the coding sequence for the T7 major capsid protein (referred to here as tg10), has been highly successful in producing a wide range of proteins in *E. coli* (Studier et al. (1986) J. Mol. Biol. 189:113-130; Rosenberg et al. (1987) Gene 56:125-135; Studier et al. (1990) Met. Enzymology 185:60-89; Studier (2005) Protein Expr. Purif. 41:207-234). T7 RNA polymerase selectively initiates transcription at a promoter sequence (referred to here as a T7 promoter) large enough to be unlikely to be found by chance in a bacterial genome. No sequences that appear likely to serve as promoters for T7 RNA polymerase are found in completely sequenced genomes of *E. coli* expression strains such as BL21(DE3). T7 RNA polymerase is highly active and processive, and is capable of producing mRNA for a wide range of coding sequences linked to a T7 promoter in cloning vectors such as the commonly used pET vectors. This highly selective transcription and efficient tg10-directed translation can commandeer the resources of the *E. coli* cell so efficiently that, in favorable cases, almost all of the protein produced shortly after T7 RNA polymerase is induced or delivered to the cell is that specified by the cloned gene (referred to here as the target protein), which can accumulate to become more than half of the total protein in the cell.

The original pET vectors and their many successors were derived from the multi-copy plasmid pBR322, which has the colE1-type replicon from pMB1 (Rosenberg et al (1987) and Bolivar et al. (1977) Gene 2:95-113). The cloning site in the tet gene is oriented so that transcription from the T7 promoter is opposite to that from the tet promoter. Basal transcription by *E. coli* RNA polymerase at this site is low enough that most coding sequences can be cloned and maintained in the absence of T7 RNA polymerase, even if the target protein is toxic to the host cell. A convenient way to produce target proteins is for the host to supply inducible T7 RNA polymerase which, upon induction, directs expression of the target gene from a T7 promoter. However, T7 RNA polymerase is so active and tg10 so efficient that basal expression in the uninduced cell can produce enough target protein to prevent establishment of an inducible expression strain if the target protein is sufficiently toxic to the host cell. This problem was encountered in the earliest attempts to use inducible T7 RNA polymerase to produce proteins from

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cloned genes, and several improvements have reduced basal levels of target protein in uninduced cells, thereby stabilizing expression strains and allowing a wider range of proteins to be produced.

One early improvement was to supply a small amount of T7 lysozyme, which binds to T7 RNA polymerase and inhibits transcription, thereby reducing basal production of target protein (Studier (1991) J. Mol. Biol. 219:37-44). Upon induction, more T7 RNA polymerase is made than can be inhibited by the T7 lysozyme and high levels of target protein can be produced.

Another solution was to place a binding site for lac repressor (referred to as an operator) immediately downstream of the start site for T7 RNA polymerase (Giordano et al. (1989) Gene 84:209-219; Dubendorff et al. (1991) J. Mol. Biol. 219:45-59; and U.S. Pat. No. 6,537,779 to Kara et al. (2003; published Feb. 4, 1999), the contents of which are incorporated herein by reference). One such construct, referred to as a T7lac promoter (Dubendorff, et al. (1991)) has found wide use in pET vectors. Bound lac repressor reduces transcription from a T7lac promoter, thereby reducing basal production of the target protein. Expression of T7 RNA polymerase in BL21(DE3) and several other expression hosts is also controlled by lac repressor, so that an inducer that reduces affinity of the lac repressor for its operators both turns on production of T7 RNA polymerase and unblocks the T7 promoter, allowing high-level production of target protein.

A third type of solution is to express T7 RNA polymerase from an inducible *E. coli* promoter that has lower basal expression than the L8, lacUV5 promoter used in BL21 (DE3) (Miroux et al. (1996) J. Mol. Biol. 260:289-298; Schlegel et al. (2015) Cell Reports 10:1758-1766 and the arabinose-inducible pBAD promoter in BL21-AI (Invitrogen, now Life Technologies, a Thermo Fisher Scientific Brand)). In BL21-AI, induction of the pBAD promoter by arabinose and unblocking the T7lac promoter by an inducer of the lac repressor would both be necessary for maximal production of target protein.

However, even with these improved vectors or expression strains, some target proteins are toxic enough that inducible strains for producing them are unstable and can be maintained only with special care, or are so toxic that inducible expression strains cannot be established at all. This problem was encountered in trying to maintain and express gene 5.3 of bacteriophage T7, which specifies a protein of unknown function that is predicted to contain 118 amino acids (Dunn et al. (1983) J. Mol. Biol. 166:477-535).

Therefore, there remains a need for improved cloning and expression vectors and systems that provide tighter control of expression and improved methods of cloning into the same.

SUMMARY OF THE INVENTION

The present invention provides vectors and methods for cloning, maintaining, and expressing coding sequences in inducible T7 expression systems.

In one embodiment, the present invention provides a T7 Promoter Control Region polynucleotide including SEQ ID NO. 17, 18, 19, 23, 24, 25, 29, 30, 31, 32, 33, 34, 35, 38, 39, 40, 41, 44, 45, 46, and 47.

In one embodiment, the present invention provides a Translation Initiation Efficiency Region polynucleotide including SEQ ID NO. 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 87, 88, 89, 90, 91, 92, and 93.

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In one embodiment, the present invention provides an Expression Control Region polynucleotide that includes a T7 Promoter Control Region polynucleotide and a Translation Initiation Efficiency Region polynucleotide.

In one embodiment, the present invention provides a vector that includes an Expression Control Region polynucleotide.

In one embodiment, the present invention provides a vector for directional cloning of at least one target DNA by asymmetric ligation. The vector includes a cloning site with upstream and downstream outward-facing asymmetric recognition sequences for at least one Type IIS restriction endonuclease that cuts outside of its recognition sequence to produce single-stranded overhangs of arbitrary nucleotide sequence. The recognition sequences are situated such that cutting said vector with said at least one Type IIS endonuclease produces two vector fragments: 1) a cloning-acceptor fragment having a different asymmetric overhang at each end; and 2) a counter-selection fragment containing both of said outward-facing recognition sequences and having asymmetric overhangs complementary to those of said cloning-acceptor fragment.

The nucleotide sequences of said 4 asymmetric overhangs are designed so that only 2 of the 10 possible pairwise alignments between said overhangs can form ungapped perfectly base-paired substrates for efficient joining by a DNA ligase to join either strand, thereby regenerating said vector and additionally so that the number of consecutive perfect base-pairs from either ungapped end of each of the remaining 8 said possible pairwise alignments is minimized, thereby minimizing potential joining of either DNA strand at any of said remaining 8 possible pairwise aligned overhangs by said DNA ligase, and so that the potential for base-pairing adjacent to any ungapped end in each of the remaining 8 possible pairwise alignments is minimized so as to decrease probability of being joined by said DNA ligase.

In one embodiment, the present invention provides a method for directional cloning of at least one target DNA in a cloning vector disclosed herein. The method includes: 1) cutting said vector with said at least one Type IIS restriction endonuclease to produce said cloning-acceptor fragment and said counter-selection fragment; 2) providing said at least one target DNA having asymmetric single-stranded overhangs designed such that the only pairwise alignments of said overhangs of said at least one target DNA and of said cloning-acceptor fragment that can form ungapped perfectly base-paired substrates for efficient joining by a DNA ligase are those whose sequential ligation generates a directional clone of said at least one target DNA in said vector cloning-acceptor fragment or those whose ligation rejoins said cloning-acceptor and said counter-selection fragments, whereas each of the greater number of other possible pairwise alignments of said overhangs has few if any complementary base pairs near its ends and therefore has a much lower probability of being joined by said DNA ligase; 3) assembling said cut vector or said cloning-acceptor fragment produced in step 1, said at least one target DNA having said asymmetric single-stranded overhangs, and said DNA ligase in a solution and incubating under conditions that promote ligation of ungapped perfectly base-paired substrates by said DNA ligase to provide ligation products; 4) transforming said ligation products into cells; and 5) selecting transformants that contain a directional clone of said at least one target DNA in said cloning-acceptor fragment of said vector.

In one embodiment, the invention provides a method for co-expression of a plurality of coding sequences from one expression vector. The method includes obtaining a clone in

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said vector by the methods disclosed herein, establishing said clone in a T7 expression host, inducing production of T7 RNA polymerase in said T7 expression host, producing target mRNA by transcription of target coding sequences from a T7 promoter in the Expression Control Region polynucleotide of said vector, and initiating production of target proteins from upstream translation initiation regions for each coding sequence.

In one embodiment, the present invention provides an engineered cell including a vector disclosed herein comprising an at least one target DNA encoding at least one target protein operationally linked to said Expression Control Region polynucleotide; and an inducible T7 RNA polymerase.

DESCRIPTION OF THE FIGURES

FIG. 1 depicts a plasmid map of pREP-1 vector, SEQ ID NO. 1.

FIG. 2 depicts examples of coding sequences that include an upstream 5'-CATG single-stranded DNA overhang and a downstream TA-3' overhang.

FIG. 3 depicts a plasmid map of pREX-S31 vector, SEQ ID NO. 62.

FIG. 4 depicts a plasmid map of pAL1 vector, SEQ ID NO. 66.

FIGS. 5A-5C depicts base pairs (bp) in all 10 ungapped pairwise alignments of 4-nt 5' overhangs for cloning single coding sequences by asymmetric ligation in three sets of pAL vectors, showing potential upstream and downstream amino-acid linkers to fusion peptides or protein domains in three reading frames (RF).

Linker amino acids (aa) in sequential positions are separated by a dash; a single aa allowed at a position is in lower case three-letter code; multiple aa allowed at a position are in upper case single-letter code; multiple aa excluded at a position are enclosed in brackets where the majority of aa are allowed.

FIGS. 6A-6B depicts all pairwise matches of overhangs for cloning 2 or 3 coding sequences by asymmetric ligation in pAL1. Overhangs of coding sequences are designed to be generated by digestion with T4 DNA polymerase in the presence of dCTP Abbreviations; SD is Shine-Dalgarno; us is upstream; ds is downstream; C at the end means complement.

In each matrix, 0 is no bp; 1 is a usual DNA bp; - is a GT match; perfect matches are circled. The two matches given between overhangs of unequal lengths are the ungapped alignments at the left and right ends.

DETAILED DESCRIPTION

The present invention provides vectors and methods for cloning, maintaining, and expressing coding sequences in inducible T7 expression systems.

The coding sequences include polynucleotides that encode for a target protein.

Strategy for Improving Vectors

Part of the strategy for improving vectors is to include an Expression Control Region as described below.

Ideally, basal expression would be reduced to a level where intervals between stochastic bursts of transcription and translation of a target gene in an uninduced cell are appreciably longer than a division cycle. If so, and if the amount of target protein produced per burst is low enough,

any expression clone could be maintained in a growing culture in an inducible state. Even if every cell in which a burst of target gene expression occurred were to be killed or prevented from dividing, continuing division of unaffected cells would increase the population of cells that remain capable of induction faster than dead cells would accumulate. The longer the average interval between stochastic bursts of target gene expression relative to division time, and the smaller the average burst of target protein produced, the greater the fraction of cells in the culture that will remain fully competent for inducible production of target protein. The amount of target protein produced upon induction of the culture will then depend primarily on the fraction of inducible cells in the culture and the effect (if any) of target protein on the ability of the induced cell to produce protein.

The strategy for obtaining vectors capable of producing virtually any target protein in an inducible T7 expression strain was to design, construct, test and improve two sets of vectors: 1) pREP vectors (FIGS. 1-2) having a cloning and retrieval site that can accept and maintain a coding sequence with the least possible expression, and 2) pREX vectors (FIG. 3) having an Expression Control Region that regulates and directs production of target protein in an inducible T7 expression strain, and an adjacent cloning site that accepts a coding sequence retrieved from a pREP clone and places it under control of the Expression Control Region. These new vectors were derived from pET vectors and were designed to eliminate superfluous DNA, minimize the potential for basal production of target protein, and to be modular, so that different components can be tested and exchanged easily through (mostly) unique restriction endonuclease cleavage sites. Several pREP vectors were designed but all of the coding sequences tested were easily cloned, maintained and retrieved from pREP-1 (SEQ ID NO. 1), which has been used almost exclusively. Many different pREX vectors have been constructed and tested, as disclosed herein.

In addition to using the T7 gene 5.3 coding sequence to test ability to maintain and express highly toxic target genes, the well-expressed T7 gene 10a coding sequence for the major T7 capsid protein has been used to test expression levels.

Construction of pREP and pREX Vectors

The pREX vectors contain approximately the minimal origin of replication of pET vectors, including the *rop* gene that ensures moderate copy numbers. However, the pREP-1 origin was further reduced to eliminate the *rop* gene, and pREP-1 has a considerably higher copy number characteristic of pUC vectors. The higher copy number of pREP-1 is meant to facilitate plasmid isolation for sequencing and retrieval of cloned coding sequences. The moderate copy number of pREX vectors is meant to make it easier to achieve lower levels of basal expression while retaining high levels of induced production of target protein typical of pET vectors.

To minimize the potential for synthesis of target mRNA by *E. coli* RNA polymerase (or RNA polymerases of any host cell), all known coding sequences in the cloning-acceptor fragment of both pREP-1 and pREX vectors plus the actively synthesized RNA I of the replication origin are oriented so that readthrough transcription that might reach a cloned target coding sequence will produce anti-sense RNA. RNA II, which primes replication of the plasmid in the opposite orientation of RNA I is the only known exception, and the *t1t2* double transcription terminator of the *rrnB* ribosomal RNA operon placed immediately upstream of the

cloning and retrieval site in pREP-1 and the Expression Control Region of pREX vectors (and the pAL vectors derived from them) is meant to minimize potential synthesis of target mRNA from possible readthrough transcription from RNA II or unknown or adventitious promoters for a host RNA polymerase.

A different selective agent together with a common counter-selection module facilitates cloning and transfer of coding sequences between pREP-1 and pREX vectors. The *cat* gene confers resistance to chloramphenicol in pREP-1 and the *kan* gene confers resistance to kanamycin in pREX vectors. Other genes have been contemplated and may be used herein. Examples of other genes that can be used for selection include the *amp^R* gene which confers resistance to ampicillin, and the *tet^R* gene which confers resistance to tetracycline.

