Leptin Receptor Deficiency is Associated With Upregulation of Cannabinoid 1 Receptors in Limbic Brain Regions

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ABSTRACT Leptin receptor dysfunction results in overeating and obesity. Leptin regulates hypothalamic signaling that underlies the motivation to hyperphagia, but the interaction between leptin and cannabinoid signaling is poorly understood. We evaluated the role of cannabinoid 1 receptors (CB1R) in overeating and the effects of food deprivation on CB1R in the brain. One-month-old Zucker rats were divided into unrestricted and restricted (fed 70% of unrestricted rats) diet groups and maintained until adulthood (4 months). Levels of relative binding sites of CB1R (CB1R binding levels) were assessed using [3H] SR141716A in vitro autoradiography. These levels were higher (except cerebellum and hypothalamus) at 4 months than at 1 month of age. One month CB1R binding levels for most brain regions did not differ between Ob and Lean (Le) rats (except in frontal and cingulate cortices in Le and in the hypothalamus in Ob). Four month Ob rats had higher CB1R binding levels than Le in most brain regions and food restriction was associated with higher CB1R levels in all brain regions in Ob, but not in Le rats. CB1R binding levels increased between adolescence and young adulthood which we believe was influenced by leptin and food availability. The high levels of CB1R in Ob rats suggest that leptin’s inhibition of food-intake is in part mediated by downregulation of CB1R and that leptin interferes with CB1R upregulation under food-deprivation conditions. These results are consistent with prior findings showing increased levels of endogenous cannabinoids in the Ob rats corroborating the regulation of cannabinoid signaling by leptin. Synapse 62:637–642, 2008.

INTRODUCTION Cannabis has been known and used for its analgesic and appetitive properties. The active compound delta(9)tetrahydrocannabinol (Δ9-THC), the endogenous cannabinoid anandamide (AEA) (N-Arachidonoyl-lethanolamide), and the cannabinoid receptor 1 (CB1R) agonist, WIN55,212-2, increased food intake in rodents (Gomez et al., 2002; Jarbe and DiPatrizio, 2005; Williams and Kirkham, 2002) by binding with high affinity to the CB1R (Bayewitch et al., 1996; Felder et al., 1993). Furthermore, studies have shown that activation of CB1R results in fatty-acid synthesis (Osei-Hyiaman et al., 2005) and lipogenesis, both strong markers of obesity (Cota et al., 2003; Osei-Hyiaman et al., 2005). There is also evidence that endogenous cannabinoids may regulate food preferences from a study that showed a correlation between CB1R density and the intake of palatable (sweet and high-fat food) (Harrold et al., 2002). Since CB1R activation has been associated with obesity-related behavioral manifestations such as overeating and lipogenesis, many efforts have focused on discovering an

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“antimarijuana” like drug in hopes of finding a treatment for obesity (Gadde and Allison, 2006).

Indeed, CB1R inactivation has been associated with a decrease in food intake and weight gain. Studies have shown that the CB1R antagonist, Rimonabant (SR141716A), reduced body weight and food intake in obese (Ob) Zucker rats (Vickers et al., 2003) and obese mice (Ravinet Trillou et al., 2003). This CB1R antagonist also suppressed food intake induced by administration of the endocannabinoid anandamide into the ventromedial hypothalamus (Jamshidi and Taylor, 2001). Another CB1R antagonist, AM 251, has also been shown to reduce food intake and weight gain in Ob rats (Chambers et al., 2004). Furthermore, Ob mice treated with CB1R antagonists gained less weight and showed lower levels of the obesity-related messengers (insulin, glucose, and leptin) compared with age-matched untreated animals (Poirier et al., 2005). Finally, mice lacking CB1R are resistant to the effects of diet induced obesity and when fed a high fat-diet show lower food intake, and a diminished response to the rewarding properties of food compared with controls (Sanchis-Segura et al., 2004). Thus, it is not surprising that CB1R antagonists are a main target in the development of medications to treat obesity.

CB1R and endogenous cannabinoids are distributed throughout the brain. However, the role of endocannabinoids in the hypothalamus has been most consistently linked with regulation of food intake. Indeed leptin appears to decrease food intake in part by reducing the levels of endogenous cannabinoids (anandamide and 2-arachidonoyl glycerol) in the hypothalamus (Di Marzo et al., 2001). Here we assess the role of leptin on CB1R expression in the brain. For this purpose we compared the regional brain distribution of CB1R in obese Zucker rats, which are leptin deficient (Chua et al., 1996), with that of their lean counterparts at different developmental stages and diet conditions. We hypothesized that leptin inhibits CB1R expression in the brain and thus Ob rats would have higher levels of CB1R than their lean counterparts; an effect that would be accentuated in food deprivation conditions.