The counter-selection module (SEQ ID NO. 2) constitutively expresses the *ccdB* toxin of the *E. coli* F factor from the upstream expression signals for the *ccdA* anti-toxin gene, which has been deleted. The *ccdB* toxin kills cells used for cloning and expression by interrupting the action of DNA gyrase, thereby eliminating the background of unwanted colonies containing uncut or reconstituted vectors on transformation plates for selecting clones (Bernard et al. (1994) *Gene* 148:71-74; and U.S. Pat. Nos. 6,180,407 and 5,910,438 to Bernard et al., the contents of which are incorporated herein by reference). Recognition sites for several restriction endonucleases were eliminated from the counter-selection module to ensure that counter-selection would not be inactivated in cloning or retrieval processes involving pREP, pREX or pAL vectors. All three types of vectors must be maintained in and produced from a host that is resistant to the *ccdB* toxin. The *gyrA462* mutation provides resistance and strain DB3.1TM (Invitrogen, *E. coli* RR1 *gyrA462 endA recA*) is a suitable host. However, the *ccdB* SurvivalTM strain that replaced DB3.1 in the Invitrogen/Life Technologies catalog, which attempts to counter *ccdB* toxin by producing *ccdA* anti-toxin, was found to be unsatisfactory because vectors maintained in this host frequently lost the ability to kill the usual transformation hosts.

Other counter selection genes have been contemplated and may be used in the vectors disclosed herein. Examples of other counter selection genes include *sacB*, *rpsL*, *tetAR*, *pheS*, *thyA*, *lacY*, and *gata-1*. See Reyrat et al., *Infection and Immunity*, September 1998, p. 4011-4017.

Both pREP and pREX vectors have clusters of recognition sites for other restriction endonucleases following the downstream acceptor site, in anticipation that they might be useful for cloning in special situations (FIGS. 1 and 3); however, they have not been used and are not included in pAL vectors (FIG. 4). The Tphi terminator of transcription by T7 RNA polymerase from T7 DNA follows these superfluous cloning sites in both pREP and pREX vectors. Besides limiting extensive readthrough transcription past the target mRNA in pREX after induction, Tphi provides a 3' stem-loop structure that makes target mRNA considerably more stable than most *E. coli* mRNAs.

Cloning and Retrieval Site in pREP-1 and Acceptor Site in pREX Vectors

Any coding sequence with an upstream 5'-CATG single-stranded DNA overhang and a downstream TA-3' overhang can be perfectly paired to complementary acceptor overhangs in the cloning and retrieval site of the cloning-acceptor fragment of pREP-1 to provide substrates for rapid ligation (FIG. 2 and SEQ ID NOs. 3 and 4). The resulting

clone contains restriction endonuclease cleavage sites that allow retrieval of the coding sequence with the same overhangs. (Nucleotide sequences are given left to right in the 5' to 3' direction unless specified otherwise, using the DNA single-letter designations for both DNA and RNA.) The ATG in the upstream overhang of the coding sequence specifies the initiation codon and the downstream TA immediately follows the codon for the last amino acid of the coding sequence. The cloning-acceptor fragment of pREX vectors accepts the upstream 5'-CATG overhang at a complementary overhang produced by cleavage at a unique NcoI site that ends the Expression Control Region (SEQ ID NO. 6), and accepts the downstream TA-3' overhang at a complementary overhang produced by cleavage at a unique BseRI cleavage site (SEQ ID NO 7).

The downstream TA acceptor in pREX vectors is followed by an A nucleotide to create a TAA termination codon, thereby directing expression of the exact target coding sequence. Equivalent pairs of pREP and pREX vectors could also be constructed with TG instead of TA as the downstream overhang (FIG. 2 and SEQ ID NO. 5).

The CATG and TA overhangs were chosen for flexibility to produce target proteins fused to N- and/or C-terminal peptides or protein domains supplied by coding sequences in different pREX vectors through linkages as short as 0-3 amino acids. However, much more construction and modification of pREX vectors than anticipated was required to obtain the desired significant reductions in basal expression, and fusions to target proteins have instead been developed in the more convenient and versatile pAL vectors.

Various Type IIS restriction endonucleases have a non-palindromic (asymmetric) recognition sequence and cut at specific positions outside of the recognition site regardless of the sequence at the cut sites. Since the distance between recognition and cleavage sites can be appreciably different for different enzymes, two or more recognition sequences for different Type IIS and Type II restriction endonucleases can be placed so that each of the enzymes cuts to produce the same acceptor overhang for cloning. The upstream cloning and retrieval site in pREP-1, designated Tr(CATG) (FIG. 2 and SEQ ID NO. 3), contains 6-bp recognition sequences for three pairs for Type IIS and Type II restriction endonucleases, placed so that each member of a pair cuts to produce 5'-CATG overhangs at the same position, so that an acceptor overhang can optionally be generated at any of three sites. In clockwise (left-to-right) order the three pairs of recognition sequences for this first set of restriction nucleases is BbsI-PciI, BsaI-NcoI and BsmBI-BspHI and they are preceded by a unique BsiWI site and followed by a unique AgeI site. Likewise, the downstream cloning and retrieval site, designated Tr(TA) (FIG. 2 and SEQ ID NO. 4), contains 6-bp recognition sequences for two triplets of Type IIS restriction endonucleases, placed so that each member of a triplet cuts to produce TA-3' overhangs at the same position, so that an acceptor overhang can optionally be generated at either of two sites. The clockwise order of the two triplets of recognition sequences for this second set of restriction endonucleases is BtsI-BseRI-BsgI and BsrDI-EciI-BpmI and they are preceded by an EagI site and followed by a unique Sad site. At least one recognition sequence in each pair or triplet is unique in pREP-1, so each of the three upstream and two downstream acceptor overhangs can be generated individually. As noted above, equivalent pairs of pREP and pREX vectors could also be constructed with TG instead of TA as the downstream acceptor overhang (FIG. 2 and SEQ ID NO. 5).

The vast majority of coding sequences will lack a 6-bp recognition site for at least one of the six restriction endonucleases in the upstream set and at least one of the six restriction endonucleases in the downstream set. Such coding sequences can be amplified by PCR using forward and reverse primers that each supply the recognition and cleavage sequence in the position and orientation such that cutting the PCR product with two restriction endonucleases that do not cut within the coding sequence will generate the upstream 5-CATG and downstream TA-3' (or TG-3') overhangs for cloning. The coding sequence can be retrieved from the pREP-1 clone for transfer to a pREX vector by cutting with the same two restriction endonucleases used to produce the DNA fragment cloned even if other cleavage sites are present elsewhere in the pREP vector, because the overhangs at outside sites will usually not compete significantly for cloning into the pREX acceptor overhangs. If necessary, coding sequences can be mutated or chemically synthesized to eliminate one or more internal cleavage sites for enzymes used in cloning and retrieval, but this was not necessary for any of the coding sequences tested.

Other Measures to Minimize Potential Expression of Active Target Protein in pREP-1

In addition to the previously discussed measures taken to prevent or reduce potential transcription of cloned coding sequences in pREP-1, the sequences ahead of the initiation codon for the cloned target protein in the three upstream cloning acceptors are designed, as much as possible, to be unfavorable for ribosome binding and initiation of target protein synthesis from any target mRNA that might happen to be made. Furthermore, the target coding sequence does not terminate at either of the two TA-3' downstream cloning acceptors in pREP-1 (SEQ ID NO. 1) but continues for 54 amino acids past the first acceptor site and 11 amino acids past the second acceptor site before terminating, thereby generating C-terminal fusions that may diminish the toxicity of any target protein that might happen to be produced. Every toxic T7 gene tested and every target coding sequence obtained from collaborators who were unable to obtain clones in pET vectors was easily cloned, maintained and retrieved from pREP-1.

Expression Control Region of pREX and pAL Vectors

The Expression Control Region of the invention includes a T7 Promoter Control Region and Translation Initiation Efficiency Region as described below.

The DNA sequence between the end of the t1t2 double transcription terminator sequence and the ATG initiation codon for the target protein in pREX and pAL vectors is referred to here as the Expression Control Region (ECR). As with many pET and other vectors, the ECRs of pREX and pAL vectors are derived from elements in T7 DNA that direct expression of the T7 gene 10 major capsid protein, the most highly expressed protein during T7 infection, combined with one or two binding sites for lac repressor (Rosenberg et al. (1987); Studier et al. (1990); Dubendorff et al. (1991); U.S. Pat. No. 6,537,779 (2003)). Expression Control Regions of pET-3a, pET-11d, ZA #11 and ZA #13 are aligned with the ECR of T7 gene 10 in Table 1 to highlight similarities and differences (SEQ ID NOs. 8-12).

Expression Control Regions of pREX and pAL vectors are here divided into a T7 Promoter Control Region (TPCR), which directs transcription of the target gene, and adjacent

Translation Initiation Efficiency Region (TIER), which largely determines how well target mRNA competes for binding to ribosomes and initiation of translation to produce target protein. The TPCRs and TIERs that comprise the ECRs of different pREX and pAL vectors are also aligned with the ECRs of T7 gene 10 and previous vectors in Table 1. The TPCRs of pREX and pAL vectors disclosed herein are identified in Tables 1 and 3 and/or with SEQ ID NOs. 13-47, and TIERs disclosed herein are identified in Tables 1-3 and/or with SEQ ID NOs. 48-59.

In clockwise order (left to right in Table 1), the TPCRs in those pREX and pAL vectors that have a single lac operator comprise: a BsiWI site and overlapping six base pairs of T7 DNA ahead of the T7 gene 10 promoter sequence; the 17-bp upstream T7 promoter sequence; 1-4 G residues that initiate RNA from the T7 promoter; and a downstream minimal lac operator that is linked to the XbaI site that begins the TIER. The link between TPCR and TIER is 0-4 bp long or through a single base-pair overlap between the last base pair of the lac operator and first base pair of the XbaI site (Tables 1 and 3). A second lac operator upstream of the T7 promoter in some pREX and pAL vectors begins immediately after a SacII site and comprises a 37-bp lacO1 operator with its natural extended palindrome positioned ahead of the BsiWI site at or near the optimal distance from the downstream lac operator for repression of transcription, as determined by Muller et al. (1996) J. Mol. Biol. 257:21-29.

In certain embodiments, the cgtacgaaat sequence at the 5' terminus of the TPCR may be substituted with another restriction enzyme site, no restriction site, or a nucleic acid sequence of identical length that does not contain a restriction site.

Modifications to the terminal sequence of the TIER have been contemplated. For example, the XbaI sequence (TCTAGA) and NcoI sequence (CATG) of the TIER may be modified. In particular, the XbaI sequence can be replaced with another restriction enzyme site or no restriction enzyme site at all. However, in certain embodiments, it is important to maintain a sequence of six nucleic acid residues at the 5' terminal end of the TIER region polynucleotide.

Combinations of T7 Promoter Control Region and Translation Initiation Efficiency Region have been contemplated. Any T7 Promoter Control Region may be paired with any Translation Initiation Efficiency Region.

Different elements in the Expression Control Region and short linker sequences between them have been the primary focus of testing and modification to reduce basal expression and improve the capabilities of pREX vectors and the pAL vectors derived from them. Five types of variation were evaluated (Tables 1-3): 1) initiating target mRNA with 1, 2, 3 or 4 consecutive G residues ahead of a minimal lac operator; 2) the relative effectiveness of a downstream asymmetric lacO1 or perfectly symmetric lacS minimal operator; 3) different lengths and compositions of base pairs linking the downstream lac minimal operator with the Translation Initiation Efficiency Region; 4) different lengths and compositions of base pairs within the TIER; and 5) adding the 37-bp lacO1 and extended palindrome upstream of the T7 promoter at or near the optimal distance for repression. Effects of these targeted variations are disclosed in following sections.

The TIER in most pREX vectors disclosed herein is tg10 (Table 1 and SEQ ID NO. 48) and pREX vectors having tg10 may be identified simply by their TPCR designation, e.g. LS31 (designations are defined in Table 3). Other TIERs

investigated are given in Table 2, and pREX vectors may be identified specifically by a compound TPCR-TIER designation to certify which TIER it contains, e.g. S31-tr5 or S31-tg10.

lacIt Gene in pREX and pAL Vectors

The lacI gene in pET vectors is meant to supply enough lac repressor to saturate all of the lac operators in an uninduced T7 expression strain such as BL21(DE3) in which bound lac repressor blocks expression of T7 RNA polymerase and the lac operon by *E. coli* RNA polymerase from different lac promoters in the chromosome and also blocks expression of target protein by T7 RNA polymerase from a T7lac promoter in the multi-copy vectors. However, transcription and translation-initiation signals for producing lac repressor from the lacI gene are known to be relatively weak, and it seemed likely that supplying a higher concentration of lac repressor in the cell might increase occupancy of the lac operators and thereby reduce basal expression of both T7 RNA polymerase and target protein with little effect on induced expression. Accordingly, two modifications to the sequence upstream of the lacI gene (SEQ ID NO. 60) were made in a pREX L44 vector, which has an Expression Control Region equivalent to that of pET-11d (Table 1 and SEQ ID NOs. 10 and 13): 1) introduction of the lacI^q mutation in the promoter for expressing lacI, a single-bp change in the -35 region (GCGCAA to GtGCAA) that is known to increase transcription of lacI by *E. coli* RNA polymerase (Calos (1978) Nature 274:762-765); and 2) changing the initiation codon for lac repressor from GTG to ATG and exchanging the upstream 13 base pairs with an AT-rich sequence with an appropriately spaced Shine-Dalgarno (SD) sequence to increase translation of lacI mRNA, a modification designated lacIt (SEQ ID NO. 61). A pREX vector containing both modifications was found to be unstable, presumably because the concentration of lac repressor was high enough to stress the cell. However, lacIt without the lacI^q mutation is well tolerated in pREX vectors and production of T7 10a protein after induction is comparable to that obtained in pET vectors. Therefore, lacIt has been included in pREX and pAL vectors rather than the natural sequence upstream of the lacI coding sequence, in the expectation that it generates a somewhat higher concentration of lac repressor and greater occupancy of lac operators in the uninduced cell, thereby reducing basal expression of both T7 RNA polymerase and the target protein more effectively.