**MATERIALS AND METHODS**

**Animals**

One-month-old male Zucker Obese {fa/fa} (Ob; n = 20) and Lean {Fa/FA} (Le; n = 20) rats were used. Rats were divided into four groups, 10 rats in each group. Specifically: (i) Ob rats with unrestricted (U) food access, (ii) Ob rats with restricted (R) food access, (iii) Le U rats, and (iv) Le R rats. At 1 month of age half the rats were placed on restricted food access (70% of ad libitum fed animals) and the rest on free access to food (ad libitum). Rats were obtained from Harlan (Indianapolis, IN) and all experiments were conducted in conformity with the National Academy of Sciences Guide for the Care and Use of Laboratory Animals and Brookhaven National Laboratory Institutional Animal Care and Use Committee protocols.

**Behavior assessment—Food intake and weight**

Rats were fed a standard (Purina) laboratory rat chow. Food intake was monitored daily at 1500 h and all rats were weighed every other day. Restricted diet rats were fed daily at 1500 h and the amount of food given was continuously adjusted to 70% of food intake of ad libitum pair-fed rats.

**CB1R autoradiography**

Each animal was deeply anesthetized with a mixture of Xylazine (10 mg/kg) and Ketamine (100 mg/kg). The brain was rapidly removed and frozen in an isopentane and dry-ice bath and stored in a −80°C freezer. The brain was transferred to the cryostat and sections of 14 μm thick were cut at −22°C and then stored at −80°C until binding. Slides were then preincubated at room temperature for 10 min in 50 mM Tris-HCl buffer (pH 7.4) and were then incubated in binding buffer (50 mM Tris-HCl, 0.4 nM [3H] SR 141716A) (Amersham Biosciences, Piscataway, New Jersey) at room temperature for 90 min. To determine nonspecific binding some slides in parallel were similarly incubated in binding buffer in the presence of 100 μM HU-210 (Tocris Bioscience, Ellisville, Missouri). Next, the slides were washed 3 × 30 min in ice-cold 50 mM Tris-HCl and then rapidly rinsed in ice-cold D2O. Slides were then dried overnight at room temperature in a desiccator and placed in a glass slide cassette for image acquisition scanning and analysis using a β-Imager 2000 (Biospace Mesures, FR). Scans were acquisitioned in the β-Imager 2000 for 6 h at high resolution. Using β vision+ software (Biospace Mesures, FR, USA), regions of interest (ROIs) were drawn on the left and right: striatum (ST), nucleus accumbens (NAc), cerebellum (CB), cingulate (CC), frontal (FC), parietal (PC), insulate (IC) cortices, and hypothalamus (HYP) of each brain slice. These data were averaged and expressed initially in counts per minute per millimeter square (cpm/mm²) and were subsequently converted into microcuries per gram (μCi/g) of tissue by using a brain homogenate standard of known radioactivity value and mass (provided by Y. Piyis at Brookhaven National Laboratory) taking into account apparatus recovery (90%) and autoabsorption (40%) due to section thickness (Langlois et al., 2001; Poisnel et al., 2006). The μCi/g estimation is a relative measure of radioactivity/mass of tissue since the tissue section and homogenate differ in their respective tissue densities.

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RESULTS

Body weight

Body weight was examined in each group over a 4-month period. A three-way ANOVA revealed significant main effects in body weight with respect to strain ($F = 461.683; df = 1, 79; P < 0.001$), diet ($F = 320.027; df = 1, 79; P < 0.001$), and age ($F = 5288.733; df = 1, 79; P < 0.001$). At 1 month, there were no significant differences in weight between any of the groups (Ob U: $110.3 \pm 9.61$ g; Le U: $119.3 \pm 10.81$ g; Ob R: $107.4 \pm 11.47$ g; Le R: $114.7 \pm 8.38$ g). At 4 months, Ob U rats showed significantly higher weight gain than Le R ($t = 23.873; P < 0.05$), Le U ($t = 26.545; P < 0.05$), and Ob R ($t = 28.154; P < 0.05$). Similarly, at 4 months, Le U and Ob R showed significantly higher weight gain compared with Le R ($t = 7.441; P < 0.05$ and $t = 5.832; P < 0.05$) respectively (Ob U: $564.23 \pm 15.4$ g; Le U: $408.84 \pm 7.81$ g; Ob R: $364.14 \pm 7.04$ g; Le R: $270.55 \pm 4.22$ g). There were no weight differences between the Ob R and the Le U rats.

Food intake

Throughout the study food intake was monitored in the two unrestricted rat groups. A two-way ANOVA showed significant main effects in body weight with respect to strain ($F = 461.683; df = 1, 79; P < 0.001$) and age ($F = 5288.733; df = 1, 79; P < 0.001$). At 1 month, there were no differences in food intake between Ob and Le animals (Ob U: $19.7 \pm 0.56$ g; Le U: $19.46 \pm 1.16$ g). At 4 months Ob rats had higher food intake levels compared with Le ($t = 6.477; P < 0.05$) (Ob U: $33.85 \pm 0.82$ g; Le U: $23.83 \pm 0.46$ g). Specifically, Ob rats consumed on average an additional 10 g of food than Le U rats (30% more). Finally, food intake levels were significantly higher at 4 compared with 1 month for both Ob ($t = 13.722; P < 0.05$) and Le rats ($t = 9.604; P < 0.05$).

CB$_1$R autoradiography

The findings from this study were as follows: (1) at 1 month of age most brain regions did not differ between Ob and Le animals (except lower relative CB$_1$R binding levels in frontal and cingulate cortices in Le and in the hypothalamus in Ob); (2) CB$_1$R were significantly higher at 4 months than at 1 month of age in all animals (in all brain regions except the CB, and HYP); (3) Ob had significantly higher relative CB$_1$R binding levels than Le animals (4 months of age) (except in HYP); and (4) diet restriction was associated with higher relative CB$_1$R binding levels in all brain regions in Ob but not in Le animals (4 months of age) (except in the HYP). These findings are described later.

Brain sections were analyzed for [$^{3}$H] SR 141716A CB$_1$R binding at 1 and 4 months (Fig. 1). A one-way ANOVA showed no significant ($P > 0.05$) differences between the left and right for all eight brain regions of interest (ROI) examined: (ST, NAc, CB, CC, FC, PC, IC, and HYP). The left and right values from each ROI were combined and treated as a single ROI value for the purpose of maximizing statistical power. A two-way ANOVA on CB$_1$R levels revealed a significant group ($F = 55.72; df = 5.7289; P < 0.001$) and group
X brain region interaction effect ($F = 3.62$; $df = 35.7289$; $P < 0.05$); along with several significant pairwise multiple comparisons using the Holm-Sidak method (see later).

**Age effects**

At 4 months Ob R rats had significantly higher relative CB$_{1}$R binding levels than 1-month-old OB U in IC ($t = 11.41$, $P < 0.001$), NAc ($t = 9.88$, $P < 0.001$), PC ($t = 9.33$, $P < 0.001$), FC ($t = 8.49$, $P < 0.001$), CC ($t = 7.85$, $P < 0.001$), ST ($t = 7.42$, $P < 0.001$), but lower than 1-month-old Le U rats ($t = 2.48$, $P < 0.05$) in the HYP (Fig. 1). Similarly, 4-month-old Ob U rats also showed significantly higher relative CB$_{1}$R binding levels than 1-month-old Ob U rats in the ST ($t = 2.46$, $P < 0.05$), NAc ($t = 4.23$, $P < 0.001$), CC ($t = 3.33$, $P < 0.001$), FC ($t = 3.61$, $P < 0.001$), PC ($t = 4.23$, $P < 0.001$), and in the IC ($t = 5.09$, $P < 0.001$) (Fig. 1). Differences were also significant in 4-month-old Le R rats compared with 1-month-old Le U rats in the ST ($t = 2.21$, $P < 0.05$), NAc ($t = 3.70$, $P < 0.001$), CC ($t = 5.22$, $P < 0.001$), the FC ($t = 5.75$, $P < 0.001$), the PC ($t = 3.92$, $P < 0.001$), and the IC ($t = 5.34$, $P < 0.001$), which Le R rats showed higher CB$_{1}$R binding than Le U rats (Fig. 1). Le R 4-month-old rats also showed higher binding in the IC ($t = 3.34$, $P < 0.001$) and NAc ($t = 1.98$, $P < 0.05$) than 1-month-old Ob U rats (Fig. 1). At 4 months, Le U rats showed higher respective CB$_{1}$R binding than 1 month Le U rats in the ST ($t = 2.44$, $P < 0.05$), NAc ($t = 3.72$, $P < 0.001$), CC ($t = 5.00$, $P < 0.001$), PC (5.70, $P < 0.001$), and in the IC ($t = 5.26$, $P < 0.001$) (Fig. 1). Overall these results show an increase in CB$_{1}$R binding with respect to age, except in the CB and HYP.