During initial construction of pREX vectors, the lacIt gene was oriented so as to be transcribed in the same direction as the target gene. After changing the orientation, it was found that the unintended lacIt orientation by itself prevented establishment of a 5.3 clone that could be maintained in an otherwise identical vector having lacIt in the intended orientation. Apparently, a significant fraction of *E. coli* RNA polymerases that initiate transcription at the weak lacI promoter continue past the t1t2 double transcription terminator and through the uninduced Expression Control Region to produce functional target mRNA. The intended lacIt orientation (transcribed in the opposite direction from the target gene) is standard in pREX and pAL vectors. The t1t2 termination module has been retained but could possibly be eliminated with little consequence.

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EXAMPLES

The present invention is illustrated in further details by the following non-limiting examples.

Testing and Improvement of pREX Vectors

The host strain used for initial cloning has been XL1Blue-MR, referred to here as XL1B (from Stratagene, now Agilent Technologies, Inc.), which lacks known DNA restriction systems, requires thiamine for growth, and has no gene to supply T7 RNA polymerase. The hosts used for expression under control of inducible T7 RNA polymerase are BL21(DE3) (from Novagen, now EMD Millipore) and BL21-AI (from Invitrogen, now Life Technologies, part of Thermo Fisher Scientific) as well as newly isolated variants of BL21(DE3) described in a later section

The first pREX vector tested had the Expression Control Region L44-tg10, equivalent to that of pET-11d (Table 1). This vector produced high levels of T7 gene 10a capsid protein but was unable to maintain the gene 5.3 coding sequence received from pREP even in XL1B (Tables 2 and 3). Since XL1B does not supply T7 RNA polymerase, inability to establish a functional 5.3 clone implies that the target gene is somehow being transcribed by *E. coli* RNA polymerase, which can initiate transcription at a wide range of sequences. It seemed likely that one or more weak promoters for *E. coli* RNA polymerase upstream of the target coding sequence was directing production of enough of the highly toxic 5.3 protein that clones could not be tolerated.

Examination of the tg10 sequence identified two sequences, TAattT and TAactT (underlined in Table 2), that contain the most highly conserved base-pairs in the TATAAT consensus sequence for the -10 region of promoters for *E. coli* RNA polymerase, either of which would direct transcription of target mRNA. Expecting that eliminating or altering one or both of these sequences might reduce basal expression sufficiently to allow stable maintenance of clones capable of producing functional 5.3 protein, deletions and alterations between the XbaI site and the Shine-Dalgarno (SD) sequence of pREX L44-tg10 were constructed and tested (Table 2). As a control, the entire Translation Initiation Efficiency Region between the XbaI and NcoI sites was replaced with the equivalent fragment from pREP-1, which had been designed to be unfavorable for binding ribosomes and initiating target protein synthesis (tr3 in Table 2).

As summarized in Table 2, the 5.3 coding sequence could be cloned in pREX L44-tr3 and maintained with little evidence of stress in XL1B and both T7 expression hosts BL21(DE3) and BL21-AI. Other alterations to tg10 in L44 also enabled 5.3 clones to be established in XL1B, although usually at lower efficiency and with smaller colony sizes than 10a clones, indicating that some basal expression directed by *E. coli* RNA polymerase remained in most or all of them. These clones could also be maintained in BL21-AI but were more stressful to varying degrees in BL21(DE3), consistent with a higher basal level of T7 RNA polymerase. Induction of clones in BL21-AI stopped the growth of the culture, indicating that active 5.3 protein was produced; however, not enough protein accumulated to be apparent in a stained gel after SDS-electrophoresis of whole-cell extracts.

Controlling Rates of Translation Initiation

Each of the above pREX L44 vectors having a variant TIER was also tested for ability to produce 10a protein in

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BL21(DE3). As expected, the tr3 replacement of essentially the entire tg10 TIER with a sequence that lacks recognizable translation initiation signals reduces production of 10a protein to a level barely if at all detectable in stained electrophoresis gels

(Table 2). Unexpectedly, L44-tr1, which retains the entire Shine-Dalgarno (SD) sequence through the initiation codon but only 5 of the 28 base pairs of tg10 upstream of the SD sequence, produced only barely detectable amounts of 10a protein. Interestingly, production of 10a protein increases gradually with increasing retention of tg10 sequence upstream of the SD sequence and reaches levels typical of L44-tg10 when ~15 or more base pairs are retained (Table 2).

Natural T7 gene 10 mRNA begins with a 21-bp stem-loop structure that ends immediately ahead of the XbaI site that begins tg10 and continues with a 28-nucleotide (nt) largely unstructured AT-rich sequence ahead of the SD sequence AAGGAG (Table 1). Target mRNA from L44 vectors also begins with a large stem-loop structure that ends immediately ahead of the XbaI site, in this case a 29-bp interrupted palindrome that encompasses the lacO1 operator. A likely explanation for the reduction of protein production caused by deletions between the XbaI site and SD sequence of the tg10 TIER is that positioning a rather large and stable RNA structure too close to the SD sequence interferes with binding of the mRNA to the ribosome and reduces the frequency of initiation of synthesis of target protein. This apparent ability to control rate of translation simply by changing the distance between a relatively stable RNA structure and an intrinsically strong, relatively unstructured upstream protein-initiation sequence may prove useful, as the solubility of some target proteins has been reported to increase when rates of protein production are reduced. The same strategy should also be applicable to the 20-bp completely symmetrical palindrome in the lacS operator in T7lac promoters constructed subsequent to these experiments (Table 1).

Vectors that Reduce Basal Expression by T7 RNA Polymerase and *E. coli* RNA Polymerase

Although deletions in tg10 allowed pREX clones of the coding sequence for the highly toxic 5.3 protein of T7 to be maintained and expressed in BL21-AI, most of these clones could be established only with considerable difficulty if at all in BL21(DE3) (Table 2). Therefore, changes to other elements in the Expression Control Region of pREX vectors were tested for ability to reduce basal transcription of the target gene by the somewhat higher basal levels of T7 RNA polymerase in BL21(DE3) and thereby increase the stability and range of clones that can be established and expressed in BL21(DE3) or any inducible T7 expression host. Surprisingly, some combinations of modifications that do not affect tg10 at all reduce basal transcription of target gene not only by T7 RNA polymerase but also by *E. coli* RNA polymerase to levels where pREX or pAL clones of the 5.3 coding sequence and every other coding sequence tested, many of which specify proteins highly toxic to the host, can be maintained and expressed in BL21(DE3).

The approach was to try to enhance the ability of bound lac repressor to block transcription of the target gene by T7 RNA polymerase. The frequency of T7 RNA polymerase transcription through an operator saturated with specifically bound lac repressor is known to increase with distance between the T7 promoter and operator (Dubendorff et al. (1991) and Giordano et al. (1989)), so decreasing the

distance between T7 promoter and lac operator might be expected to decrease basal transcription of target gene by T7 RNA polymerase. The T7lac promoter in pET vectors initiates RNA chains 4 bp ahead of the AATT sequence that begins the 21-bp interrupted palindrome that constitutes the minimal natural lacO1 operator (pET-11d and pREX L44-tg10 in Table 1). The RNA produced by T7 RNA polymerase begins GGGGAA, a sequence very similar to the conserved GGGAGA start of the RNAs from the six strongest T7 promoters in T7 DNA. Addition of CCCC residues at the other end of the T7lac operator extends the palindrome by 4 perfect base pairs, similar to extensions of the palindrome flanking the natural lacO1 operator (upstream 37-bp lacO1 in Table 1).

Three approaches to decreasing basal transcription of the target gene by T7 RNA polymerase were taken: 1) exploring effects on basal and induced expression of moving the lac operator closer to the RNA start position; 2) replacing the 21-bp natural lacO1 operator with a 20-bp operator that is a perfect palindrome (referred to here as lacS), which is known to bind lac repressor with a higher affinity (Muller et al. (1996) J. Mol. Biol. 257:21-29); and 3) adding a 37-bp lacO1 operator with its extended natural palindrome upstream of the T7 promoter-lacS sequence such that the center-to-center spacing between the two operators is 70.5 bp for T7 promoters that begin RNA with GGGAA, the spacing expected for maximum repression (Muller et al. (1996)). For convenience in vector construction, the spacing is 69.5 bp for T7 promoters that begin RNA with GGAA, which seems to have comparable effect. The names, nucleotide sequences and SEQ ID NOS. of different T7 Promoter Control Regions and Translation Initiation Efficiency Regions that comprise Expression Control Regions of pREX and pAL vectors are given in Tables 1-3, and results of cloning and expression tests are given in Tables 2 and 3.

Initial experiments to test the effect of moving the lacO1 operator closer to the T7 promoter by progressive shortening of the four consecutive G residues that begin the RNA produced from the T7lac promoter used in pET vectors also shortened the four consecutive C residues between the lac operator and tg10, so the observed improvements in ability to clone the 5.3 coding sequence were probably due primarily to shortening the linker between the operator and tg10, as discussed in more detail below (Table 3). Shortening the RNA start from GGGGAA to GGGAA had no apparent effect on levels of 10a protein produced after induction in either BL21(DE3) or BL21-AI; shortening to GGAA had little if any effect on level of 10a protein produced in BL21(DE3) but moderately reduced the level produced in BL21-AI; and shortening to GAA severely reduced 10a protein production in both hosts. All of the natural T7 promoters in T7 DNA begin RNA with at least two of the first three nucleotides being G, and apparently a GAA start is unfavorable. In one embodiment, the RNA start includes GGGGAA, GGGAA, GGAA, GGAGAA, GAGGAA, GAGAA, AGGAA, or AGGGAA. Examples of T7 Promoter Control Regions having the afore-mentioned RNA start sequences are disclosed herein and provided, for example, in SEQ ID NOS. 13-32 and 94-102.

T7lacS promoter-operator combinations appeared to be superior to T7lacO1 combinations for maintaining and expressing 5.3 clones in all configurations in which they were compared, and expression levels of 10a protein after induction were comparable. Therefore, the symmetric lacS operator was used in most tests of effects of moving the lac operator closer to the T7 promoter and shortening the sequence between the AATT that ends the lacS operator and

the XbaI recognition sequence TCTAGA that begins tg10 (Table 3). Substituting a G residue for one or both of the A residues that begin the minimal lac operator sequence decreased the ability to maintain and express 5.3 clones (not shown in Table 3), presumably due to weaker binding of lac repressor to the altered operator sequence.

The length of consecutive C residues separating the lac operator from tg10 was found to have a substantial effect on the level of basal expression of the target gene due to transcription by *E. coli* RNA polymerase. Vectors linking these two regions through CCCC were unable to clone the 5.3 coding sequence in XL1B regardless of changes in spacing between the T7 promoter and lac operator or the presence of a second operator upstream of the T7 promoter (Table 3). This configuration, also found in many pET vectors and in vectors described by Kara et al. (U.S. Pat. No. 6,537,779), is apparently unsuitable for cloning and expressing a considerable range of proteins that stress *E. coli* expression hosts. Changing the linker between lacS and tg10 to AGGG gave marginal improvement in some configurations but was generally unsatisfactory as well (not shown in Table 3). Shortening CCCC to CCC or CC without changing the number of G residues that begin the RNA improves ability to clone 5.3 in XL1B, although cells are obviously stressed in some configurations, and transfer to BL21(DE3) or BL21-AI for expression can remain problematic or impossible (Table 3). The best configurations for cloning 5.3 in XL1B and for transferring clones to and expressing them in hosts that supply T7 RNA polymerase are when the linkage between lacS and tg10 consists of a single C, no base-pair at all, or where the lacS operator and XbaI site overlap by a single T. Presumably, replacing the single C with an A, G, or T residue at this position would have a similar effect. Reducing basal expression due to transcription of the target gene by *E. coli* RNA polymerase in this way is superior to deleting potential promoter elements in tg10 (compare Tables 2 and 3).

Simply moving the lacS operator one or two bp closer to the T7 promoter than the 4-bp distance in the T7lac promoter appears to have had minimal effect on decreasing basal transcription by T7 RNA polymerase (Table 3). However, placing a lacO1 operator with its natural extended palindrome upstream of the T7 promoter sequence at a spacing between the lacO1 and lacS operators close to that reported to provide maximum repression (Muller et al. (1996) J. Mol. Biol. 257:21-29) substantially improves the ability to clone, maintain and express the 5.3 coding sequence. This enhancement could be due to the ability of tetramer lac repressor to bring the two operators together, forming a DNA loop that interferes with binding of T7 RNA polymerase, or an increase in occupancy of the lacS repressor-binding site, or both.

Controlling Rates of Target Protein Production

Induced levels of 10a protein in BL21(DE3) are comparably high for all vector configurations in which target mRNA begins with GG, GGG, or GGGG and whether or not an upstream operator is present (Table 3). However, at least three different levels of induced expression are apparent in BL21-AI, depending on these factors (Table 3). The lower levels of production in BL21-AI seem likely to be due to a lower induced level of T7 RNA polymerase in BL21-AI.

Controlling rates of production of target protein by combining different T7 Promoter Control Regions and Translation Initiation Affinity Regions characterized in Tables 1-3 in different vectors and hosts may be useful in producing

functional proteins if significant numbers of proteins have increased solubility or are better able to fold correctly at lower rates of transcription or translation, as reported for some proteins. The ability to control basal and induced transcription by T7 RNA polymerase or other T7-like RNA polymerases, combined with cell-specific translation signals might also have application in producing proteins in other types of cells or for synthetic biology

Asymmetric Ligation and Cloning in pAL Vectors

Although the pREP-pREX system has been useful both for testing different vector configurations and for producing proteins, a simpler cloning system for efficient and high-throughput cloning of coding sequences in vectors that incorporate the improvements developed in pREX vectors has been designed and implemented. The method is referred to here as asymmetric ligation and the vectors for cloning by asymmetric ligation are called pAL vectors.