**Diet and strain effects**

At 1 month, Ob U rats displayed significantly higher respective CB$_{1}$R binding levels than Le U rats in both the CC ($t = 4.93$, $P < 0.001$) and FC ($t = 5.30$, $P < 0.001$), but lower although not significant in the CB ($t = 0.24$; $P > 0.05$) and HYP ($t = 1.47$, $P > 0.05$) (Fig. 1). In contrast, at 4 months, Ob rats displayed significantly higher CB$_{1}$R binding in most brain regions than Le rats (except in the CB and HYP). The greatest CB$_{1}$R binding levels were observed in the Ob R rats. Specifically Ob R had higher CB$_{1}$R levels in ST than Ob U ($t = 4.66$, $P < 0.001$), Le U ($t = 6.24$, $P < 0.001$), and Le R ($t = 6.48$, $P < 0.001$); in NAc than Ob U ($t = 5.52$, $P < 0.001$), Le U ($t = 7.26$, $P < 0.001$), and Le R ($t = 7.64$, $P < 0.001$); in CC than Ob U ($t = 4.21$, $P < 0.001$), Le U ($t = 6.21$, $P < 0.001$), and Le R ($t = 6.58$, $P < 0.001$); in PC than Ob U ($t = 4.51$, $P < 0.001$), Le U ($t = 6.45$, $P < 0.001$), and Le R ($t = 6.89$, $P < 0.001$); in PC than Ob U ($t = 4.74$, $P < 0.001$), Le U ($t = 6.93$, $P < 0.001$), and Le R ($t = 7.57$, $P < 0.001$); in IC than Ob U ($t = 5.89$, $P < 0.001$), Le U ($t = 7.83$, $P < 0.001$), Le R ($t = 8.15$, $P < 0.001$), but lower in HYP than Le U ($t = 2.61$, $P < 0.05$) (Fig. 1).

Diet restriction significantly affected brain regional CB$_{1}$R binding levels in the Ob but did not affect the respective levels in the Le rats (4 months of age), except in the HYP. Specifically CB$_{1}$R binding levels were significantly higher in Ob R than Ob U in all regions; but not significantly higher ($t = 0.65$, $P > 0.05$) in the CB and lower in the HYP ($t = 0.01$, $P > 0.05$). In contrast at 4 months, respective CB$_{1}$R binding levels did not differ in any of the brain regions between the Le R and the Le U rats, except in the HYP.

**DISCUSSION**

Here we show significant differences in CB$_{1}$R binding expression in the brain between Ob and Le rats; specifically in frontal, limbic, and striatal regions, CB$_{1}$R binding levels were higher in Ob than Le rats. Since Ob rats are deficient in leptin receptor function this suggests that leptin inhibits CB$_{1}$R expression in regions that process rewarding and motivational signals associated with food intake (Kelley et al., 2005). When leptin levels are low and the animals are on a restricted diet, CB$_{1}$R expression increased in Ob but not in the Le rats. This suggests that leptin receptor activity may play a role in inhibiting CB$_{1}$R in the brain. Leptin’s regulation may be developmentally triggered since the differences between strains, both for CB$_{1}$R expression as well as food intake and weight, were observed at 4 months but not at 1 month of age. In addition, we also show that CB$_{1}$R levels increased with age; CB$_{1}$R were higher at 4 months than at 1 month of age in both strains.

**Strain effects on CB$_{1}$R: Leptin signaling**

Zucker obese rats have been widely used as a model of obesity and characterized by a mutation in the leptin receptor (Chua et al., 1996) leading to obesity, diabetes, and hyperphagia (Fetissov et al., 2000). In our study, the only observed differences in CB$_{1}$R between Ob and Le strains of Zucker rats at 1 month of age occurred in the CC, FC (Ob $>$ Le rats), and HYP (Le $>$ Ob). This suggests that the CB$_{1}$R in these specific brain structures was affected by leptin-receptor deficiency or behavioral and physiological manifestations associated with this deficiency. Leptin activation inhibits endocannabinoid release (Jo et al., 2005) in the HYP, which is neuroanatomically connected both with the FC and CC (Morecraft et al., 1992; Pajolla et al., 2001). Work done in ob/ob mice (leptin deficient) recently has shown a higher concentration of 2-AG (an endocannabinoid) in the HYP relative to

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wild-type mice (Di Marzo et al., 2001). Therefore, increased CB₁R binding levels in Ob rats at this early age in the CC and FC may reflect neuroadaptative changes associated with increased endocannabinoid release, which may be involved in increased food intake regulated by leptin (Di Marzo et al., 2001).

Compared with 1-month-old adolescent rats, adult 4-month-old rats’ CB₁R binding levels showed significant differences between Ob and Le rats in all brain regions examined apart from the CB. Indeed, it is possible that leptin does not regulate CB₁R in the CB. Defective leptin signaling is associated with elevated hypothalamic, but not cerebellar levels of endocannabinoids in Zucker rats (Di Marzo et al., 2001). On the other hand, the increased levels of CB₁R binding in brain regions involved in the rewarding effects of food (NAc, IC, CC, CPU, FC, and PC) suggests that CB₁R in these regions may contribute to hyperphagia observed in 4-month-old Ob rats (Durham and Truett, 2006; Kurtz et al., 1989). Whereas there is clear evidence that leptin modulates cannabinoid signaling in HYP, our findings suggest that the modulatory role of leptin on cannabinoid signaling extends to other brain regions.

**Developmental effects on CB₁R**

All rats showed significantly higher CB₁R binding at 4 months compared with 1 month of age in all regions except for the CB of Le rats and the HYP of Ob rats. This suggests that in the rodent there is an increase in CB₁R binding levels in most of the brain regions in the transition from childhood to adolescence. The increases in CB₁R with age are consistent with prior rodent studies showing gradual increases in CB₁R mRNA in the FC, hippocampus, basal ganglia, and CB between the fetal period and adulthood (Berrendero et al., 1999) and with human postmortem studies showing increases in CB₁R mRNA and CB₁R binding levels between prenatal and postnatal brain development (Mato et al., 2003; Wang et al., 2003). The increases in CB₁R we observed with age may contribute to the developmentally related increases in food intake and weight gain.

The developmental changes in CB₁R, which differed between the strains, paralleled the changes in weight gain and food intake. Ob and Le rats did not differ at 1-month but at 4-months of age; Ob rats weighed 39% more and consumed 42% more than their lean counterparts. This is consistent with the developmental regulation of leptin receptors in rat pituitary and HYP (Morash et al., 2003).

**Diet effects on CB₁R**

Food restriction significantly reduced body weight at 4 months of age in both Ob and Le rats; Ob R rats weighed 55% less than Ob U rats and Le R rats weighed 51% less than Le U rats. However food restriction increased CB₁R in Ob rats (deficient leptin receptors) but not in the Le rats (normal leptin receptors). This suggests that leptin receptors may exert an inhibitory role on CB₁R upregulation under conditions of food deprivation. Since leptin levels drop with food deprivation, this suggests that constitutive activity of leptin receptors may be involved in this inhibition. However, to our knowledge constitutive activity of leptin receptors has previously been documented only for peripheral tissues (adipose tissue, placenta, and stomach) (Morash et al., 1999). Thus, further work is required to assess if constitutive activity for leptin receptors occurs in the brain.

Modulation of CB₁R by diet is consistent with the recognized role of CB₁R in feeding behavior (Gomez et al., 2002; Williams and Kirkham, 2002) and blocking CB₁R reduces food intake in both Le and Ob Zucker rats, with a greater effect on Ob rats (Vickers et al., 2003). Moreover, CB₁R knockout mice are resistant to diet-induced obesity, show lower food intake, and show a diminished response to rewarding stimuli compared with controls (Sanchis-Segura et al., 2004). The CB₁R antagonist, Rimonabant (SR141716A), reduced body weight and decreased food intake (of obese Zucker rats (Bensaid et al., 2003; Vickers et al., 2003) and mice (Ravinet Trillou et al., 2003)); reduced ethanol intake (in C57BL/6 mice (Arnone et al., 1997; Thanos et al., 2005a); Sardinian alcohol-prefering (SP) rats (Colombo, 1997; Colombo et al., 2004); alcohol preferring (P) rats (Thanos et al., 2005b); and decreased place preference for alcohol (Houchi et al., 2005; Thanos et al., 2005a).

In summary, the high levels of CB₁R levels in limbic cortical and subcortical brain region of obese Zucker rats suggests that leptin’s regulation of food intake is in part mediated by CB₁R in areas of the brain involved with the rewarding and motivational aspects of food intake. The increases in CB₁R in Ob but not in Le animals with diet restriction when leptin levels are low, suggests that constitutive leptin receptor activity interferes with CB₁R upregulation.

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