Many cloning vectors, including pREP and pREX vectors, have directional cloning sites generated by cutting with restriction endonucleases that produce a different symmetric (self-complementary) overhang at each end of the cloning-acceptor fragment. These cleavages produce 3' hydroxyl and 5' phosphate ends that are substrates for covalent joining by a DNA ligase such as T4 DNA ligase, which efficiently joins double-stranded DNAs that have the same symmetric overhang (such as 5'-CATG or TA-3' in pREP and pREX vectors) by sealing the nick at each end of perfectly base-paired overhangs. In a ligation reaction mixture, the equilibrium between base-paired and unpaired symmetric overhangs greatly favors unpaired overhangs, but the rate of ligation increases with ligase concentration and, under appropriate conditions, T4 DNA ligase can join both nicks in essentially all perfectly paired 4-nt overhangs in less than a minute at room temperature (Shore et al. (1981) *Proc. Natl. Acad. Sci. USA* 78:4833-4837).

Directional cloning by ligation of symmetric overhangs requires two steps: 1) an initial ligation to join one end of a cloning-acceptor fragment to the complementary end of a target fragment through perfectly paired symmetric overhangs, and 2) a subsequent ligation to join the other ends through their perfectly paired symmetric overhangs to complete the circular clone before either end can be joined through perfectly paired overhangs to another linear fragment. Ungapped perfectly paired symmetric overhangs direct efficient ligation between any linear molecules in the reaction mixture indiscriminately, and the many competing nonproductive reactions mean that the desired clone is typically a minor component of ligation products. Nevertheless, the desired pREP or pREX clone can usually be obtained simply by transformation of the ligation mixture, because strong selection for antibiotic resistance and counter-selection against the *ccdB* module will eliminate most of the unwanted clones.

Ligases have long been known to join double-stranded DNAs at ungapped perfectly base-paired overhangs much more rapidly than where imperfectly aligned overhangs have gaps or mismatches at or near the ends to be joined (e.g. Wu et al. (1989) *Gene* 76:245-254 for T4 DNA ligase). Asymmetric ligation takes advantage of this specificity to greatly increase the efficiency of directional cloning. Asymmetric overhangs of all DNAs in the ligase reaction mixture are designed so that the only ungapped perfectly base-paired overhang alignments are those whose ligation will produce the desired clone or regenerate the cloning vector and, crucially, such that all of the other possible overhang align-

ments have few if any aligned base pairs near their ends and therefore a much smaller probability of being ligated. In the pAL vectors constructed so far, overhangs in the cloning-acceptor fragment are generated by the Type IIS restriction endonuclease *BsaI*, which cuts to one side of an asymmetric recognition site to produce 4-nt 5' overhangs that can have any sequence. The only two *BsaI* recognition sites in these vectors flank the same *ccdB* counter-selection module used in pREP and pREX vectors and are oriented outward, so that cutting with *BsaI* produces a cloning-acceptor fragment with the desired asymmetric overhang at each end and a *ccdB*-containing fragment that contains both *BsaI* recognition sequences and has asymmetric overhangs complementary to those of the cloning-acceptor fragment (map of pAL1 in FIG. 4). Of course, any Type IIS restriction endonucleases that cut outside of their recognition sequence to produce 4-nt 5', 3-nt 5' or possibly even 2-nt 5' or 2-nt 3' overhangs of arbitrary sequence would also be suitable for generating asymmetric overhangs for cloning. For example, restriction endonucleases that provide a 4-nt 5' overhang include *BsaI*, *BbvI*, *BcoDI*, *BsmFI*, *BsmBI*, *FokI*, *BfuAI*, *BbsI*, and *SfaNI*; restriction endonucleases that provide 3-nt 5' overhang include *SapI*; 2-nt 5' overhang include *FauI*; and 2-nt 3' include *BseRI*.

To clone a single DNA in a pAL vector by asymmetric ligation, the target DNA must have the asymmetric 4-nt 5' overhang at each end that is perfectly complementary to the asymmetric 4-nt 5' overhang at the appropriate end of the cloning-acceptor fragment of the vector. The overhangs in a target DNA are routinely generated in a 5-minute reaction at room temperature in which the 3' exonuclease activity of T4 DNA polymerase removes the first four nucleotides at each 3' end of the double-stranded target DNA but stops at the fifth nucleotide because the only deoxynucleotide triphosphate (dNTP) present in the reaction mixture can be incorporated at this position by the polymerase activity, thereby preventing removal of additional nucleotides. The target DNA being digested has typically been amplified by PCR with a high-fidelity polymerase that produces blunt ends, using two PCR primers that have the appropriate five-nucleotide sequence at each 5' end, and the T4 DNase polymerase reaction mixture also contains T4 polynucleotide kinase to phosphorylate the 5' ends of the overhangs to make them substrates for ligation. Any target DNA that can be amplified by PCR can be cloned in this way regardless of whether it contains (or lacks) a site for cleavage by certain restriction endonucleases. Fragments suitable for cloning could also be produced by Type IIS cleavages at the ends of PCR amplified DNA, but only if the enzyme does not also cut within the sequence to be cloned.

A limitation in the use of T4 DNA polymerase in the presence of a single dNTP to generate overhangs for cloning is that none of the four nucleotides in an overhang should be the same as the fifth nucleotide from the 5' end. However, control tests of 5-minute reactions at room temperature with T4 DNA polymerase in the absence of any dNTP but in the presence of T4 polynucleotide kinase (to phosphorylate the 5' ends of overhangs) found that ligation of DNA treated in this way to *BsaI*-cut pAL1 produced ~20-50% as many clones as obtained when the target DNA had been digested in the presence of the appropriate single dNTP. These few control tests suggest that the limitation on composition of the overhang and requirement for a specific nucleotide at the fifth position from the 5' end could probably be bypassed if necessary or desirable in selecting pairs of 4-nt overhangs for cloning by asymmetric ligation, and perhaps other exonucleases could also be used to generate overhangs. How-

ever, so many different pairs of asymmetric 4-nt 5' overhangs are suitable for cloning by asymmetric ligation that the limitations of digestion by T4 DNA polymerase are relatively insignificant and these additional possibilities were not pursued. Chemical synthesis of DNA is becoming progressively cheaper and more accessible so that simply purchasing double-stranded DNA with a codon-optimized coding sequence and any specified overhang sequences is a feasible alternative.

The initial pAL vectors were designed to produce the target protein itself or fused to an N- and/or C-terminal peptide or protein domain, and a variety of Expression Control Regions and fusion partners are represented among pAL vectors already constructed and tested (Table 4 and SEQ ID NOs. 66-78). Three pairs of upstream and downstream overhangs of target DNA generated by T4 DNA polymerase are cloned by asymmetric ligation in different subsets of these vectors: CCATg-cAGTG and ACTCg-cAGTG are generated in the presence of dCTP, and CTGCa-tAGCG are generated in the presence of dTTP. Sequences are given in the 5' to 3' orientation left to right with the four overhang nucleotides capitalized; consequently, the 5' end of the upstream PCR primer for amplifying a target coding sequence is the five-nucleotide sequence given, and the 5' end of the downstream PCR primer is the complement of the five-nucleotide sequence given.

The pattern of base pairing in each of the 10 possible ungapped pairwise alignments between the four different 4-nt overhangs present in an asymmetric ligation reaction for cloning a single coding sequence are shown in FIGS. 5A-5C as a base pair matrix for each of the three pairs of overhangs. The two pairs of complementary overhangs that produce the desired clone each have a standard complementary DNA base-pair at all four positions of the alignment, the best substrate for ligation (represented as 1111 in the three base-pairing matrices illustrated in FIGS. 5A-5C). The remaining eight pairwise alignments have at most two of the four positions being either a standard base-pair (1001, 1010, or 0110) or the presumably much weaker G-T base-pair (indicated by a dash in FIGS. 5A-5C, i.e., 0-0 or -00-), which makes these eight overhang pairs much worse substrates for ligation. None of the imperfectly matched pairs of overhangs has more than 1 consecutive standard base-pair at either end. Also shown in FIGS. 5A-5C are the possible combinations of two or three amino acids to link upstream and downstream peptides or protein domains to the target protein in each of the three reading frames.

Cloning by asymmetric ligation in these pAL vectors is so efficient that transformation of 1 µl of a 5-minute room-temperature ligation reaction mixture typically produces tens of thousands of clones in XL1B or thousands in BL21(DE3) or BL21-AI, almost all of which are the desired clone. This is ~1-2 orders of magnitude more colonies than were typical in cloning by ligation of the symmetric overhangs in pREP or pREX vectors. Long coding sequences are also cloned efficiently by asymmetric ligation; the 4239-bp coding sequence for a 1413 amino-acid transporter cloned in pAL1 produced too many transformed colonies to count, and the six clones analyzed each contained the entire coding sequence. The presence of the ccdB counter-selection module in pAL vectors allows efficient cloning directly in BsaI-cut pAL vectors without having to purify the cloning-acceptor fragment. In typical ligation reactions, the cut vector and the fragment to be cloned are at equi-molar concentration. The overhangs of the cloning-acceptor fragment will pair perfectly for ligation only to the overhangs of the target fragment that produce the desired clone or to the

overhangs of the ccdB fragment to regenerate the vector. Since the ccdB toxin prevents growth of cells containing incompletely cut or reconstituted vector, the vast majority of transformed colonies that grow in the presence of kanamycin (the selective antibiotic for pAL vectors) contain the desired clone. As with pREP and pREX vectors, pAL vectors must be maintained in a host such as DB3.1, which is resistant to the effects of the ccdB toxin.

Deriving pAL Vectors from pREX Vectors

The modular construction of pREX vectors made it relatively easy to derive the first pAL vectors from pREX S31-tg10 (FIG. 3 and SEQ ID NO. 62) to produce pAL1, 5, 11-14 and 21-24 (Table 4, which includes SEQ ID NOs.). The DNA fragments between the XbaI-AgeI and EagI-Acc65I sites flanking the ccdB counter-selection module were replaced with synthetic oligonucleotides or appropriately processed PCR products to: 1) introduce unique outward facing BsaI sites flanking the ccdB module (FIG. 4); 2) provide asymmetric overhangs to link target coding sequences to Expression Control Regions containing either the tg10 or tr5 Translation Initiation Efficiency Region, which provide different rates of translation (Table 2); and 3) specify N-terminal and/or C-terminal fusions to the target protein (Table 4). In these pAL vectors, direct linkage of the target coding sequence to the ATG initiation codon at the end of the Expression Control Region is through the CCATg overhang, which allows complete flexibility for all remaining codons that specify the target protein (Tables 1 and 4). The ATG initiation codon for N-terminal fusions to the target protein is in a CATATG NdeI site at the end of the Expression Control Region, as in T7 DNA (Tables 1 and 4).

N-terminal fusions are inflexible in an individual vector but C-terminal fusions are optional because the coding sequence to be cloned can either include a termination codon ahead of the overhang that joins it to the vector or can link in-frame through the overhang to the sequence encoding a C-terminal fusion. Examples of N-terminal and C-terminal fusions have been contemplated and include affinity tag (HIS, FLAG, Myc, or HA), fluorescent tag (GFP and CFP); targeting peptide (pelB or dsbA); and dimerization domain (dsbC).

The N-terminal fusions represented in the vectors listed in Table 4 are a Met-Ala-Ser-6His-Ser affinity tag and pelB or dsbA leader sequences that target proteins to the periplasm by the Sec or SRP pathway respectively. The optional C-terminal fusions represented are a Ser-Gly-6His affinity tag and a 73-amino-acid dimerization domain of the *E. coli* dsbC protein (a periplasmic disulfide isomerase) with or without an additional Gly-Ser-6His affinity tag.

The collection of pREX vectors already tested for ability to maintain and express target coding sequences is a valuable resource for constructing useful pAL vectors. Modular construction makes it easy to convert any pREX vector to a pAL vector simply by replacing the XbaI-Acc65I or NcoI-Acc65I fragment that contains the ccdB counter-selection module with the comparable fragment from any pAL vector (see, for example, FIGS. 3 and 4, and Tables 1-4). The pAL1 vector derived from pREX S31-tg10 has proved to be highly versatile for cloning and testing expression of a variety of coding sequences with or without a C-terminal 6His affinity tag. Anticipating that the somewhat lower basal and induced expression levels accessible with the Expression Control Regions of the S21-tg10 (SEQ ID NO. 63), LS31-tg10 (SEQ ID NO. 64) and LS21-tg10 (SEQ ID NO. 65) pREX vectors will be advantageous for some target proteins, the pAL2

(SEQ ID NO. 67), pAL3 (SEQ ID NO. 68) and pAL4 (SEQ ID NO. 69) vectors (Table 4) were derived from these pREX vectors by replacing the NcoI-Acc65I fragments of the pREX vectors with the comparable fragment of pAL1. Exchange of Translation Initiation Efficiency Regions or introduction of specific N- or C-terminal fusions into pAL vectors is easily done by taking advantage of the unique SacII, BsiWI, XbaI, NcoI, AgeI, EagI, and Acc65I sites in pAL vectors (Table 1 and FIG. 4).

Cloning and Co-Expressing More than One Coding Sequence in pAL Vectors

Ability to co-express more than one coding sequence from a single vector promises to be useful for producing functional protein complexes that contain more than one protein, especially when individual proteins are unstable or insoluble in the absence of their interacting partner(s).

Two or three coding sequences having appropriate overhangs can be directionally cloned routinely in a single asymmetric ligation reaction for co-expression in a pAL vector. Four coding sequences for co-expression and four fragments linked to make a single coding sequence have also been cloned in pAL1, but with some difficulty.

Coding sequences for T7 late proteins transcribed by T7 RNA polymerase during infection are translated efficiently from relatively stable mRNAs that contain several different coding sequences in tandem (Dunn et al. (1983) J. Mol. Biol. 166:477-535). The relative stability of T7 mRNAs is attributed primarily to stable stem-loop structures at their 3' ends, one of which is generated by termination at the Tphi transcription terminator in T7 DNA, which terminates transcription of target DNA in pAL vectors (and in pREX and many pET vectors). Each of the coding sequences in these multi-gene T7 mRNAs almost always has its own translation-initiation region with a strong SD sequence such as GGAG, GAGG or GGAGG separated from an ATG initiation codon by an AT-rich sequence ~5-9 nucleotides long containing few G residues. Therefore, sequences that link coding sequences for co-expression in pAL vectors have been designed to have these features.

To test cloning efficiency and co-expression as a function of length of the AT-rich region, coding sequences for the T7 gene 9 and 10a proteins were joined to each other through asymmetric overhangs of length 4 to 10 nucleotides containing only A and T residues and directionally cloned in single asymmetric ligation reactions for co-expression in pAL1. The (A,T) overhangs were designed to minimize complementary base pairs at or near the ends of all eight or more possible pairwise alignments outside of the three perfectly complementary alignments that produce the desired clone. Overhangs for joining the two coding sequences as well as overhangs for joining them to the pAL1 cloning-acceptor fragment were generated by treatment of the two PCR products with T4 DNA polymerase in the presence of dCTP. The sequences taaggagc (A, T)_n and the complement of _n(A, T) gatg were joined by ligation to form the junction taaggagc_n(A, T)_ngatg, where the underlined taa is the termination codon of the first coding sequence, the underlined atg is the initiation codon of the second coding sequence, and _n(A, T)_n represents the complementary asymmetric overhangs of different lengths at the 5' end of one PCR primer for amplifying each coding sequence (with the overhang for ligation to the cloning-acceptor fragment of pAL1 being generated at the other end). Each asymmetric ligation reaction produced hundreds of clones in XL1B, almost all of them correct, and auto-induction in BL21(DE3)

produced comparable amounts of the two proteins in each of the seven clones. Therefore, a strategy of supplying a strong SD sequence joined to an ATG initiation codon through AT-rich overhangs between 4 and 10 nt long appears to work well for cloning and co-expression if nonproductive ligations are sufficiently minimized.

Cloning two or three coding sequences by asymmetric ligation for co-expression from pAL1 typically yields hundreds of colonies, almost all of which are correct. The usual junction between two coding sequences has been taaggagacTATTAATg (SEQ ID NO. 79), where the termination codon of the first coding sequence and the initiation codon of the second coding sequence are underlined. The 7-nt asymmetric sequence capitalized and its complement are the linker 5' overhangs generated by T4 DNA polymerase in the presence of dCTP from the PCR products to be joined. The junction between the second and third coding sequences has usually been taaggagacTTAATATg (SEQ ID NO. 80). The base-pairing patterns for all possible pairwise alignments of the overhangs present in cloning two or three coding sequences in pAL1 by asymmetric ligation using these overhangs are shown in FIGS. 6A-6B. The three or four ungapped, perfectly base-paired overhangs that can be efficiently ligated to produce the desired clone are represented as 1111, 1111111, and 1111 for cloning two coding sequences and 1111, 1111111, 1111111, and 1111 for cloning three coding sequences; and are circled in FIGS. 6A-6B. The 26 or 48 mismatched aligned pairs apparently are poor enough substrates for ligation that undesired ligation products do not significantly interfere with ability to obtain the desired clone.

To test possible position effects on expression levels, the T7 gene 8, 9, and 10a proteins were cloned for co-expression in pAL1 in all six possible gene orders, using the above two junction sequences. The three proteins in each of the gene orders were readily apparent in a stained gel after SDS-electrophoresis of whole-cell extracts of auto-induced cultures. In general, each protein was produced at a distinctly higher level when its coding sequence was in the first position to be transcribed relative to the second position, with perhaps a further slight decrease in the third position.

In attempts to extend simultaneous cloning by asymmetric ligation to four DNA fragments in pAL1, cloning efficiency was greatly reduced and only a small fraction of the clones were correct. This drastic reduction in success rate probably results from a combination of the increasing ratio of imperfectly to perfectly aligned overhangs and an unavoidable increase in lengths of perfectly paired bases at the ungapped ends of imperfectly aligned overhangs as the total number of overhangs increases. Nevertheless, one set of four coding sequences was cloned by asymmetric ligation for co-expression and another set of four DNA fragments was correctly linked to produce a single coding sequence for a multi-domain protein of 4832 amino acids. It may be possible to increase efficiency sufficiently to support routine cloning and co-expression of four or more coding sequences by using all four nucleotides in the overhangs that join coding sequences (for example, by using de novo synthesized DNAs) or by increasing the stringency of ligase for perfectly paired bases. Increasing the salt concentration to 200 mM or adding 5 mM spermidine to the ligation reaction mixture was reported by Wu et al. (1989) Gene 76:245-254 to increase the specificity of T4 ligase for perfectly-paired nicks relative to imperfectly paired sequences by 10-fold or

more. Other ligases might also have greater specificity. These potential simple remedies have yet to be explored.

Variants of BL21(DE3) with Reduced Basal Expression

In the course of testing the ability of different pREX vectors to maintain and express clones of T7 gene 5.3 in BL21(DE3), several different 5.3 mutants were isolated from larger colonies that arose on plates used to titer populations stressed by basal expression. Analysis of large colonies from a set of plates that had mistakenly lacked the selective antibiotic found that a significant fraction had lost the plasmid, some retained expression clones having mutations that reduced expression or produced impaired 5.3 protein, and some retained apparently unaffected expression plasmid and therefore seemed likely to be variants of BL21(DE3) that could better tolerate 5.3 expression clones. Some white sectors of growth in colonies that were dying on LB plates because of unintended induction at saturation also gave rise to such variants. Potential variants of BL21(DE3) were readily isolated free of their expression plasmid simply by growing colonies on plates that lack the selective antibiotic and contain the inducer IPTG. Candidates were then tested for ability to support plaque formation by the T7 deletion mutant 4107, which lacks the entire gene for T7 RNA polymerase and cannot form a plaque unless active T7 RNA polymerase is provided by the host cell (Studier et al. (1986)). Plaque formation and size in the absence or presence of inducer are indicators of relative amounts of basal and induced T7 RNA polymerase supplied by the host cell. BL21(DE3) itself gave small plaques in the absence of inducer and large plaques in its presence. This plaque assay identified variants of BL21(DE3) that gave even smaller plaques or no detectable plaques in the absence of inducer and a range of plaque sizes in the presence of inducer, from no detectable plaques to large plaques comparable to those obtained with BL21(DE3) itself.

The upstream region controlling expression of T7 RNA polymerase from the chromosome of BL21(DE3) extends from the end of the *lacI* gene to the beginning of the coding sequence for T7 RNA polymerase (SEQ ID NO. 81) and contains: the L8 mutation, a G to A transition in the binding site for the cAMP receptor protein (CRP); the GT to AA double mutation that converts the -10 region of the *lac* promoter from the wild-type TATgtT to the stronger -10 consensus sequence TATAAT of the *lacUV5* promoter; the *lacO1* operator; the coding sequence for an N-terminal fragment of *lacZ*; and 26 base pairs of T7 DNA ahead of the ATG initiation codon for T7 RNA polymerase. The only sequence difference between *E. coli* K12, the source of this control region in BL21(DE3), and the same region of the *lac* operon in BL21(DE3) is the last base pair ahead of the minimal *lacO1* operator (the sixth base pair downstream of the -10 sequence of the *lac* promoter), where K12 has a G and BL21(DE3) an A. It seemed likely that at least some of the BL21(DE3) variants would have changes in this upstream control region, in the coding sequence for T7 RNA polymerase, or both.

The entire sequence of the upstream control region between the end of *lacI* and the beginning of T7 RNA polymerase was sequenced in a set of 22 of the BL21(DE3) variants identified as large-colony variants that arose under stress by a 5.3 clone and categorized by T7 deletion mutant 4107 plaque test. The only sequence differences found in 15 of the 22 variants are conversion of the L8 and UV5 mutations to the wild-type sequence and conversion of the

base pair ahead of *lacO1* from the G of K12 to the A of BL21(DE3), a strong indication that these changes were due to gene conversion from the *lac* operon elsewhere in the BL21(DE3) genome. The same explanation holds for another 3 variants, which differ from these 15 only in retaining the L8 mutation in the CRP binding site, the same variant isolated and studied by Miroux et al. (1996) *J. Mol. Biol.* 260:289-298 and Schlegel et al. (2015) *Cell Reports* 10:17581766. The remaining 4 variants are identical to BL21(DE3) in this region. Three of these variants are the only ones that failed to show detectable plaques in the presence of inducer, and the fourth gave the smallest plaques observed in the presence of inducer, suggesting that these variants may have mutations in T7 RNA polymerase that decrease its activity or inactivate it.

The entire coding sequence for T7 RNA polymerase was determined for 8 of the 22 variants of BL21(DE3), selected to represent the types distinguishable by the combination of plaque assay and the sequence determined for the upstream control sequence. Four variants that increase the repertoire of useful T7 expression hosts are here referred to as BL21(DE3)v1, v2, v3, and v4. The v1 variant (SEQ ID NO. 82) has a detectable but reduced level of basal expression relative to BL21(DE3) but retains full induced expression, as indicated by plaque size in the absence and presence of inducer and by the levels of T7 10a protein produced from a pREX LS31 clone by auto-induction. This variant is representative of 13 of the 15 variants that converted the upstream control region to the wild-type *lac* promoter. It has no changes in the T7 RNA polymerase sequence. The v2 variant (SEQ ID NO. 83) has no basal expression detectable by the plaque assay and a somewhat lower induced expression relative to BL21(DE3), as indicated by a somewhat smaller plaque size in the presence of inducer and lower 10a expression levels by auto-induction. This variant is representative of the 3 variants that converted all but the L8 mutation in the CRP binding site in the upstream control region to wild type. It likewise has no changes in the T7 RNA polymerase sequence. The v3 variant (SEQ ID NO. 84) has no basal activity detectable by the plaque assay and an induced activity comparable to that of v2. Sequencing found that this variant is identical to v1 in the upstream control region but has acquired a mutation of alanine-102 of T7 RNA polymerase to threonine. Another of the 8 fully sequenced variants is identical to this one. The v4 variant (SEQ ID NO. 85) has no basal activity detectable by the plaque assay and a somewhat lower induced activity than v2 and v3. This variant is identical to BL21(DE3) in the upstream control region and has acquired a mutation of proline-818 of T7 RNA polymerase to glutamine. The relative activity of the alanine-102 and proline-818 mutant RNA polymerases is difficult to assess from these results, because v3 should have a higher level of induced expression than v4. The 3 remaining sequence variants have no basal or induced activity detectable by the plaque assay: two of them have an amber termination mutation at glutamine-36 and the third has an 8-bp duplication at leucine-706, which changes the reading frame.

Variants that reduce basal expression of target protein increase the range of target proteins that can be stably maintained and produced in inducible T7 expression strains. Hosts that supply mutant but functional T7 RNA polymerases, such as the two isolated here or others that could be isolated and characterized by the methods disclosed here, may have unique advantages for adjusting rates of transcription to enhance the production of properly folded or secreted and functional target proteins.

The present disclosure contemplates T7 expression strains that contain the coding sequence for T7 RNA polymerase discussed above. Examples of T7 expression strains include BL21 CodonPlus and Lemo21(DE3).

General Methods for Cloning into pAL Vectors

Commonly used growth media can be suitable for growing *E. coli* hosts for cloning, transformation, preparation of freezer stocks, isolation of plasmids and maintenance of clones for expressing most target proteins in pREX and pAL vectors. However, variability in complex components in most media can cause unintended induction and instability (Studier, 2005). To avoid or minimize such problems, fully defined growth media made entirely from purified components are used routinely (Studier (2014) *Methods in Molecular Biology* (Clifton, N.J.) 1091:17-32; and Studier, U.S. Pat. Nos. 8,241,887 and 7,704,722). For induction of expression, the auto-induction media and methods of Studier (2014) and Studier, U.S. Pat. Nos. 7,560,264; 7,759,109; and 8,399,217 are useful, the contents of which are incorporated herein by reference.

Studier (2014) describes non-inducing growth media MDAG-135+B1 (B1=1 μ M thiamine) and MDAG-11+B1. MDAG-135+B1 is suitable for growth to high density of well-aerated cultures of DB3.1, XL1B, BL21 and strains derived from BL21, such as BL21(DE3) and BL21-AI. Kanamycin at a concentration of 100 μ g/ml is routinely used for selection and maintenance of pREX and pAL clones in these media, where little if any unintended induction should occur. Indeed, no target protein has been found to be so toxic to the host that an expressible gene could not readily be cloned in an appropriate pREX or pAL vector in XL1B and transferred to BL21(DE3) and BL21-AI in these media. Even strains showing signs of stress could often be grown to high density with good yields of plasmid. MDAG-135+B1 is also good for preparation of chemically competent DB3.1 for transformation (currently not commercially available).

MDAG-11+B1 has a lower concentration of glucose, which limits acid production at saturation in less well aerated conditions. This medium (plus selective antibiotic) is suitable for selection of transformants on 1% agar plates, and colonies typically remain viable on these plates for weeks in the refrigerator. Plates can be stored refrigerated before use but should be well equilibrated to room temperature for several hours before placing in a 37° C. incubator, to prevent formation of small air bubbles in the agar. This medium is also used for 0.7% agar stabs for mailing cultures or for poorly aerated or standing cultures.

Freezer stocks for long-term storage are made by mixing 1 ml of culture with 0.1 ml of 80% glycerol in a 2-ml plastic freezer tube and placing directly in a -70° C. freezer. Cultures of most clones are grown to saturation before freezing but should not be allowed to grow beyond mid-log phase before freezing if the clone is highly toxic to the host. Cultures are inoculated from freezer stocks by scraping some frozen culture from the surface without thawing the rest of the tube.

Standard procedures are used for isolating plasmids, high-fidelity PCR amplifications of DNA, enzyme treatments, purification of amplified or enzymatically treated DNA and transformation of chemically competent cells. Quality and concentration of purified DNA are routinely determined by Nanodrop Spectrometry and agarose gel electrophoresis. Typical commercial products include: Wizard Plus SV Mini-prep DNA Purification System for purifying plasmids (Promega); Accuprime Pfx SuperMix for high-fidelity PCR

(Invitrogen/Life Technologies); synthetic DNA primers (Integrated DNA Technologies); illustra GFX PCR DNA and Gel Band Purification Kit (GE Healthcare Life Sciences); DNA Clean & Concentrator-5 Kit (Zymogen); BsaI-HF, other restriction enzymes, T4 DNA polymerase, T4 ligase, T4 polynucleotide kinase, NTPs, dNTPs and reaction buffers (New England Biolabs); XL1B-MR Chemically Competent cells (Agilent); BL21(DE3) Chemically Competent cells (Novagen); and BL21-AI Chemically Competent cells (Invitrogen/Life Technologies). In a five-minute reaction at room temperature, purified PCR product (or blunt-end synthetic DNA) is treated with T4 DNA polymerase in the presence of the appropriate dNTP to generate the overhangs for cloning into pAL vectors and, in the same reaction mixture, with T4 Polynucleotide Kinase and ATP to phosphorylate the 5' ends of the overhangs. For convenience, the reaction volume is usually 50 μ l and contains ~10 nM DNA, 0.2 mM dNTP, 1 unit of T4 DNA polymerase, 1 unit of T4 polynucleotide kinase and BSA at a concentration of 100 μ g/ml in New England Biolabs T4 DNA Ligase Reaction Buffer (50 mM Tris-HCl, 10 mM MgCl₂, 10 mM DTT, 1 mM ATP, pH 7.8). The reaction has also been scaled to 20 μ l volume when the amount of synthetic DNA was limiting. The reaction is stopped by adding 3 μ l of 0.5 M EDTA per 50 μ l of reaction mixture and the treated DNA is purified by the GFX process, eluting the microspin column with a volume of low TE (1 mM Tris, 0.1 mM EDTA, pH 8) equal to the reaction volume.

Purified pAL plasmid is digested with BsaI-HF, which is then heat-inactivated for 20 minutes at 65° C. The cut DNA is ethanol-precipitated for at least 1 hour at -70° C., washed, dried and dissolved in low TE. This DNA can be used immediately for cloning but, for convenience, we usually make a larger batch, analyze a sample by agarose gel electrophoresis to verify BsaI cutting and nanodrop concentration, and store aliquots at -20° C. for future cloning. In limited testing, purification of the acceptor fragment seemed to offer little if any advantage.

In another five-minute reaction at room temperature, the DNA to be cloned, whether overhangs were generated by T4 DNA polymerase or in the design of synthetic DNA, is ligated to the BsaI-cut pAL vector. A reaction volume of 20 μ l contains 1 nM concentration each of the cut vector and the DNA to be cloned and 400 units of T4 DNA ligase in New England Biolabs T4 DNA Ligase Reaction Buffer (50 mM Tris-HCl, 10 mM MgCl₂, 10 mM DTT, 1 mM ATP, pH 7.8). Typically, 1 μ l of the reaction mixture is used immediately for transformation of 25 μ l of chemically competent cells and the remainder is stored at -20° C. for further use, if needed.

Definitions

As used herein, a "host cell" includes any cell capable of proliferating the DNA vectors disclosed herein. In one embodiment, the host cell is a prokaryotic cell. Examples of suitable host cells include BL21 (DE3) and derivatives thereof.

As used herein, "prokaryote" and "prokaryotic cell" refer to cells which do not contain a nucleus and whose chromosomal material is thus not separated from the cytoplasm. Prokaryotes include, for example, bacteria. Prokaryotic host cells particularly embraced by the present invention include those amenable to genetic manipulation and growth in culture. Exemplary prokaryotes routinely used in recombinant protein expression include, but are not limited to, *E. coli*, *Bacillus licheniformis* (van Leen, et al. (1991) Bio/

Technology 9:47-52), *Ralstonia eutropha* (Srinivasan, et al. (2002) Appl. Environ. Microbiol. 68:5925-5932), *Methylobacterium extorquens* (Belanger, et al. (2004) FEMS Microbiol Lett. 231(2):197-204), *Lactococcus lactis* (Oddone, et al. (2009) Plasmid 62(2):108-18) and *Pseudomonas* sp. (e.g., *P. aeruginosa*, *P. fluorescens* and *P. syringae*). Prokaryotic host cells can be obtained from commercial sources (e.g., Clontech, Invitrogen, Stratagene and the like) or repositories such as American Type Culture Collection (Manassas, Va.). In particular embodiments, the prokaryotic host cell is *E. coli*.

As used herein, a “vector” is a DNA molecule to which heterologous DNA may be operatively linked so as to bring about replication of the heterologous DNA. Vectors are conventionally used to deliver DNA molecules to cells, including *E. coli* cells that are typically used in a majority of cloning or protein expression applications. Examples of vectors include plasmid, cosmid, and phages.

Type II restriction endonucleases are enzymes that cleave DNA at different positions close to or within their recognition sequences. Examples include XbaI, NcoI, and NotI.

Type IIS restriction endonucleases are enzymes that cleave DNA outside of their recognition sequences, to one side. Examples of type IIS restriction endonucleases include FokI, AlwI, and BsaI.

As used herein, a “target DNA” includes a polynucleotide. The term “polynucleotide” as used herein is defined as a chain of nucleotides. Polynucleotide includes DNA and RNA.

Furthermore, nucleic acids are polymers of nucleotides. Thus, nucleic acids and polynucleotides as used herein are interchangeable. One skilled in the art has the general knowledge that nucleic acids are polynucleotides, which can be hydrolyzed into the monomeric “nucleotides.” The monomeric nucleotides can be hydrolyzed into nucleosides. As used herein polynucleotides include, but are not limited to, all nucleic acid sequences which are obtained by any means available in the art, including, without limitation, recombinant means, i.e., the cloning of nucleic acid sequences from a recombinant library or a cell genome, using ordinary cloning technology and polymerase chain reaction (PCR), and the like, and by synthetic means.

As used herein, the terms “peptide,” “polypeptide,” and “protein” are used interchangeably, and refer to a compound having amino acid residues covalently linked by peptide bonds. A protein or peptide must contain at least two amino acids, and no limitation is placed on the maximum number of amino acids that can be included in a protein’s or peptide’s sequence. Polypeptides include any peptide or protein having two or more amino acids joined to each other by peptide bonds. As used herein, the term refers to both short chains, which also commonly are referred to in the art as peptides, oligopeptides, and oligomers, for example, and to longer chains, which generally are referred to in the art as proteins, of which there are many types. “Polypeptides” include, for example, biologically active fragments, substantially homologous polypeptides, oligopeptides, homodimers, heterodimers, variants of polypeptides, modified polypeptides, derivatives, analogs, and fusion proteins, among others. The polypeptides include natural peptides, recombinant peptides, synthetic peptides, or a combination thereof.

A “heterologous” peptide or polypeptide means a peptide or polypeptide encoded by a non-host DNA molecule. The heterologous peptide or polypeptide may be toxic to the host cell when expressed by way of an expression vector.

Further, unless expressly stated to the contrary, “or” refers to an inclusive “or” and not to an exclusive “or”. For example, a condition A or B is satisfied by any one of the following: A is true (or present) and B is false (or not present), A is false (or not present) and B is true (or present), and both A and B are true (or present).

In this specification, groups of various parameters containing multiple members are described. Within a group of parameters, each member may be combined with any one or more of the other members to make additional sub-groups. For example, if the members of a group are a, b, c, d, and e, additional sub-groups specifically contemplated include any one, two, three, or four of the members, e.g., a and c; a, d, and e; b, c, d, and e; etc.

While there have been described what are presently believed to be the preferred embodiments of the present invention, those skilled in the art will realize that other and further changes and modifications may be made thereto without departing from the spirit of the invention, and it is intended to claim all such modifications and changes as come within the true scope of the invention.

TABLE 1

Sequences of Expression control Region (ECR) in T7 DNA; pET, pPREX, pAL vectors; ZA#11, ZA#13 in US 6,537,779									
SEQ ID	Expression Control Region (ECR)								
NO.	T7 promoter	T7 RNA start and palindrome	XbaI	tg10	ndeI				
8	T7 DNA	ACTTCG AAAT TAATACGACTCACTATA GGGAGACC ACAAC GGTTTCCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATACATATG							
9	pBT-3a	cccgCG AAAT TAATACGACTCACTATA GGGAGACC ACAAC GGTTTCCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATACATATG							
		RNA lacOI palindrome link							
10	pET-11d	cccgCG AAAT TAATACGACTCACTATA GGGG AATTGTGAGCGGATAACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAAAGTATAC CATG							
11	ZA#11	attcCG AAAT TAATACGACTCACTATA GGGG AATTGTGAGCGGATAACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAAAGTATACATATG							
		lacS palindrome							
12	ZA#13	attcCG AAAT TAATACGACTCACTATA GGGG AATTGTGAGCGGATAACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAAAGTATACATATG							
		attcCG AAAT TAATACGACTCACTATA GGGG AATTGTGAGC GCTCACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAAAGTATACATATG							
pPREX	pAL	T7 Promoter Control Region (TPCR)	Translation Initiation Efficiency Region (TIER)						
TPCR		T7 promoter RNA lacOI palindrome link XbaI tg10, SEQ ID No: 48 NcoI							
13	L44	cgtacg AAAT TAATACGACTCACTATA GGGG AATTGTGAGCGGATAACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
14	L33	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGCGGATAACAATT CCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
15	L22	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGCGGATAACAATT CC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
16	L1m	cgtacg AAAT TAATACGACTCACTATA G AATTGTGAGCGGATAACAATT C CTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
		lacS palindrome							
20	S34	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
21	S33	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT CCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
22	S32	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT CC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
23	S31	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT C TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG	pAL1						
24	S30	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
25	S3m	cgtacg AAAT TAATACGACTCACTATA GGG AATTGTGAGC GCTCACAATT CTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
26	S24	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT CCCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
27	S23	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT CCC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
28	S22	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT CC TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
29	S21	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT C TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG	pAL2						
30	S20	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT TCTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
31	S2m	cgtacg AAAT TAATACGACTCACTATA GG AATTGTGAGC GCTCACAATT CTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							
32	S1m	cgtacg AAAT TAATACGACTCACTATA G AATTGTGAGC GCTCACAATT CTAGA AATAAATTTGTTTAACTTTAAGAAAGGAGATATAC CATG							

TABLE 1-continued

Sequences of Expression control Region (ECR) in T7 DNA; pET, pREX, pAL vectors; ZA#11, ZA#13 in US 6,537,779					
Upstream 37-bp lacO1 with extended natural palindrome (and spacer sequence) in pREX-LS vectors, pAL3 and pAL4					
SEQ ID NO.	SacII	lacO1 spacer	BsiWI	T7 promoter	RNA
86	CGTA CCGCGG	TTGTGTGG <u>AATTGTGAGCGGATAACAATT</u> <u>TCACACAG</u> AACACAGTCCCT cgtacG AAAT TAATACGACTCACTATA GG			

Bases in the 37-bp extended natural palindrome of lacO1 are underlined
lacO1 operators are in the same orientation as in the lac operon and in pET vectors
Center-to-center spacing between the lacO1 operators is 70.5 bp in pAL3 and pREX-LS3 vectors and is 69.5 bp in pAL4 and pREX-LS2 vectors

TABLE 2

Deletions and sequence changes between the Thai site and Shine-Dalgarno (SD) sequence of the tg10 Translation Initiation Efficiency Region (TIER) affect ability to clone T7 gene 5.3 and to produce T7 gene 10a protein in pREX L44.											
SEQ		Translation Initiation Efficiency Region (TIER)						T7 lac-5.3		T7 lac-10a	
ID NO.	TIER	XbaI <SD bp	XbaI site	Shine-Dalgarno sequence	Start codon	transformants XL1B	BL21 (DE3)	BL21 Al	induced protein		
48	tg10	28	TCTAGA	<u>AATAATTTT</u> GTTT <u>AACTTT</u> AAG	AAGGAG ATATACC ATG	0	0	0	+++		
49	tr4	24	TCTAGA	AAT <u>TTC</u> <u>TTTAACTTT</u> AAG	AAGGAG ATATACC ATG	sm	(tiny)	+	+++		
50	tr11	16	TCTAGA	<u>TA</u> ACTTTAAG	AAGGAG ATATACC ATG	+	(tiny)	+	+++		
51	tr10	15	TCTAGA	<u>AACTTT</u> AAG	AAGGAG ATATACC ATG				+++		
52	tr9	14	TCTAG	<u>AACTTT</u> AAG	AAGGAG ATATACC ATG	+	(tiny)	+	(+++)		
53	tr8	14	TCTAGA AA	<u>TTT</u> AAG	AAGGAG ATATACC ATG	+	(sm)	+	(+++)		
54	tr2	13	TCTAG	<u>ACTTT</u> AAG	AAGGAG ATATACC ATG	+	sm	+	++		
55	tr7	12	TCTAGA AA	<u>TA</u> AG	AAGGAG ATATACC ATG	+	tiny	+	(++)		
56	tr6	10	TCTAGA	<u>TA</u> AG	AAGGAG ATATACC ATG	+	tiny	+	+		
57	tr5	8	TCTAG	<u>AAG</u>	AAGGAG ATATACC ATG	+	sm	+	+		
58	tr1	5	TCT	<u>AG</u>	AAGGAG ATATACC ATG	+	med	+	(+)		
59	tr3	12	TCTAGA	<u>AGACTA</u>	CATGTG GTCTCCC ATG	+	+	+	0?		

XbaI→SD column gives the number of base pairs ahead of the Shine-Dalgarno (SD) sequence in the Translation Initiation Efficiency Region (or a comparable number for tr3, which has no obvious SD sequence)
Potential -10 sequences for *E. coli* promoters (TataaT) are underlined.
A potential -10 sequence (TAGACT) created by deletion in tr2 appears to have little effect on basal expression in pREX L44-tr2, as judged by ability to clone and maintain gene 5.3 in XL1B and BL21(DE3)

TABLE 3

Ability to clone and express T7 5.3 and 10a coding sequences in different pREX vectors													
SEQ ID		Configuration of Expression Control Region (ECR)							5.3 clones in			10a clones	
		downstream of T7 promoter sequence							pREX transformed colonies			Protein level	
		RNA											
NO.	TPCR	start	lac operator		Linker	XbaI			XL1B	DE3	Al	DE3	Al
13	L44	GGGG	AATT	O1	AATT	CCCC	TCTAGA	tg10	0	0	0	+++	+++
14	L33	GGG	AATT	O1	AATT	CCC	TCTAGA	tg10	tn + sm	0	0	+++	+++
15	L22	GG	AATT	O1	AATT	CC	TCTAGA	tg10	sm	0	str	+++	++
16	L1m	G	AATT	O1	AATT		CTAGA	tg10	sm	0	sm	?	(+)
20	S34	GGG	AATT	S	AATT	CCCC	TCTAGA	tg10	0	0	0	+++	+++
21	S33	GGG	AATT	S	AATT	CCC	TCTAGA	tg10	md + tn	0	0	+++	(+++)
22	S32	GGG	AATT	S	AATT	CC	TCTAGA	tg10	mlg + tn	sm	mlg	+++	(+++)
23	S31	pAL1	GGG	AATT	S	AATT	C	TCTAGA	(lg)	sm	mlg	+++	(+++)
24	S30	GGG	AATT	S	AATT		TCTAGA	tg10	mlg	sm	mlg	+++	(+++)
25	S3m	GGG	AATT	S	AATT		CTAGA	tg10	mlg	sm	mlg	+++	(+++)
26	S24	GG	AATT	S	AATT	CCCC	TCTAGA	tg10	0	0	0	+++	++
27	S23	GG	AATT	S	AATT	CCC	TCTAGA	tg10	tn + md	0	0	+++	++
28	S22	GG	AATT	S	AATT	CC	TCTAGA	tg10	mlg + tn	sm	mlg	+++	++
29	S21	pAL2	GG	AATT	S	AATT	C	TCTAGA	mlg	smd	mlg	+++	++

TABLE 3-continued

Ability to clone and express T7 5.3 and 10a coding sequences in different pREX vectors													
SEQ ID	pREX	Configuration of Expression Control Region (ECR)							5.3 clones in			10a clones	
		downstream of T7 promoter sequence							pREX transformed colonies			Protein level	
		RNA					TIER SEQ #48						
NO.	TPCR	start	lac operator		Linker	XbaI		XL1B	DE3	A1	DE3	A1	
30	S20	GG	AATT	S	AATT		TCTAGA tg10	mlg	smd	mlg	+++	++	
31	S2m	GG	AATT	S	AATT		CTAGA tg10	mlg	smd	mlg	+++	+++	
32	S1m	G	AATT	S	AATT		CTAGA tg10	smd	med	mlg	(+)	+	
36	LS34	GGG	AATT	S	AATT	CCCC	TCTAGA tg10	0	0	0	+++	++	
37	LS33	GGG	AATT	S	AATT	CCC	TCTAGA tg10	lg	0	str	(+++)	++	
38	LS32	GGG	AATT	S	AATT	CC	TCTAGA tg10	lg	mlg	lg	+++	++	
39	LS31	pAL3	GGG	AATT	S	AATT	C	TCTAGA tg10	lg	mlg	lg	+++	(++)
40	LS30	GGG	AATT	S	AATT		TCTAGA tg10	lg	mlg	lg	+++	(++)	
41	LS3m	GGG	AATT	S	AATT		CTAGA tg10	lg	mix	lg	+++	+	
42	L24	GG	AATT	S	AATT	CCCC	TCTAGA tg10	0	0	0	(+++)	+	
43	LS23	GG	AATT	S	AATT	CCC	TCTAGA tg10	mlg	0	str	+++	+	
44	LS22	GG	AATT	S	AATT	CC	TCTAGA tg10	lg	mlg	lg	(+++)	+	
45	LS21	pAL4	GG	AATT	S	AATT	C	TCTAGA tg10	lg	mlg	lg	+++	+
46	LS20	GG	AATT	S	AATT		TCTAGA tg10	lg	mlg	lg	+++	+	
47	LS2m	GG	AATT	S	AATT		CTAGA tg10	lg	mlg	lg	+++	+	

T7 Promoter Control Region (TPCR) designations for pREX and pAL vectors (full spectrum in Table 1)

L indicates a minimal 21-bp lacO1 asymmetric operator downstream of the T7 promoter sequence

S indicates an extended 37-bp lacS symmetric operator downstream of the T7 promoter sequence

LS indicates an extended 37-bp lacO1 operator downstream and a 20-bp lacS operator downstream of the T7 promoter sequence, separated by 69.5 or 70.5 bp center-to-center for maximum repression

Asymmetric lacO1 operators have the same orientation as in the lac operon and in pET vectors

The second integer gives the number of C residues linking the downstream operator to the XbaI site

An "m" indicates that the T of the downstream operator is also the first T of the XbaI site

Translational Initiation Efficiency Region (TIER) is tg10 in all vectors in the table (effects of other TIER seqs in Table 2)

pREX vectors are identified unambiguously by a TPCR-TIER compound name; if TPCR only is given, the TIER is tg10

Attempts to maintain pREX clones of T7 gene in the hosts XL1B, BL21(DE3), and BL21-A1 are summarized:

0 did not obtain transformants with active 5.3

str cultures obviously stressed, susceptible to overgrowth by mutants

tn some or most clones were inactivated by tn10 insertion in SL1B

sm small or late-developing colonies

smd small-medium colonies

med medium colonies

mlg medium-large colonies

lg large colonies

mix mixture of colonies of different sizes

TABLE 4

Configurations flanking the cloning sites of different pAL vectors, and sequences at ends of PCR products for generating 4-nt 5' overhangs for ligation to pAL cloning acceptors						
SEQ ID#	Vector	ECR	N-terminal fusion	Cloning site and counter-selection module	Optional C-terminal fusions	Blunt ends of PCR for 4-nt 5' overhangs upstream-downstream
						T7 DNAF + dCTP
66	pAL1	S31-tg10		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-Gly-6His	-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
67	pAL2	S21-tg10		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-Gly-6His	-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
68	pAL3	LS21-tg10		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-Gly-6His	-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
69	pAL4	LS21-tg0		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-Gly-6His	-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
70	pAL5	S31-tg5		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-Gly-6His	-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
71	pAL11	S31-tg10		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
72	pAL12	S31-tr5		-NcoI- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CCATG-NNCAGTG</u> 3'
73	pAL13	S31-tg10	-NdeI-NheI-6His-	<u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-TAA -Acc65I	-BamHI-TAA -Acc65I	5' <u>ACTCG-NNCAGTG</u> 3'
74	pAL14	S31-tr5	-NdeI-NheI-6His-	<u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-TAA -Acc65I	-BamHI-TAA -Acc65I	5' <u>ACTCG-NNCAGTG</u> 3'
						T4 DNAP + dTTP
75	pAL21	S31-tg10	-NdeI- pelB	- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CTGCA-NNTAGCG</u> 3'
76	pAL22	S31-tr5	-NdeI- pelB	- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CTGCA-NNTAGCG</u> 3'
77	pAL23	S31-tg10	-NdeI- dsbA	- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CTGCA-NNTAGCG</u> 3'
78	pAL24	S31-tr5	-NdeI- dsbA	- <u>BsaI</u> -AgeI-ccdB-EagI- <u>BsaI</u> -NNC-Ser-dsbc(21-93)-BamHI-6His-Acc65I	-BamHI-6His-Acc65I	5' <u>CTGCA-NNTAGCG</u> 3'

The outward facing BsaI sites for cloning are underlined, as are the codons in the PCR primers that will fuse the target coding sequence in the correct reading frame for N- or C- terminal C-terminal fusions are optional. To terminate a coding sequence without a C-terminal fusion, the termination codon (default TAA) is places ahead of the 5-bp C-terminal fusion sequence in the PCR primer. C-terminal fusions represented in these vectors include 6His affinity tags and the dimerization domain of dsbc. N-terminal fusions include 6His affinity tags, the N-terminal pelB leader sequence, which directs secretion into the periplasm by the Sec pathway, and the N-terminal dsbA leader sequence, which directs into the periplasm by the SRP pathway. Achieving optimal secretion and/or folding of target protein may require matching the rate of target protein production to the secretion and/or folding capacity of the cell. Basal and induced expression will be higher with the tg10 upstream translation sequence than the tr5 and will be generally higher in BL21(DE3) than in BL21-AL. Other rates of target protein production could be achieved with other vector-host configurations.

INCORPORATION OF SEQUENCE LISTING

A Sequence Listing conforming to the rules of WIPO Standard ST.25 is hereby incorporated by reference. Said

Sequence Listing has been filed as an electronic document via PatentCenter in ASCII text. The electronic document, created on Feb. 16, 2023, is entitled "IP2014-019-03-_PCT-US_ST25.txt", and is 147,312 bytes in size.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 102

<210> SEQ ID NO 1

<211> LENGTH: 2650

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 1

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tatatactga tatctatacc cgaagtatgt caaaaagagg tgtgctatgc agtttaaggt    180
ttacacctat aaaagagaga gccgttatcg tctgtttgtg gatgtacaga gtgatattat    240
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agtctcccggt gaactttacc cgggtggtgca tatcggggat gaaagctggc gcatgatgac    360
caccgatatg gccagtgtgc cagtctccgt tatcggggaa gaagtggctg atctcagcca    420
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cctctaaacg ggtcttgagg ggttttttgc tgaaagctta cgcggcgccc tgccactcat    720
cgcagtactg ttgtaattca ttaagcattc tgccgacatg gaagccatca caaacggcat    780
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tttccgaagg taactggctt cagcagagcg cagataccaa atactgtcct tctagtgtag   1800
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tgctatgcag tttAaggttt acacctataa aagagagagc cgttatcgtc tgtttTgtgga	180
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acgtctgctg tcagataaag tctcccgTga actttaccCG gtggTgcata tcgggggatga	300
aagctggcgc atgatgacca ccgatatggc cagtgtgcca gtctccgtta tcggggaaga	360
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<220> FEATURE:
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<400> SEQUENCE: 21

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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 22

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<210> SEQ ID NO 23
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 23

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<400> SEQUENCE: 24

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<210> SEQ ID NO 25
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<220> FEATURE:
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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 26

cgtagcgaat taatagcact cactatagga attgtgagcg ctcacaattc ccc 53

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<220> FEATURE:
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<220> FEATURE:
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aatacgactc actatagggg aattgtgagc gtcacaatt	100	
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<400> SEQUENCE: 36

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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 37

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aatacgactc actataggga attgtgagcg ctcacaattc cc 102

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

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aatacgactc actataggga attgtgagcg ctcacaattc c 101

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<213> ORGANISM: Artificial Sequence
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aatacgactc actataggga attgtgagcg ctcacaattc 100

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<211> LENGTH: 102
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aatacgactc actataggaa ttgtgagcgc tcacaattcc cc 102

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<220> FEATURE:
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<400> SEQUENCE: 43

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aatacgactc actataggaa ttgtgagcgc tcacaattcc c 101

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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 44

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aatacgactc actataggaa ttgtgagcgc tcacaattcc 100

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<400> SEQUENCE: 45

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aatacgactc actataggaa ttgtgagcgc tcacaattc 99

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aatacgactc actataggaa ttgtgagcgc tcacaatt 98

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<220> FEATURE:		
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<212> TYPE: DNA		
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<220> FEATURE:		
<223> OTHER INFORMATION: Synthetic sequence		
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<213> ORGANISM: Artificial Sequence		
<220> FEATURE:		
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<211> LENGTH: 30		
<212> TYPE: DNA		
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<223> OTHER INFORMATION: Synthetic sequence		
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<400> SEQUENCE: 60

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

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tatactgata tctatacccg aagtatgtca aaaagagggtg tgctatgcag tttaaggttt 240
acacctataa aagagagagc cgttatcgtc tgthttgtgga tgtacagagt gatattattg 300
acacgcctgg gcgacgcatg gtgatcccc tggccagtgc acgtctgctg tcagataaag 360
tctcccgtag actttaccgg gtggtgcata tcggggatga aagctggcgc atgatgacca 420
ccgatatggc cagtgtgcca gtctccgtha tcggggaaga agtggctgat ctacagccacc 480
gcgaaaatga catcaaaaac gccattaacc tgatgttctg gggaatataa cggccggagg 540
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cctgcatttc gcgtaagact caagtcttat aactacgtaa ggatcctaag tttaaacagc 660
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

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<210> SEQ ID NO 64
<211> LENGTH: 4542
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 64

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<210> SEQ ID NO 65

<211> LENGTH: 4541

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 65

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ctgcatttct cgtaagactc aagtcttata actacgtaag gatcctaagt ttaaacagcc	660
cgggcattta aatcagcgat cgcaagggtac cgctgagcaa taactagcat aacccttggt	720
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<210> SEQ ID NO 66

<211> LENGTH: 4363

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 66

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cctataaaaag agagagccgt tatcgtctgt ttgtggatgt acagagtgat attattgaca	300
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<210> SEQ ID NO 67

<211> LENGTH: 4362

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 67

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cggagggtgg	cgggcaggac	gcccgccata	aactgccagg	catcaaatta	agcagaaggc	4320
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<210> SEQ ID NO 68

<211> LENGTH: 4422

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 68

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ctaaaagcca	gataacagta	tgcgtatttg	cgcgctgatt	tttgcggtat	agaatatata	180
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<210> SEQ ID NO 69
 <211> LENGTH: 4421
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 69

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taaaagccag ataacagtat gcgtatttgc gcgctgattt ttgcggtata agaatatata	180
ctgatatcta taccggaagt atgtcaaaaa gaggtgtgct atgcagttta aggtttacac	240
ctataaaaga gagagccgtt atcgtctgtt tgtggatgta cagagtgata ttattgacac	300
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tggacatcac caccatcatc actaaggtag cgctgagcaa taactagcat aacccttgg	600
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tcataaaaat cactcgcatc aaccaaaccg ttattcattc gtgattgcgc ctgagcgaga	1020
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catcctgacg gatggccttt ttgcgtttct acaaaactct ccgtaccgcg gttgtgtgga	4380
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<210> SEQ ID NO 70
<211> LENGTH: 4343
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 70

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caacaatatt ttcacctgaa tcaggatatt cttctaatac ctggaatgct gttttcccg	1140
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<210> SEQ ID NO 71

<211> LENGTH: 4585

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 71

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<210> SEQ ID NO 72

<211> LENGTH: 4565

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 72

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<210> SEQ ID NO 73
<211> LENGTH: 4595
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 73

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cggagaccgg taaaatacat aaggcttact aaaagccaga taacagtatg cgtatttgcg	180
cgtgatTTT tgcggtataa gaatatatac tgatatctat acccgaagta tgtcaaaaag	240
aggTgtgcta tgcagTTta ggtttacacc tataaaagag agagccgTTa tcgtctgttt	300
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agtgcacgtc tgctgtcaga taaagtctcc cgtgaacttt acccggtggt gcatatcggg	420
gatgaaagct ggcgcatgat gaccaccgat atggccagtg tgccagtctc cgttatcggg	480
gaagaagtgg ctgatctcag ccaccgcgaa aatgacatca aaaacgcat taacctgatg	540
ttctggggaa tataacggcc ggtctccagt gatgacggc caattcaaca aacgttagcc	600
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ctgactaaca gcggcgtgtt gtacatcacc gatgatggta aacatatcat tcaggggcca	720
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ttgaatgcgg gatcctaagg taccgctgag caataactag cataaccctt tggggcctct	840
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tcaaatgaaa	ctgcaattta	ttcatatcag	gattatcaat	accatatttt	tgaaaaagcc	960
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ccagagaaaa	atcaccttag	gtcactgccc	gctttccagt	cgggaaacct	gtcgtgccag	3240
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<210> SEQ ID NO 74
<211> LENGTH: 4575
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 74

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ggtctccagt	gatgacgcgg	caattcaaca	aacgttagcc	aaaatgggca	tcaaaagcag	600
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<210> SEQ ID NO 75

<211> LENGTH: 4652

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 75

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aggcttacta aaagccagat aacagtatgc gtatttgcgc gctgattttt gcggtataag	240
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agtaggacaa atccgcggg agcggatttg aacgttgca agcaacggcc cggagggtgg	4560
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<210> SEQ ID NO 76

<211> LENGTH: 4632

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 76

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gccaaccag cgatggctgc agagaccgg aaaatacata aggcttacta aaagccagat	180
aacagtatgc gtatttgccg gctgattttt gcggtataag aatatatact gatatctata	240
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cccgttggtg	catatcgggg	atgaaagctg	gcgcagatg	accaccgata	tggccagtgt	480
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ccgaacgacc	gagcgcagcg	agtcagtgag	cgaggaagcg	gaagagcgcc	tgatgcggta	2640
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acaaactctt cc	4632

<210> SEQ ID NO 77
<211> LENGTH: 4643
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

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<400> SEQUENCE: 77

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gtttagtttt agcgttcagc gcaagcgctg cagagaccgg taaaatacat aaggcttact	180
aaaagccaga taacagtatg cgtatttgcg cgctgatttt tgcggtataa gaatatatac	240
tgatatctat acccgaagta tgtcaaaaag aggtgtgcta tgcagtttaa ggTTtAcacc	300
tataaaagag agagccgtta tegtctgttt gtggatgtac agagtgatat tattgacacg	360
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atggccagtg tgccagtctc cgttatcggg gaagaagtgg ctgatctcag ccaccgcgaa	540
aatgacatca aaaacgccat taacctgatg ttctggggaa tataacggcc ggtctcaagc	600
gatgacgagg caattcaaca aacgttagcc aaaatgggca tcaaaagcag cgatattcag	660
cccgcgcccg tagctggcat gaagacagtt ctgactaaca gcggcgtgtt gtacatcacc	720
gatgatggta aacatatcat tcaggggcca atgtatgacg ttagcggcac ggctccggtc	780
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<210> SEQ ID NO 78
<211> LENGTH: 4623
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 78

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cgtatttgcg cgctgatttt tgcggtataa gaatatatac tgatatctat acccgaagta	240
tgtcaaaaag aggtgtgcta tgcagtttaa ggtttacacc tataaaagag agagccgtta	300
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gaagacagtt ctgactaaca gcggcgtgtt gtacatcacc gatgatggta aacatatcat	720
tcaggggcca atgtatgacg ttagcggcac ggctccggtc aatgtcacca ataagatgct	780
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<210> SEQ ID NO 79
<211> LENGTH: 17
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 79

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<210> SEQ ID NO 80
<211> LENGTH: 17
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 80

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<210> SEQ ID NO 81
<211> LENGTH: 3257
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 81

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<210> SEQ ID NO 82
<211> LENGTH: 3257
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence

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<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 82

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cagctatgac catgattacg gattcactgg ccgctgtttt acaacgtcgt gactgggaaa	180
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<210> SEQ ID NO 83

<211> LENGTH: 3257

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 83

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cagctatgac catgattacg gattcactgg ccgtcgtttt acaacgtcgt gactgggaaa	180
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<211> LENGTH: 3257
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 84

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cagctatgac catgattacg gattcactgg ccgtcgtttt acaacgtcgt gactgggaaa	180
accctggcgt tacccaactt aatcgccttg cagcacatcc ccctttcgcc agctggcgta	240
atagcgaaga ggcccgacc gatcgccctt cccaacagtt gcgcagcctg aatggcgaat	300
ggcgctttgc ctggtttccg gcaccagaag cggtgccgga aagctggctg gagtgcgatc	360
ttcctgaggg cgatactgtc gtcgtccctt caaactggca gatgcacggg tacgatgcgc	420
ccatctacac caacgtgacc tatccatta cggtcfaatcc gccgtttgtt cccacggaga	480
atccgacggg ttgttactcg ctcacattta atgttgatga aagctggcta caggaaggcc	540
agacgcgaat tatttttgat ggcgtcggga tctgatccgg atttactaac tggaagaggc	600
actaaatgaa cacgattaac atcgctaaga acgacttctc tgacatcgaa ctggctgcta	660
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gtgatgtact	ggctgatttc	tacgaccagt	tcgctgacca	gttgacagag	tctcaattgg	3180
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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 85

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cagctatgac	catgattacg	gattcactgg	ccgtcgtttt	acaacgtcgt	gactgggaaa	180
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ggcgctttgc	ctggtttccg	gcaccagaag	cggtgccgga	aagctggctg	gagtgcgac	360
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agacgcgaat	tatttttgat	ggcgtcggga	tctgatccgg	atttactaac	tggaagaggg	600
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tcccgttcaa	cactctggct	gaccattacg	gtgagcggtt	agctcgcgaa	cagttggccc	720
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<210> SEQ ID NO 86
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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 86

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gtacgaaatt aatacgactc actatagg 88

<210> SEQ ID NO 87
<211> LENGTH: 43
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

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ctagaaataa ttttgttttaa ctttaagaag gagatatacc atg 43

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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: n is a, c, g, or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(6)
<223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 88

nnnnnaata atttgtttta actttaagaa ggagatatatc catg 44

<210> SEQ ID NO 89
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<212> TYPE: DNA
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<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: n is a, c, g, or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(5)
<223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 89

nnnnnaataa ttttgtttta ctttaagaag gagatatacc atg 43

<210> SEQ ID NO 90
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 90

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<210> SEQ ID NO 91

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<211> LENGTH: 44
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 91

ctagaaataa ttttgtttaa ctttaagaag gagatataca tatg 44

<210> SEQ ID NO 92
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: n is a, c, g, or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(6)
<223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 92

nnnnnaata attttgttta actttaagaa ggagatatatc atatg 45

<210> SEQ ID NO 93
<211> LENGTH: 44
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: n is a, c, g, or t
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<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(5)
<223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 93

nnnnnaataa ttttgtttaa ctttaagaag gagatataca tatg 44

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<220> FEATURE:
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<400> SEQUENCE: 94

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<210> SEQ ID NO 95
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic sequence

<400> SEQUENCE: 95

taatacgact cactataggg aattgtgagc gctcacaatt 40

<210> SEQ ID NO 96
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<220> FEATURE:	
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<210> SEQ ID NO 98	
<211> LENGTH: 41	
<212> TYPE: DNA	
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<220> FEATURE:	
<223> OTHER INFORMATION: Synthetic sequence	
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taatacgact cactatagga gaattgtgag cgctcacaat t	41
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<212> TYPE: DNA	
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<223> OTHER INFORMATION: Synthetic sequence	
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<212> TYPE: DNA	
<213> ORGANISM: Artificial Sequence	
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taatacgact cactataagg gaattgtgag cgctcacaat t	41

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The invention claimed is:

1. A vector comprising:

a T7 Promoter Control Region polynucleotide selected from the group consisting of: SEQ ID NO: 17, 18, 19, 23, 24, 25, 29, 30, 31, 32, 33, 34, 35, 38, 39, 40, 41, 44, 45, 46, 47, 94, 95, 96, 97, 98, 99, 100, 101, and 102; and

a Translation Initiation Efficiency Region polynucleotide selected from the group consisting of SEQ ID NO: 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 87, 88, 89, 90, 91, 92, and 93;

wherein,

the T7 Promoter Control Region polynucleotide is joined at the 3' end to the 5' end of the Translation Initiation Efficiency Region polynucleotide; and

a spacer comprising between 0, 1, or 2 nucleotide residues is disposed between the T7 Promoter Control Region polynucleotide and the Translation Initiation Efficiency Region polynucleotide.

2. The vector of claim 1 wherein said vector comprises at least one of:

a site for directional cloning of a target coding sequence by ligation to an acceptor overhang that is complementary to an upstream overhang 5'-CATG sequence and ligation to a downstream acceptor overhang comprising TA-3' or TG-3';

a cloning-acceptor fragment wherein all transcribed elements except the primer for DNA replication are in an orientation opposite to that of the target coding sequence; and

a DNA sequence between said upstream and downstream acceptor overhangs of said site for directional cloning comprises a counter-selection module.

3. The vector of claim 1 wherein said vector comprises at least one of: a lacI_t enhanced translation start (SEQ ID NO: 61); and

a sequence for constitutive expression of the ccdB toxin of *E. coli* F factor (SEQ ID NO: 2).

4. A vector for directional cloning of at least one target DNA by asymmetric ligation comprising:

a cloning site with upstream and downstream outward-facing asymmetric recognition sequences for at least one Type IIS restriction endonuclease, said recognition sequences being situated such that cutting said vector with said at least one Type IIS endonuclease produces two vector fragments:

1) a cloning-acceptor fragment having a different asymmetric overhang at each end; and

2) a counter-selection fragment containing both of said outward-facing recognition sequences and having asymmetric overhangs complementary to those of said cloning-acceptor fragment; and

wherein the nucleotide sequences of said 4 asymmetric overhangs are designed so that only 2 of the 10 possible pairwise alignments between said overhangs form ungapped perfectly base-paired substrates for a DNA ligase to join either strand, thereby regenerating said vector, and additionally so that the number of consecutive perfect base-pairs from either ungapped end of each of the remaining 8 said possible pairwise alignments is minimized, thereby minimizing potential joining of either DNA strand at any of said remaining 8 possible pairwise aligned overhangs by said DNA ligase.

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5. The vector of claim 4 comprising at least one of:

an Expression Control Region polynucleotide positioned in said cloning-acceptor fragment so as to direct expression of at least one coding sequence in said directionally cloned at least one target DNA, wherein the Expression Control Region polynucleotide comprises:

a T7 Promoter Control Region polynucleotide selected from the group consisting of: SEQ ID NO: 17, 18, 19, 23, 24, 25, 29, 30, 31, 32, 33, 34, 35, 38, 39, 40, 41, 44, 45, 46, 47, 94, 95, 96, 97, 98, 99, 100, 101, and 102; and

a Translation Initiation Efficiency Region polynucleotide selected from the group consisting of SEQ ID NO: 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 87, 88, 89, 90, 91, 92, and 93; wherein,

the T7 Promoter Control Region polynucleotide is joined at the 3' end to the 5' end of the Translation Initiation Efficiency Region polynucleotide; and

a cloning-acceptor fragment wherein all transcribed elements except the primer for DNA replication are in an orientation opposite to that of the target coding sequence.

6. The cloning and expression vector of claim 5 comprising at least one of:

a lacI_t enhanced translation start (SEQ ID NO: 61); and counter-selection fragment comprising a sequence for constitutive expression of the ccdB toxin of *E. coli* F factor (SEQ ID NO: 2).

7. The vector of claim 6 wherein said upstream and downstream outward-facing recognition sequences comprise BsaI recognition sequence and wherein cutting by BsaI produces two vector fragments as follows:

1) a cloning-acceptor fragment having upstream and downstream acceptor overhangs; and

2) a counter-selection fragment.

8. The vector of claim 7 wherein comprising at least one of:

said upstream acceptor overhang comprises the complement of the first two nucleotides of the initiation codon of said Expression Control Region polynucleotide;

said vector comprises a coding sequence for a peptide or protein domain initiated at said Expression Control Region polynucleotide and capable of N-terminal fusion to said target protein through said upstream acceptor overhang;

said vector comprises a coding sequence for a peptide or protein domain following said downstream acceptor overhang and capable of C-terminal fusion to said target protein through said downstream acceptor overhang;

said vector comprises a coding sequence for a peptide or protein domain initiated at said Expression Control Region polynucleotide and capable of N terminal fusion to said target protein through said upstream acceptor overhang, and a coding sequence for a peptide or protein domain following said downstream acceptor overhang and capable of C-terminal fusion to said target protein through said downstream acceptor overhang; and

said upstream acceptor overhang is the complement of 5'-CCAT and said downstream acceptor overhang is 5'-AGTG.

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